National Comprehensive Cancer Network®

NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®)

Genetic/Familial High-Risk Assessment: Colorectal

Version 3.2019 — December 13, 2019

NCCN.org

Continue

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Genetic/Familial High-Risk Assessment: Colorectal

NCCN Guidelines Index
Table of Contents
Discussion

*Dawn Provenzale, MD, MS/Chair ¤ Þ
Duke Cancer Institute

*Samir Gupta, MD/Vice-chair ¤
UC San Diego Moores Cancer Center

Dennis J. Ahnen, MD ¤
University of Colorado Cancer Center

Lee-May Chen, MD ¶
UCSF Helen Diller Family
Comprehensive Cancer Center

Daniel C. Chung, MD ¤ Δ
Massachusetts General Hospital
Cancer Center

Gregory Cooper, MD ¤
Case Comprehensive Cancer Center/
University Hospitals Seidman Cancer
Center and Cleveland Clinic Taussig
Cancer Institute

Dayna S. Early, MD ¤
Siteman Cancer Center at Barnes-
Jewish Hospital and Washington
University School of Medicine

Francis M. Giardiello, MD, MBA ¤
The Sidney Kimmel Comprehensive
Cancer Center at Johns Hopkins

William Grady, MD ¤
Fred Hutchinson Cancer Research
Center/Seattle Cancer Care Alliance

Michael J. Hall, MD, MS † Δ
Fox Chase Cancer Center

Amy L. Halverson, MD ¶
Robert H. Lurie Comprehensive Cancer
Center of Northwestern University

Stanley R. Hamilton, MD ≠
The University of Texas
MD Anderson Cancer Center

Heather Hampel, MS, CGC Δ
The Ohio State University Comprehensive
Cancer Center - James Cancer Hospital
and Solove Research Institute

Sigurdis Haraldsdottir, MD, PhD †
Stanford Cancer Institute

Priyanka Kanth, MD, MS ¤
Huntsman Cancer Institute
at the University of Utah

Jason B. Klapman, MD ¤
Moffitt Cancer Center

Audrey J. Lazenby, MD ≠
Fred & Pamela Buffett Cancer Center

Xavier Llor, MD, PhD ¤ Þ
Yale Cancer Center/
Smilow Cancer Hospital

Patrick M. Lynch, MD, JD ¤
The University of Texas
MD Anderson Cancer Center

Arnold J. Markowitz, MD ¤
Memorial Sloan Kettering Cancer Center

Robert J. Mayer, MD † Þ
Dana-Farber/Brigham and Women's
Cancer Center

June Mikkelson, MS, CGC Δ
Roswell Park Cancer Institute

Reid M. Ness, MD, MPH ¤
Vanderbilt-Ingram Cancer Center

Shajan Peter, MD ¤
O'Neal Comprehensive
Cancer Center at UAB

Scott E. Regenbogen, MD ¶
University of Michigan
Rogel Cancer Center

Niloy Jewel Samadder, MD ¤
Mayo Clinic Cancer Center

Thomas P. Slavin Jr, MD Δ
City of Hope
National Medical Center

Jennifer M. Weiss, MD, MS ¤
University of Wisconsin
Carbone Cancer Center

NCCN
Ndiya Ogba, PhD
Mary Dwyer, MS

Δ Cancer genetics	¥ Patient advocacy
¤ Gastroenterology	¶ Surgery/Surgical oncology
Þ Internal medicine	* Discussion Writing Committee Member
† Medical oncology	
≠ Pathology	

NCCN Guidelines Panel Disclosures

Continue

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Genetic/Familial High-Risk Assessment: Colorectal

NCCN Genetic/Familial High-Risk Assessment: Colorectal Panel Members
Summary of the Guidelines Updates

High-Risk Colorectal Cancer Syndromes
- Assessment for Hereditary Colorectal Cancer Syndrome (HRS-1)
- Principles of Cancer Risk Assessment and Counseling (HRS-A)

Non-Polyposis Syndrome
- Lynch Syndrome (Hereditary Nonpolyposis Colorectal Cancer) (LS-1)
  - Principles of IHC and MSI Testing for Lynch Syndrome (LS-A)
  - Cancer Risks in Lynch Syndrome by Gene Compared to the General Population (LS-B)

Polyposis Syndromes
- Adenomatous Polyposis Testing Criteria (POLYP-1)
- Familial Adenomatous Polyposis/AFAP (FAP/AFAP-1)
  - Familial Adenomatous Polyposis (FAP-1)
    - ◊ Surgical Options for Treating the Colon and Rectum in Patients with FAP (FAP-A)
  - Attenuated Familial Adenomatous Polyposis (AFAP-1)
  - *MUTYH*-Associated Polyposis (MAP-1)
- Peutz-Jeghers Syndrome (PJS-1)
- Juvenile Polyposis Syndrome (JPS-1)
- Serrated Polyposis Syndrome (SPS-1)
- Colonic Adenomatous Polyposis of Unknown Etiology (CPUE-1)
- Multi-Gene Testing (GENE-1)

Clinical Trials: NCCN believes that the best management for any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

To find clinical trials online at NCCN Member Institutions, click here: nccn.org/clinical_trials/clinicians.aspx.

NCCN Categories of Evidence and Consensus: All recommendations are category 2A unless otherwise indicated.

See NCCN Categories of Evidence and Consensus.

The NCCN Guidelines® are a statement of evidence and consensus of the authors regarding their views of currently accepted approaches to treatment. Any clinician seeking to apply or consult the NCCN Guidelines is expected to use independent medical judgment in the context of individual clinical circumstances to determine any patient's care or treatment. The National Comprehensive Cancer Network® (NCCN®) makes no representations or warranties of any kind regarding their content, use or application and disclaims any responsibility for their application or use in any way. The NCCN Guidelines are copyrighted by National Comprehensive Cancer Network®. All rights reserved. The NCCN Guidelines and the illustrations herein may not be reproduced in any form without the express written permission of NCCN. ©2019.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Genetic/Familial High-Risk Assessment: Colorectal

NCCN Guidelines Index
Table of Contents
Discussion

Updates in Version 3.2019 of the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal from Version 2.2019 include:

MS-1 The Discussion section was updated to reflect the changes in the algorithm.

Updates in Version 2.2019 of the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal from Version 1.2019 include:

LS-B 1 of 2

- Cancer Risks in LS by Gene Compared to the General Population table, the breast cancer risk for *MLH1* was changed from "12%–25%" to "12%-17%."

Updates in Version 1.2019 of the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal from Version 1.2018 include:

High-Risk Colorectal Cancer Syndromes

HRS-1

- Assessment for hereditary CRC syndrome
  - First question was revised, "Is there a personal history *or family history* of a known *pathogenic variant in a colorectal polyposis or cancer gene* ~~genetic mutation or known genetic mutation in the family~~?
  - If No, the criteria was revised, "Personal *or family* history of:" and "Family history of: >1 relative with polyposis" was removed.
- Footnote b was added, "Pathogenic variant includes likely pathogenic variant. Slavin TP, Van Tongeren LR, Behrendt CE, et al. Prospective study of cancer genetic variants: Variation in rate of reclassification by ancestry. J Natl Cancer Inst 2018;110:1059-1066."
- Footnote c was added, "Irrespective of degree of relatedness." (Also for LS-1 and POLYP-1)

HRS-2

- Qualifier "≥5 serrated polyps" was revised by adding "proximal to sigmoid colon."
- After qualifier >10 adenomas, "Rare genetic causes of multiple adenomatous polyps" was added with corresponding footnote i.

HRS-3

- The "Criteria for the Evaluation of Lynch Syndrome" were reorganized by personal history, family history and increased model-predicted risk for Lynch syndrome.
- Personal history, 3rd sub-bullet was revised by removing, "diagnosed ≤60 y"
- Increased model-predicted risk for Lynch syndrome, sub-bullet was revised, "An individual with ~~a LS-related cancer or unaffected individual with~~ a ≥5% risk..."

HRS-A

- The first three pages of the Principles of Cancer Risk Assessment and Counseling are a new section in the guidelines.

HRS-A 4 of 6

- For family history of cancer and expanded pedigree, 2nd bullet was changed from "Minimal data set on each affected relative" to "Recommended data on each affected relative" and two additional sub-bullet were added, "Birth resulting from sperm or egg donor" and "History of allogeneic (related or unrelated donor) bone marrow transplant."

Lynch Syndrome

LS-1

- The algorithm and footnotes on this page were extensively revised.

LS-2

- Lynch Syndrome Management
  - Other Extracolonic Cancers
    - ◊ Gastric and small bowel cancer recommendation was revised, "...Also, individuals of Asian descent *(or from countries with high background incidence of gastric cancer)* may have increased risk for stomach cancer and may benefit from surveillance."
    - ◊ Urothelial cancer recommendation was revised, "*There is no clear evidence to support surveillance for urothelial cancers in LS. Surveillance may be considered in* selected individuals such as with a family history of urothelial cancer or individuals with MSH2 pathogenic variants (especially males) *as these groups appear to be at higher risk* ~~may want to consider screening surveillance~~."
  - Footnotes
    - ◊ Footnote o was added, "Patients who may benefit from a shorter 1- versus longer 2-year interval include those with risk factors such as history of CRC, male sex, MLH1/MSH2 pathogenic variant, age >40 years, and history of adenoma. See Discussion." (Also for LS-5)
    - ◊ Footnote q was revised from, "Patients with LS can consider ongoing clinical trials for pancreatic cancer screening" to "If screening is performed, it should be considered in high-volume centers with multidisciplinary teams, preferably within research protocols. The International Cancer of the Pancreas Screening (CAPS) consortium recommends that patients with LS with one first-degree relative with pancreatic cancer should be considered for screening."

Continued

UPDATES

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Genetic/Familial High-Risk Assessment: Colorectal

NCCN Guidelines Index
Table of Contents
Discussion

Updates in Version 1.2019 of the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal from Version 1.2018 include:

- 1st finding, No pathologic findings, follow-up was revised by adding, "Continued surveillance *every 1–2 y*"
- 3rd finding, Adenomas,
  - 1st bullet was revised by removing, "Complete endoscopic polypectomy with follow-up colonoscopy every 1–2 y ~~depending on: Location, character, Surgical risk, Patient preference for colonoscopy frequency~~."
  - 4th finding was revised, "Adenomas not amenable to endoscopic resection ~~or high-grade dysplasia~~."
  - 5th finding was added, "Adenomas with high-grade, dysplasia."
- Footnote t was added, "Surgery is recommended if resection was not en bloc or if dysplasia involved the resection margin, whereas surveillance may be considered if an en bloc complete excision was performed."

LS-A 1 of 5

- Bullet was added to page, "The panel recommends universal screening of all CRCs to maximize sensitivity for identifying individuals with Lynch syndrome and to simplify care processes. Counseling by an individual with expertise in genetics is not required prior to routine tumor testing. An infrastructure needs to be in place to handle the screening results."
- General
  - 1st bullet, 1st sentence was revised by removing, "(an inherited mutation of one of the MMR genes or EPCAM)."
  - 2nd bullet was revised, "*The panel recommends a universal screening strategy be the primary approach to identify CRC patients with LS. However, in other lower resource settings, other historic criteria for selecting patients for testing may be relevant.* The Bethesda criteria (~~See LS-1~~ See Discussion) are intended to help identify CRC patients whose tumors should be tested for MMR defects, by MSI and/or IHC analysis, thereby identifying patients with a greater chance of having LS. ~~Although more sensitive than the Amsterdam criteria, up to 50% of patients with LS do not meet even the revised Bethesda Guidelines~~."
- IHC, 2nd bullet was revised by adding, "If abnormal IHC is followed by germline testing and no LS-causing pathogenic variants are identified, the panel strongly recommends proceeding with MLH1 methylation analysis of the tumor. Patients who have normal germline testing and MLH1 hypermethylation are likely to have sporadic cancer and should be treated as such taking into account their family history."

LS-A 3 of 5

- Pros and Cons of Universal Tumor Screening *with IHC and/or MSI* for LS Using Colonoscopy-Based Biopsy Versus Surgical Resection Specimen
  - Surgical testing considerations, Pros, a bullet was removed, "Patient may be less likely to be lost to follow-up"

LS-A 5 of 5

- Footnote j was added, "These tumor testing results may also have implications for treatment in cases that are sporadic or hereditary. See The NCCN Guidelines for Colon Cancer Guidelines for more information on pathologic review and the impact on management (COL-B 4 of 6). Consult with an expert if the scenario is not covered by this table."
- Footnote d was revised, "... it is recommended that these patients and their close relatives be managed based on their family history and NOT as if they have LS. ~~Regardless of the results of tumor sequencing, these patients and their close relatives should be managed based on their family history and NOT as if they have LS unless their family history warrants it~~. If double somatic pathogenic variants are identified..."

LS-B 1 of 2

- The table, "Cancer Risks in Lynch Syndrome by Gene Compared to the General Population" was extensively revised.

Adenomatous Polyposis Testing Criteria

POLYP-1

- The name of the page changed from "APC and MUTYH Genetic Testing Criteria" to "Adenomatous Polyposis Testing Criteria."
- For specific APC and MUTYH testing criteria was removed and replaced with adenomatous polyposis testing criteria.
- The remainder of the page was revised according to "Pathogenic variant(s) known" and "No known pathogenic variants in any polyposis gene."
- Footnotes
  - Footnote b was added, "Age of onset, family history, personal history of colorectal cancer, and/or presence of other features may influence whether genetic testing is offered in these situations.
  - Footnote c was added, "There are clinically relevant yet rarer genes that can cause a polyposis that may be phenotypically indistinguishable from APC/MUTYH polyposis."
  - Footnote d was added, "Multi-gene panel should include all polyposis and colorectal cancer genes (Stanich P, et al. Clin Gastroenterol Hepatol 2018)."

Continued

UPDATES

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Updates in Version 1.2019 of the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal from Version 1.2018 include:

- Footnote f was revised, "...Full sequencing of MUTYH may be considered in an unaffected parent when the other parent has MAP. ~~If the unaffected parent is found to not have a MUTYH pathogenic variant, genetic testing in the children is not necessary to determine MAP status.~~ If the unaffected parent is found to have one MUTYH pathogenic variant, testing the children for the familial MUTYH pathogenic variants is indicated. If the unaffected parent is not tested, comprehensive testing of MUTYH should be considered in the children. *Testing for children of MUTYH heterozygotes should be offered if the other parent is also a heterozygote or could still be offered if the other parent is not a heterozygote and management would change (if they have an FDR affected with CRC) or inform reproductive risks (since their future children could be at-risk for MAP*)."

Familial Adenomatous Polyposis

FAP-1

- Footnote was removed, "A single pilot study among patients with FAP suggests the omega-3 polyunsaturated fatty acid eicosapentaenoic acid has potential to reduce size and number of polyps on follow-up (West NJ, Clark SK, Phillips RK, et al. Gut 2010;59:918-925). However, evidence is insufficient to recommend routine use, and a meta-analysis of N-3 polyunsaturated fatty acids intake and risk of CRC (not limited to FAP patients) did not show a clear protective association."

FAP-2

- Gastric cancer bullet was revised, "~~Examine stomach at time of upper endoscopy~~. Fundic gland polyps occur in a majority of FAP patients, and focal low-grade dysplasia can occur but is typically non-progressive. *The risk of gastric cancer in FAP patients appears to be increased in patients from geographic areas with high gastric cancer risk and may be elevated in the setting of certain endoscopic findings, including carpeting of fundic gland polyps, solitary polyps larger than 20 mm, and mounds of polyps. High-risk histologic features include tubular adenomas, polyps with high-grade dysplasia, and pyloric gland adenomas. Need for specialized surveillance or surgery should be considered in presence of high risk histologic features, preferably at a center of expertise*."
  - Sub-bullet was revised, "~~Adenomatous polyps of the stomach should be managed endoscopically if possible.~~ Patients with *high-risk lesions* ~~adenomatous polyps~~ that cannot be removed endoscopically, ~~or adenomas with high-grade dysplasia/or invasive cancer detected on biopsy~~ should be referred to a specialized center for consideration of gastrectomy."
- Footnote h was added, "Cap-assisted endoscopy may be adequate for visualization of the ampulla. (Kallenberg F, et al. Endoscopy 2017;49:181-185.)."

FAP-3

- Footnote j was added, "Potentially higher risk adenomas involving the papilla (ie, large ≥1 cm adenomas or adenomas extending into papilla) should be referred to an expert center for evaluation and management."

FAP-4

- Footnote k was revised by removing, "An at-risk family member can be defined as a first-degree relative of an affected individual and/or proband." (Also for footnote f on AFAP-2)
- Footnote l was added, "FAP genetic testing in children should be done by age 10 y when colon screening would be initiated. ~~In select cases where~~ *If there is intent to do* hepatoblastoma screening ~~will be pursued~~, FAP genetic testing ~~could be done~~ *should be considered* in infancy."

Attenuated Familial Adenomatous Polyposis

AFAP-2

- APC positive, surveillance was revised, "Colonoscopy beginning in late teens, then every *1–2* ~~2–3~~ y."

MUTYH-Associated Polyposis

MAP-2

- Surveillance, Extracolonic, 2nd bullet was revised, "Baseline upper endoscopy (*including complete visualization of the ampulla of Vater* beginning at age 30–35 y *[See FAP-3 for follow-up of duodenoscopic findings]*)" and corresponding footnote was added, "Cap-assisted endoscopy may be adequate for visualization of the ampulla. (Kallenberg F, et al. Endoscopy 2017;49:181-185.)." (Also for CPUE-1)

MAP-3

- Biallelic MUTYH pathogenic variant positive, surveillance was revised, "Begin colonoscopy at age 25–30 y and every *1–2* ~~2–3~~ y..."

Continued

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Genetic/Familial High-Risk Assessment: Colorectal

NCCN Guidelines Index
Table of Contents
Discussion

Updates in Version 1.2019 of the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal from Version 1.2018 include:

Peutz-Jeghers Syndrome

PJS-1

- Diagnosis, 1st bullet was added here and removed from surveillance, "The majority of cases occur due to pathogenic variants in the STK11 (LKB1) gene. Clinical genetic testing is available."

PJS-2

- Site, breast was clarified as being for women.
- Lifetime risk for breast was changed from 45%-50% to 32%-54%.
- Lifetime risk for lung was changed from 15%-17% to 7%-17%.
- Screening Procedure and Interval
  - Small intestine was revised, "... though this may be individualized, ~~or with symptoms~~. *Repeat small intestinal exam is also indicated at any time based on symptoms*.)."

Juvenile Polyposis Syndrome

JPS-1

- JPS definition, 1st bullet, 1st sub-bullet was revised, "~~At least 3 to~~ ≥5 juvenile polyps of the colon."

Serrated Polyposis Syndrome

SPS-1

- Definition, 3rd bullet was revised, "~~Currently~~, *For the majority of patients with SPS*, no causative gene ~~has been identified for serrated polyposis~~ is identifiable. *Pathogenic variants in RNF43 have been identified as a rare cause of serrated polyposis*."
- Surveillance, 1st bullet was revised, "The risk of CRC in *first-degree* relatives of individuals with serrated polyposis is ~~still unclear~~ *elevated*. ~~Pending further data it is reasonable to screen first-degree relatives at the youngest age of onset of serrated polyposis diagnosis, and subsequently per colonoscopic findings~~."

Colonic Adenomatous Polyposis of Unknown Etiology

CPUE-1

- Phenotype
  - 4th row is new, "Personal history of 11–20 adenomas."
  - 5th row was revised. "Family history of ≥100 adenomas in a first-degree relative *at age <40 y.*"
- Management/Surveillance, for both 2nd and 3rd row, a bullet was added, "Consider baseline upper endoscopy (including complete visualization of the ampulla of Vater) at baseline and repeat following duodenal surveillance guidelines on page FAP-3."
- Footnote b was revised, "~~Consider~~ *Recommend* genetic testing (See POLYP-1) in family member affected with polyposis."

Multi-Gene Testing

GENE-1

- 5th bullet, sub-bullet was added, "Reclassification of variants of uncertain significance is commonplace. Historically, over 91% of variants of uncertain significance in hereditary cancer testing have been downgraded to benign or likely benign categories. Nonetheless, clinical phenotypic correlation is warranted with further discussion with the testing laboratory if there is evidence supporting variant pathogenicity. Patient and provider guidelines and policies for follow-up of variants of uncertain significance have been developed."

GENE-4

- Note * was added, "Risk level is based on panel consensus." Also for GENE-5 and GENE-6.

GENE-7

- Table 5,
  - For MUTYH heterozygotes, 3rd bullet was revised, "~~Data are uncertain if specialized screening is warranted~~. Data are unclear as to whether specialized screening is warranted for MUTYH monoallelic carriers unaffected by CRC with no family history of CRC."

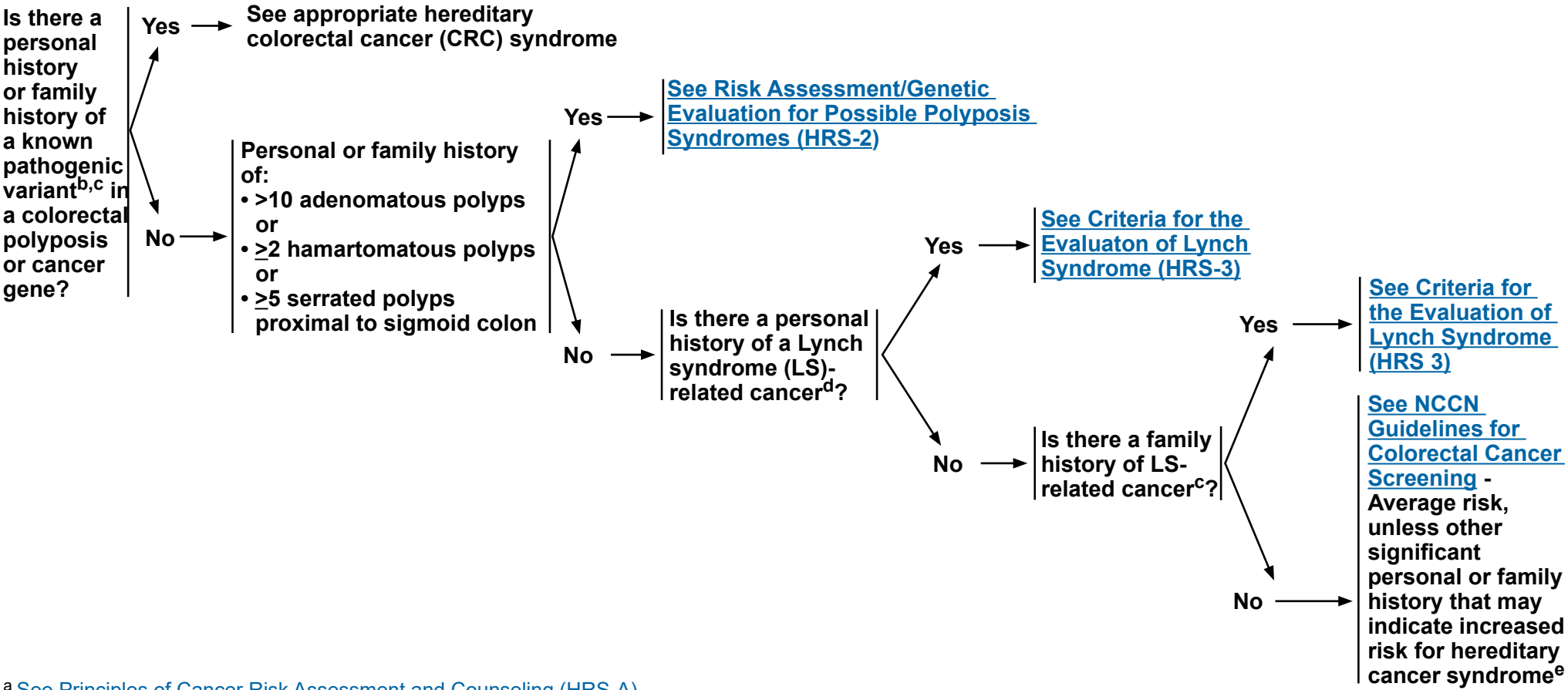
GENE-8

- Footnotes i and j was added.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

ASSESSMENT FOR HEREDITARY CRC SYNDROME[a]



[a] See Principles of Cancer Risk Assessment and Counseling (HRS-A).

[b] Pathogenic variant includes likely pathogenic variant. Slavin TP, Van Tongeren LR, Behrendt CE, et al. Prospective study of cancer genetic variants: Variation in rate of reclassification by ancestry. J Natl Cancer Inst 2018;110:1059-1066.

[c] Irrespective of degree of relatedness.

[d] LS-related cancers include colorectal, endometrial, gastric, ovarian, pancreas, ureter and renal pelvis, brain (usually glioblastoma), biliary tract, small intestinal cancers, as well as sebaceous adenomas, sebaceous carcinomas, and keratoacanthomas as seen in Muir-Torre syndrome.

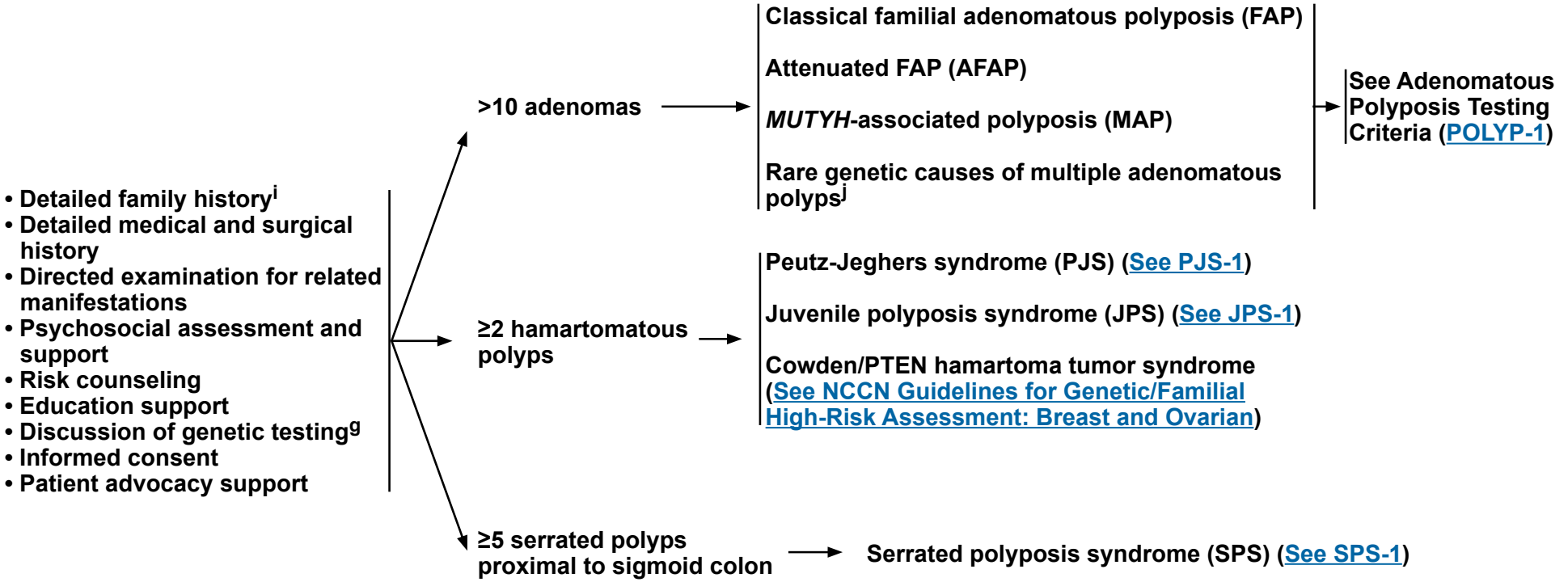
[e] Increased risk warranting genetic evaluation may be indicated by, but not restricted to personal or family history of congenital hypertrophy of the retinal pigment epithelium, osteomas, supernumerary teeth, desmoid tumor, cribriform variant of papillary thyroid cancer, brain cancer (usually medulloblastoma), and hepatoblastoma.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

RISK ASSESSMENT/GENETIC EVALUATION FOR POSSIBLE POLYPOSIS SYNDROMES[f,g,h]



[f] See Obtaining a Comprehensive Assessment for Hereditary Colorectal Cancer (HRS-A 4 of 6).

[g] Genetic counseling/patient education is highly recommended when genetic testing is offered and after results are disclosed. A genetic counselor, medical geneticist, oncologist, gastroenterologist, surgeon, oncology nurse, or other health professional with expertise and experience in cancer genetics should be involved early in counseling patients who potentially meet criteria for an inherited syndrome.

[h] If personal history of CRC and more than one syndrome might explain the presentation, consider multi-gene testing.

[i] If evaluation is based on family history of ≥1 relative with polyposis, then type of polyps in the affected relative (if known) may guide testing.

[j] Gene mutations associated with adenomatous polyposis include, but are not limited to monoallelic mutations in *GREM1*, *POLE*, *POLD1*, *AXIN2*, and biallelic mutations in *NTHL*1 and *MSH3*.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

CRITERIA FOR THE EVALUATION OF LYNCH SYNDROME

- **Known LS pathogenic variant in the family**

- **Personal history of colorectal, endometrial, or other Lynch syndrome-associated cancer**
  - **An individual with colorectal or endometrial cancer at any age with tumor showing evidence of mismatch repair (MMR) deficiency, either by microsatellite instability (MSI) or loss of MMR protein expression[k]**
  - **An individual with colorectal or endometrial cancer and any of the following:**
    - ◊ **Diagnosed <50 y**
    - ◊ **Another synchronous or metachronous LS-related cancer[d]**
    - ◊ **≥1 first-degree or second-degree relative with LS-related cancer[d] diagnosed <50 y**
    - ◊ **≥2 first-degree or second-degree relatives with LS-related cancers[d] regardless of age**
  - **An individual with a colorectal tumor with MSI-high (MSI-H) histology (ie, presence of tumor-infiltrating lymphocytes, Crohn's-like lymphocytic reaction, mucinous/signet ring differentiation, or medullary growth pattern)**

- **Family history of any of the following:**
  - **≥1 first-degree relative with colorectal or endometrial cancer diagnosed <50 y**
  - **≥1 first-degree relative with colorectal or endometrial cancer and another synchronous or metachronous LS-related cancer[d]**
  - **≥2 first-degree or second-degree relatives with LS-related cancer,[d] including ≥1 diagnosed <50 y**
  - **≥3 first-degree or second-degree relatives with LS-related cancers,[d] regardless of age**

- **Increased model-predicted risk for Lynch syndrome**
  - **An individual with a ≥5% risk[l] of having an MMR gene pathogenic variant based on predictive models (ie, PREMM5, MMRpro, MMRpredict)**

→ See Strategies For Evaluating LS (LS-1)

[d] LS-related cancers include colorectal, endometrial, gastric, ovarian, pancreas, ureter and renal pelvis, brain (usually glioblastoma), biliary tract, small intestinal cancers, as well as sebaceous adenomas, sebaceous carcinomas, and keratoacanthomas as seen in Muir-Torre syndrome.

[k] The panel recommends tumor screening for MMR deficiency for all colorectal and endometrial cancers regardless of age at diagnosis; however, germline genetic testing is generally reserved for patients with early age at diagnosis; positive family history; or abnormal tumor testing results: MSI or loss of MMR protein expression. See LS-A for details on tumor screening for Lynch syndrome.

[l] There are recent data that resulted in a lower threshold of ≥2.5% for the PREMM5 predictive model risk for having an MMR gene pathogenic variant. Based on these data, it is reasonable for testing to be done based on the ≥2.5% score result and clinical judgment. Of note, with the lower threshold, there is an increase in sensitivity, but a decrease in specificity. It is not known how this applies to the general population of unaffected individuals.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

NCCN Guidelines Version 3.2019
High-Risk Colorectal Cancer Syndromes

NCCN Guidelines Index
Table of Contents
Discussion

PRINCIPLES OF CANCER RISK ASSESSMENT AND COUNSELING

- **Cancer risk assessment and genetic counseling is highly recommended when genetic testing is offered (ie, pre-test counseling) and after results are disclosed (ie, post-test counseling).[1-5] A genetic counselor, medical geneticist, oncologist, gastroenterologist, surgeon, oncology nurse, or other health professional with expertise and experience in cancer genetics should be involved early in the counseling of patients.**

• Pre-test counseling includes: ▸ Collection of a comprehensive family history ◊ Note that when assessing family history, close blood relatives include first-, second-, and third-degree relatives on each side of the family (See HRS-A 6 of 6) ▸ Evaluation of a patient's cancer risk ▸ Generating a differential diagnosis and educating the patient on inheritance patterns, penetrance, variable expressivity, and the possibility of genetic heterogeneity ▸ Preparing the patient for possible outcomes of testing including positive (pathogenic, likely pathogenic), negative, and uncertain findings and obtaining informed consent	• Post-test counseling includes discussions of: ▸ Results along with their significance and impact and recommended medical management options ▸ Interpretation of results in context of personal and family history of cancer ▸ Informing and testing at-risk family members ▸ Available resources such as disease-specific support groups and research studies

Genetic Testing Considerations

- **Testing should be considered in appropriate high-risk individuals where it will impact the medical management of the tested individuals and/or their at-risk family members. It should be performed in a setting in which it can be adequately interpreted.[1]**
- **The probability of pathogenic/likely pathogenic variant detection associated with these criteria will vary based on family structure. Individuals with unknown or limited family history/structure may have an underestimated probability of familial pathogenic/likely pathogenic variant detection.**
- **Patients who have received an allogeneic bone marrow transplant should not have molecular genetic testing via blood or saliva samples due to unreliable test results from contamination by donor DNA. If available, DNA should be extracted from a fibroblast culture. If this source of DNA is not available, buccal brushing or skin biopsies can be considered.**
- **Comprehensive genetic testing includes full sequencing and testing for large genomic rearrangements. It is encouraged that testing be done in commercial or academic labs that are clinically approved and validated. See HRS 3 of 6.**
- **In children <18 y, genetic testing is generally not recommended when results would not impact medical management.[6]**
- **Likely pathogenic variants are often treated similarly to pathogenic variants.**

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF CANCER RISK ASSESSMENT AND COUNSELING

Genetic Testing Approach

- **If more than one family member is affected with cancers highly associated with a particular inherited cancer susceptibility syndrome, consider testing first a family member with youngest age at diagnosis, multiple primary cancers, or other cancers associated with the syndrome, or most closely related to the proband/patient. If there are no living family members with cancer that is a cardinal feature of the syndrome in question, consider testing first- or second-degree family members affected with other cancers thought to be related to the gene in question (eg, colorectal, endometrial or urothelial with LS pathogenic variants).**
- **Testing for unaffected family members when no affected member is available should be considered. Significant limitations of interpreting test results should be discussed.**
- **If no pathogenic/likely pathogenic variant is found, consider referral for expert genetics evaluation if not yet performed; testing for other hereditary cancer syndromes may be appropriate.**
- **Testing family members for a variant of unknown significance should not be used for clinical purposes. Consider a referral to research studies that aim to define the functional impact of variants such as variant reclassification programs through clinical labs or registries.**

Risk to Relatives

- **Advise about possible inherited cancer risk to relatives, options for risk assessment, and management.**
- **Recommend genetic counseling and consideration of genetic testing for at-risk relatives.**

Reproductive Options

- **For patients of reproductive age, advise about options for prenatal diagnosis and assisted reproduction, including pre-implantation genetic diagnosis. Discussion should include known risks, limitations, and benefits of these technologies.**
- **Biallelic pathogenic/likely pathogenic variants in some genes, such as *MUTYH*, and certain other genes included on gene panels, may be associated with autosomal recessive conditions. Thus, for these types of genes, consideration would be given to carrier testing the partner for pathogenic/likely pathogenic variants in the same gene if it would inform reproductive decision-making and/or risk assessment and management.[7]**

[1]Robson ME, Bradbury AR, Arun B, et al. American Society of Clinical Oncology Policy Statement Update: Genetic and Genomic Testing for Cancer Susceptibility. J Clin Oncol 2015;33:3660-3667.
[2]Berliner JL, Fay AM, Cummings SA, Burnett B, Tillmanns T. NSGC practice guideline: risk assessment and genetic counseling for hereditary breast and ovarian cancer. J Genet Couns 2013;22:155-163.
[3]American College of Obstetricians and Gynecologists; ACOG Committee on Practice Bulletins--Gynecology; ACOG Committee on Genetics; Society of Gynecologic Oncologists. ACOG Practice Bulletin No. 103: Hereditary breast and ovarian cancer syndrome. Obstet Gynecol 2009;113:957-966.
[4]Lancaster JM, Powell CB, Chen LM, Richardson DL; SGO Clinical Practice Committee. Society of Gynecologic Oncology statement on risk assessment for inherited gynecologic cancer predispositions. Gynecol Oncol 2015;136:3-7.
[5]Weitzel JN, Blazer KR, Macdonald DJ, Culver JO, Offit K. Genetics, genomics, and cancer risk assessment: State of the art and future directions in the era of personalized medicine. CA Cancer J Clin 2011;61:327-359.
[6]Committee on Bioethics; Committee on Genetics, and American College of Medical Genetics and; Genomic Social; Ethical; Legal Issues Committee. Ethical and policy issues in genetic testing and screening of children. Pediatrics 2013;131:620-622.
[7]Offit K, Levran O, Mullaney B, et al. Shared genetic susceptibility to breast cancer, brain tumors, and Fanconi anemia. J Natl Cancer Inst 2003;95:1548-1551.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF CANCER RISK ASSESSMENT AND COUNSELING

Evaluating the Source of Genetic Testing Information

- **Prior to using any germline findings for medical management, it is important to establish whether the reported findings were obtained from a laboratory that is certified by both the College of American Pathologists (CAP) and Clinical Laboratory Improvement Amendments (CLIA) to issue a report of germline findings directly to ordering health care providers. Some states (eg, New York) may have additional reporting requirements. Confirmatory germline testing through an appropriately certified laboratory is recommended when a potential pathogenic/likely pathogenic variant is identified through various data sources as noted below:**

- **Information obtained from direct-to-consumer ancestry/health-based services:**
  - **Commercial entities providing ancestry (and sometimes health) information typically do so through microarray-based single nucleotide polymorphism (SNP) testing that has not been validated for clinical use. Third-party software applications can be used by consumers to obtain an interpretation of the raw data provided by these companies. Raw data and third-party software are not able to provide information that is appropriate for medical management, as these services are not subject to quality-control processes and recent research suggests that the error rate is substantial.[8]**

- **Information obtained from tumor-only profiling (ie, without paired germline analysis):**
  - **Pathogenic/likely pathogenic variants reported by laboratories providing tumor-only profiling may be of somatic or germline origin. Although germline origin can sometimes be inferred with a high degree of confidence, confirmatory germline testing is indicated for pathogenic/likely pathogenic variants with a reasonable clinical suspicion of being of germline origin (based on patient/family history or clinical characteristics [and in some cases pathogenic/likely pathogenic variant frequency]). Somatic pathogenic/likely pathogenic variants in several genes with germline implications are common (eg, *TP53*, *STK11*, *PTEN*), and will rarely be indicative of a need for germline testing unless clinical/family history features suggest the possibility of a germline pathogenic/likely pathogenic variant.**
  - **It should be noted that the absence of reported pathogenic/likely pathogenic variants in a particular gene does not rule out the possibility of a germline pathogenic/likely pathogenic variant in that gene. Clinically indicated germline testing is still appropriate for patients meeting testing guidelines regardless of tumor profiling results.**

- **Other data sources:**
  - **Patients may have participated in research studies that include germline genomic analysis, or had some type of genomic testing because of a suspected genetic condition in their self or a relative. Incidental germline findings relating to cancer risk may have been reported.[9] In such cases, it is recommended to review the findings with a genetics professional and/or the reporting laboratory to establish whether the original report was generated by an appropriately certified laboratory, or whether confirmatory testing is recommended.**

[8]Tandy-Connor S, Guiltinan J, Krempely K, et al. False-positive results released by direct-to-consumer genetic tests highlight the importance of clinical confirmation testing for appropriate patient care. Genet Med 2018;20:1515-1521.
[9]Green R, Berg J, Grody W, et al. ACMG recommendations for reporting of incidental findings in clinical exome and genome sequencing. Genet Med 2013;15:565-574.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF CANCER RISK ASSESSMENT AND COUNSELING

Obtaining a comprehensive assessment for hereditary colorectal cancer[10]

Family history of cancer and expanded pedigree

- **It is essential to obtain a detailed family history, including:**
  - **Parents**
  - **Children**
  - **Siblings/half-siblings**
  - **Aunts and uncles**
  - **Grandparents**
  - **Great-grandparents**
  - **Cousins**
  - **Nieces and nephews**

  See Common Pedigree Symbols (HRS-A 5 of 6) and Pedigree: First-, Second-, and Third-Degree Relatives of Proband (HRS-A 6 of 6)
- **Recommended data on each affected relative:**
  - **Current age and age at diagnosis of cancer (medical record documentation of cancer is strongly encouraged)**
  - **Age and cause of death**
  - **Type of cancer (note multiple primaries)**
  - **Ethnicity/country of origin**
  - **Consanguinity**
  - **Birth resulting from sperm or egg donor**
  - **Suspected colon cancer syndromes and additional syndrome-specific features (eg, Muir-Torre syndrome, Turcot syndrome, PJS, JPS)[11]**
  - **All other inherited conditions and birth defects**
  - **History of allogeneic (related or unrelated donor) bone marrow transplant**

Detailed medical and surgical history

- **Pathology verification strongly encouraged**
- **Polyps**
- **Inflammatory bowel disease**
- **Inherited syndromes:**
  - **LS**
    - ◊ **Muir-Torre syndrome**
    - ◊ **Turcot syndrome**
  - **FAP and associated syndromes**
    - ◊ **AFAP**
    - ◊ **Gardner syndrome**
    - ◊ **Turcot syndrome**
  - **MAP**
  - **PJS**
  - **JPS**
  - **PTEN hamartoma tumor syndromes**
    - ◊ **Cowden syndrome**
    - ◊ **Bannayan-Riley-Ruvalcaba syndrome**

Directed examination for related manifestations

- **Colonoscopy**
- **Esophagogastroduodenoscopy (EGD)**
- **Indicated only if suspicion of a specific syndrome**
  - **Eye examination**
  - **Skin, soft tissue, and bone examination**
  - **Oral examination**
  - **Measurement of head circumference (≥97%, 58 cm in adult women, 60 cm in adult men)**

[10]Providers should be aware that multiple factors may limit the benefits of family history in helping to determine a patient's degree of cancer risk, including: small family size; unknown family history, eg, adoption or non-paternity; the potential for a new pathogenic variant arising in the patient (de novo pathogenic variant); variable penetrance of a pathogenic variant; autosomal recessive inheritance of risk; and mosaicism.

[11]Burt R and Neklason DW. Genetic testing for inherited colon cancer. Gastroenterology 2005;128:1696-1716.

Continued

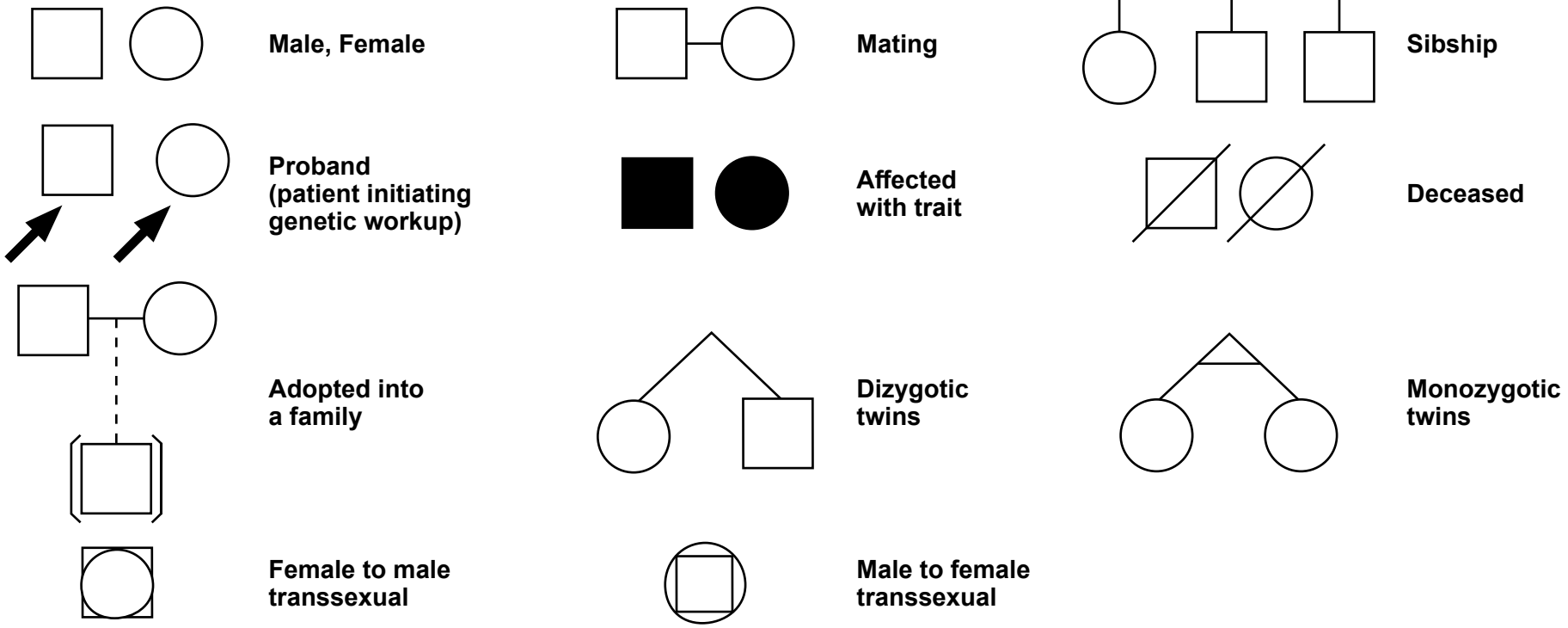
Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF CANCER RISK ASSESSMENT AND COUNSELING

COMMON PEDIGREE SYMBOLS[12]



See Pedigree: First-, Second-, and Third-Degree Relatives of Proband (HRS-A 6 of 6)

Continued

[12]Bennett RL, Steinhaus KA, Uhrich SB, et al. Recommendations for standardized human pedigree nomenclature. Pedigree Standardization Task Force of the National Society of Genetic Counselors. Am J Hum Genet 1995;56:745-752.

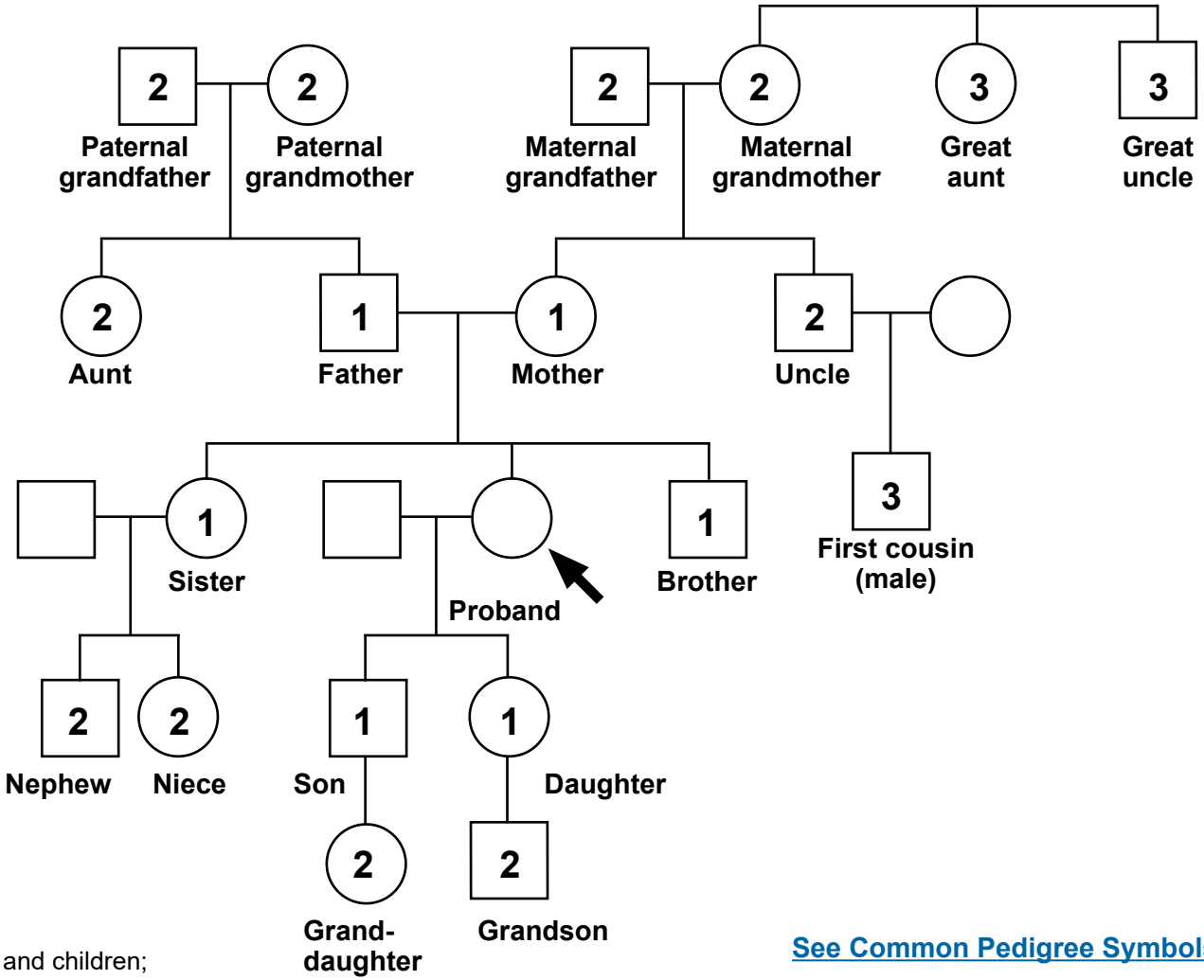
Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF CANCER RISK ASSESSMENT AND COUNSELING

PEDIGREE: FIRST-, SECOND-, AND THIRD-DEGREE RELATIVES OF PROBAND[13]



[13]First-degree relatives: parents, siblings, and children;
Second-degree relatives: grandparents, aunts, uncles, nieces, nephews, grandchildren, and half-siblings;
Third-degree relatives: great-grandparents, great-aunts, great-uncles, great-grandchildren, first cousins, and half aunts and half uncles.

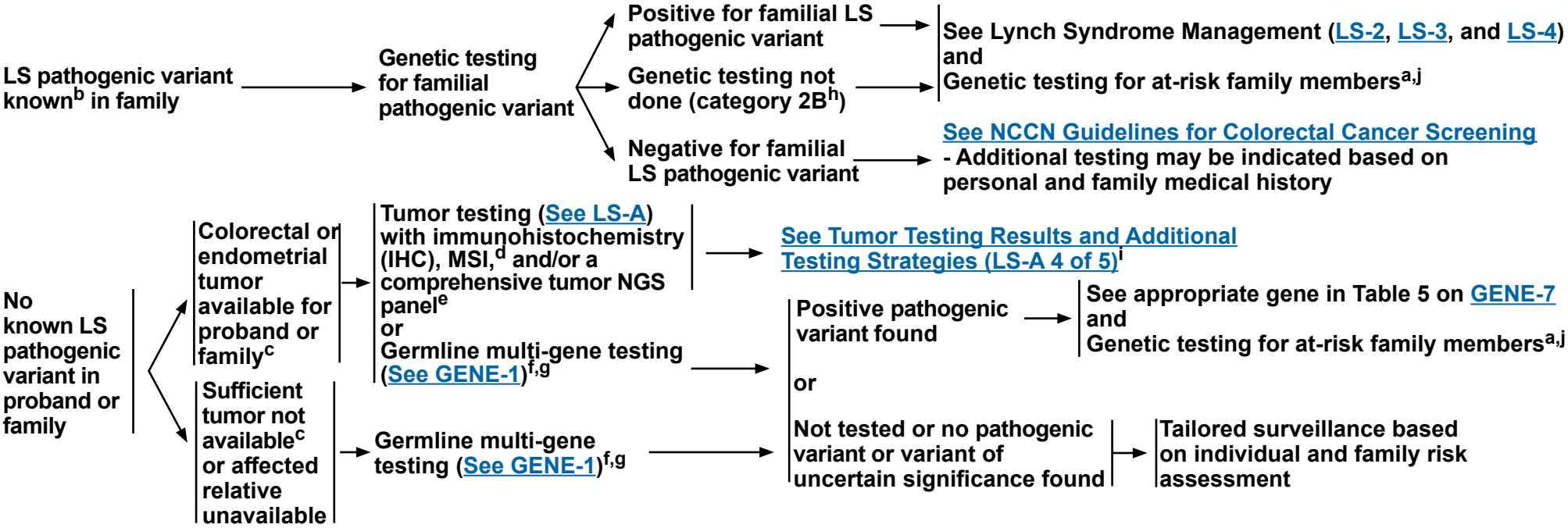
See Common Pedigree Symbols (HRS-A 5 of 6)

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

STRATEGIES FOR EVALUATING FOR LS IN INDIVIDUALS MEETING CRITERIA FOR THE EVALUATION OF LS[a]



[a] An individual with expertise in genetics should be involved in the testing process. Minimum pretest counseling (in person or through written or video) materials with pros and cons of testing should be provided. See Principles of Cancer Risk Assessment and Counseling (HRS-A 1 of 6).

[b] Irrespective of degree of relatedness.

[c] If there is more than one affected family member, first consider: youngest age at diagnosis, multiple primaries, and colorectal or endometrial cancers. Limitations of interpreting test results should be discussed if testing tumors other than colorectal or endometrial cancers. If IHC/MSI previously done, see LS-A 4 of 5.

[d] The panel recommends tumor testing with IHC and/or MSI be used as the primary approach for pathology lab-based universal screening.

[e] Tumor NGS panels should include at a minimum the MMR genes (*MLH1, MSH2, MSH6, PMS2, and EPCAM),* other known familial cancer genes, MSI, and *BRAF*.

[f] This approach may be preferred in patients with a strong family history or if diagnosed age <50 y (Pearlman R, et al. JAMA Oncol 2017;3:464-471; Yurgelun M, et al. J Clin Oncol 2017;35:1086-1095).

[g] Testing of unaffected family members when no affected member is available should be considered. Significant limitations of interpreting test results should be discussed.

[h] The recommendation to manage patients in whom genetic testing was not done using LS-management recommendations is category 2B.

[i] For individuals found to have an LS pathogenic variant, see LS management recommendations.

[j] If a first-degree relative is unavailable or unwilling to be tested, more distant relatives should be offered testing for the known pathogenic variant in the family.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

LYNCH SYNDROME MANAGEMENT

Surveillance/Prevention Strategies for *MLH1*, *MSH2*, *MSH6*, *PMS2*, and *EPCAM* Pathogenic Variant Carriers[k,l,m]

- **Colon cancer:**
  - **Colonoscopy at age 20–25 y[n] or 2–5 y prior to the earliest colon cancer if it is diagnosed before age 25 y and repeat every 1–2 y.[o]**
  - **There are data to suggest that aspirin may decrease the risk of colon cancer in LS, but optimal dose and duration of aspirin therapy are uncertain.**

→ See Follow-up of Surveillance Findings (LS-5)

Other Extracolonic Cancers

- **Gastric and small bowel cancer:**
  - **There are no clear data to support surveillance for gastric, duodenal, and more distal small bowel cancer for LS. Selected individuals with a family history of gastric, duodenal, or more distal small bowel cancer may have increased risk, and may benefit from surveillance. Also, individuals of Asian descent (or from countries with high background incidence of gastric cancer) may have increased risk for stomach cancer and may benefit from surveillance (Vasen HF, et al. Gut 2013;62:812-823). If surveillance is performed, may consider upper endoscopy with visualization of the duodenum at the time of colonoscopy every 3–5 y beginning at age 40 y.[p] Consider H. pylori testing and treating H. pylori, if detected.**
- **Urothelial cancer:**
  - **There is no clear evidence to support surveillance for urothelial cancers in LS. Surveillance may be considered in selected individuals such as with a family history of urothelial cancer or individuals with *MSH2* pathogenic variants (especially males) as these groups appear to be at higher risk. Surveillance options may include annual urinalysis starting at age 30–35 y. However, there is insufficient evidence to recommend a particular surveillance strategy.**
- **Central nervous system (CNS) cancer:**
  - **Consider annual physical/neurologic examination starting at age 25–30 y; no additional screening recommendations have been made.**
- **Pancreatic cancer:**
  - **Despite data indicating an increased risk for pancreatic cancer, the panel is not able to make a screening recommendation at this time.[q]**
- **Breast cancer:**
  - **There have been suggestions that there is an increased risk for breast cancer in LS patients; however, there is not enough evidence to support increased screening above average-risk breast cancer screening recommendations or those based on personal/family history of breast cancer. See NCCN Guidelines for Breast Cancer Screening and Diagnosis.**
- **Prostate cancer:**
  - **At present, there is insufficient evidence to recommend earlier or more frequent prostate cancer screening among men with LS. However, men with LS should be encouraged to participate in prostate cancer screening as recommended in the NCCN Guidelines for Prostate Cancer.**

[k] See Cancer Risks in Lynch Syndrome by Gene Compared to the General Population (LS-B).

[l] Other than colon and endometrial cancer, surveillance recommendations are expert opinion rather than evidence-based.

[m] The panel recognizes that there are limited population-based studies on the lifetime risk for most of the cancers related to each of these genes. Although there are some pathogenic variant-specific data available, a generalized screening approach is suggested. Screening and the option of risk-reducing surgeries should be individualized after risk assessment and counseling.

[n] For *MSH6*, consider a later age for colonoscopy initiation such as at age 30 years or 10 years younger than age of any relative with CRC. Due to limited data for the *PMS2* gene, the panel is not able to make a specific recommendation regarding later age of onset for colonoscopy.

[o] Patients who may benefit from a shorter 1- versus longer 2-year interval include those with risk factors such as history of CRC, male sex, *MLH1/MSH2* pathogenic variant, age >40 years, and history of adenoma. See Discussion.

[p] Møller P, et al. Cancer risk and survival in path_MMR carriers by gene and gender up to 75 years of age: a report from the Prospective Lynch Syndrome Database. Gut 2018;67:1306-1316.

[q] If screening is performed, it should be considered in high-volume centers with multidisciplinary teams, preferably within research protocols. The International Cancer of the Pancreas Screening (CAPS) consortium recommends that patients with LS with one first-degree relative with pancreatic cancer should be considered for screening.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Lynch Syndrome Management continued on LS-3 and LS-4

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

LYNCH SYNDROME MANAGEMENT

Surveillance/Prevention Strategies for *MLH1*, *MSH2*, *MSH6, PMS2,* and *EPCAM* Pathogenic Variant Carriers[k,l,m]

Other Extracolonic Cancers

- **Endometrial cancer:**
  - **Because endometrial cancer can often be detected early based on symptoms, women should be educated regarding the importance of prompt reporting and evaluation of any abnormal uterine bleeding or postmenopausal bleeding. The evaluation of these symptoms should include endometrial biopsy.**
  - **Hysterectomy has not been shown to reduce endometrial cancer mortality, but can reduce the incidence of endometrial cancer. Therefore, hysterectomy is a risk-reducing option that can be considered.**
  - **Timing of hysterectomy can be individualized based on whether childbearing is complete, comorbidities, family history, and LS gene pathogenic variant, as risks for endometrial cancer vary by pathogenic variant.**
  - **Endometrial cancer screening does not have proven benefit in women with LS. However, endometrial biopsy is both highly sensitive and highly specific as a diagnostic procedure. Screening via endometrial biopsy every 1 to 2 years can be considered.**
  - **Transvaginal ultrasound to screen for endometrial cancer in postmenopausal women has not been shown to be sufficiently sensitive or specific as to support a positive recommendation, but may be considered at the clinician's discretion. Transvaginal ultrasound is not recommended as a screening tool in premenopausal women due to the wide range of endometrial stripe thickness throughout the normal menstrual cycle.**
- **Ovarian cancer:**
  - **Bilateral salpingo-oophorectomy (BSO) may reduce the incidence of ovarian cancer. The decision to have a BSO as a risk-reducing option by women who have completed childbearing can be individualized. Timing of BSO should be individualized based on whether childbearing is complete, menopause status, comorbidities, family history, and LS gene pathogenic variant, as risks for ovarian cancer vary by mutated gene. Insufficient evidence exists to make specific recommendation for risk-reducing salpingo-oophorectomy (RRSO) in *MSH6* and *PMS2* pathogenic variant carriers (see LS-B). See ovarian age-specific risks on LS-B 2 of 2.**
  - **Since there is no effective screening for ovarian cancer, women should be educated on the symptoms that might be associated with the development of ovarian cancer, such as pelvic or abdominal pain, bloating, increased abdominal girth, difficulty eating, early satiety, or urinary frequency or urgency. Symptoms that persist for several weeks and are a change from a woman's baseline should prompt evaluation by her physician.**
  - **While there may be circumstances where clinicians find screening helpful, data do not support routine ovarian cancer screening for LS. Transvaginal ultrasound for ovarian cancer screening has not been shown to be sufficiently sensitive or specific as to support a routine recommendation, but may be considered at the clinician's discretion. Serum CA-125 is an additional ovarian screening test with caveats similar to transvaginal ultrasound.**
- **Consider risk reduction agents for endometrial and ovarian cancers, including discussing risks and benefits (See Discussion for details).**

[k] See Cancer Risks in Lynch Syndrome by Gene Compared to the General Population (LS-B).

[l] Other than colon and endometrial cancer, surveillance recommendations are expert opinion rather than evidence-based.

[m] The panel recognizes that there are limited population-based studies on the lifetime risk for most of the cancers related to each of these genes. Although there are some pathogenic variant-specific data available, a generalized screening approach is suggested for many surveillance strategies. Screening and the option of risk-reducing surgeries should be individualized after risk assessment and counseling.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Lynch Syndrome Management continued on LS-4.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

LYNCH SYNDROME MANAGEMENT

Surveillance/Prevention for *MLH1*, *MSH2*, *MSH6, PMS2,* and *EPCAM* Pathogenic Variant Carriers[k,l,m]

Reproductive Options

- **For patients of reproductive age, advise about options for prenatal diagnosis and assisted reproduction including pre-implantation genetic diagnosis. Discussion should include known risks, limitations, and benefits of these technologies.**
- **For patients of reproductive age, advise about the risk of a rare recessive syndrome called constitutional MMR deficiency (CMMRD) syndrome; Wimmer K, et al. J Med Genet 2014;51:355-365. If both partners are a carrier of a pathogenic variant/s in the same MMR gene or *EPCAM* (for example, if both partners carry a pathogenic variant in the *PMS2* gene), then their future offspring will be at risk of having CMMRD syndrome.**

Risk to Relatives

- **Advise patients to tell their relatives about possible inherited cancer risk, options for risk assessment, and management.**
- **Recommend genetic counseling and consideration of genetic testing for at-risk relatives.**

[k] See Cancer Risks Up to Age 70 Years in Individuals with Lynch Syndrome Compared to the General Population (LS-B).
[l] Other than colon and endometrial cancer, surveillance recommendations are expert opinion rather than evidence-based.
[m] The panel recognizes that there are limited population-based studies on the lifetime risk for most of the cancers related to each of these genes. However, there are some pathogenic variant-specific data available and a generalized screening approach is suggested. Screening and the option of risk-reducing surgeries should be individualized after risk assessment and counseling.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

SURVEILLANCE COLONOSCOPY FINDINGS	FOLLOW-UP	
No pathologic findings	Continued surveillance every 1–2 y[o,r]	
Adenocarcinomas	• See appropriate NCCN Guidelines for Treatment of Cancer by Site • For patients with colorectal adenocarcinoma, segmental or extended colectomy depending on clinical scenario should be considered[s]	
Adenomas	Complete endoscopic polypectomy with follow-up colonoscopy every 1–2 y	
Adenomas not amenable to endoscopic resection	Segmental or extended colectomy depending on clinical scenario[s]	Examine all remaining colonic mucosa every 1–2 y
Adenomas with high-grade dysplasia	Colonoscopy surveillance every 1–2 years[t] or Segmental or extended colectomy[t]	

[o] Patients who may benefit from a shorter 1- versus longer 2-year interval include those with risk factors such as history of CRC, male sex, MLH1/MSH2 pathogenic variant, age >40 years, and history of adenoma. See Discussion.
[r] May consider subtotal colectomy if patient is not a candidate for optimal surveillance.
[s] The type of surgical procedure chosen should be based on individual considerations and discussion of risk. See Definitions of Common Colorectal Resections (CSCR-B) in the NCCN Guidelines for Colorectal Cancer Screening.
[t] Surgery is recommended if resection was not en bloc or if dysplasia involved the resection margin, whereas surveillance may be considered if an en bloc complete excision was performed.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF IHC AND MSI TESTING FOR LYNCH SYNDROME

- The panel recommends universal screening of all CRCs to maximize sensitivity for identifying individuals with Lynch syndrome and to simplify care processes. Counseling by an individual with expertise in genetics is not required prior to routine tumor testing. An infrastructure needs to be in place to handle the screening results.

General

- IHC and MSI analyses are screening tests (either by themselves or in conjunction) that are typically done on colon and endometrial cancer tissue to identify individuals at risk for LS. Greater than 90% of LS tumors are MSI-H and/or lack expression of at least one of the MMR proteins by IHC. Ten percent to 15% of sporadic colon cancers exhibit abnormal IHC and are MSI-H most often due to abnormal methylation of the *MLH1* gene promoter, rather than due to LS. Mutant *BRAF* V600E is found in many sporadic MSI-H CRCs and is rarely found in LS-related CRCs. There are some tumors that will have *MLH1* methylation but lack a *BRAF* pathogenic variant. Thus, the presence of an abnormal *MLH1* IHC test increases the possibility of LS but does not make a definitive diagnosis. Confirmed diagnosis of LS is based on germline testing, when tumor-based testing scenarios or other factors raise suspicion for the diagnosis (see LS-A 4 of 5). Also, sporadic endometrial cancers may exhibit abnormal MSI/IHC due to abnormal methylation of the *MLH1* promoter. Somatic MMR genetic testing of the corresponding gene(s) (see “Plausible Etiologies” for possibilities on LS-A 4 of 5) could be performed on tumor DNA to assess for pathogenic variants that might explain the abnormal IHC and/or MSI-H results.
- The panel recommends a universal screening strategy be the primary approach to identify CRC patients with LS. However, in other lower resource settings, other historic criteria for selecting patients for testing may be relevant. The Bethesda criteria (~~See LS-1~~ See Discussion) are intended to help identify CRC patients whose tumors should be tested for MMR defects, by MSI and/or IHC analysis, thereby identifying patients with a greater chance of having LS.

IHC

- IHC refers to staining tumor tissue for protein expression of the 4 MMR genes known to be mutated in LS: *MLH1*, *MSH2*, *MSH6*, and *PMS2*. A normal IHC test implies all 4 MMR proteins are normally expressed, and thus it is unlikely that an underlying MMR gene pathogenic variant is present. An abnormal test means that at least one of the proteins is "not detected," and an inherited pathogenic variant may be present in the related gene. Loss of protein expression by IHC in any one of the MMR genes guides further genetic testing (pathogenic variant detection) to the gene(s) where protein expression is not observed or to the corresponding protein dimer. Absent expression of one or more of the 4 DNA MMR proteins is often reported as abnormal or “positive” IHC. When “positive” IHC is reported, caution should be taken in making sure that positive refers to absence of MMR protein expression, and not to presence of expression.
- Abnormal *MLH1* IHC should be followed by either germline genetic testing or tumor testing for *MLH1* methylation for colorectal or endometrial cancers. Alternatively for colorectal cancers with loss of *MLH1* on IHC, the tumor can be tested for a *BRAF* V600E pathogenic variant. Testing for *BRAF* pathogenic variants using IHC is not sufficiently sensitive in general but it may be an option for situations with insufficient tumor material for molecular testing since it only requires one slide. Presence of *MLH1* hypermethylation, *BRAF* V600E pathogenic variant, or abnormal *BRAF* V600E protein by IHC is consistent with sporadic cancer. If MLH1 promoter methylation or *BRAF* testing is normal, genetic testing is indicated (See LS-A 4 of 5). Those with a germline pathogenic variant are then identified as LS patients. *BRAF* V600E pathogenic variants are found in 69% of methylated colorectal cancers, so the absence of a *BRAF* V600E pathogenic variant does not rule out methylation. As a result, there may be a role for methylation testing to rule out Lynch syndrome in MSI-H tumors in which no *BRAF* pathogenic variant is found either prior to genetic testing or in the event genetic testing is negative. If abnormal IHC is followed by germline testing and no LS-causing pathogenic variants are identified, the panel strongly recommends proceeding with MLH methylation analysis of the tumor. Patients who have normal germline testing and MLH1 hypermethylation are likely to have sporadic cancer and should be treated as such taking into account their family history.
- If clinical suspicion for LS is high despite a normal IHC screening result, consider genetic evaluation and testing.
- There is a 5%–10% false-negative rate with IHC testing.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF IHC AND MSI TESTING FOR LYNCH SYNDROME

IHC (continued)

- **Adenomas:**
  - **IHC can also be performed on colorectal adenomas if cancer tissue is not available. Abnormal loss of staining can be identified in as many as 70%–79% of Lynch-associated adenomas. Adenoma size >10 mm and/or the presence of high-grade dysplasia within the polyp increases sensitivity of IHC for LS.[1,2,3] The suboptimal sensitivity of IHC performed on polyps means this approach should not be used to exclude LS. An abnormal polyp IHC result should be referred for genetic evaluation and testing. If *PMS2* and *MLH1* are missing, further tumor testing should be considered before referring for genetic testing.**
- **Rectal cancers treated with neoadjuvant chemotherapy and radiation therapy (RT):[4]**
  - **False abnormal IHC has been reported in rectal cancer resection specimens after neoadjuvant chemotherapy and RT. As a result, some NCCN Member Institutions avoid doing IHC on rectal cancers after neoadjuvant chemotherapy and RT. Others still perform IHC on rectal cancers after neoadjuvant chemotherapy and RT, but if expression is absent (particularly *MSH6* or *PMS2*) the testing is repeated on the pretreatment biopsy.**
- **Sebaceous neoplasms:[5-9]**
  - **The sensitivity and specificity of MMR IHC on sebaceous neoplasms in LS is much lower than that of CRC (85% vs. 92%–94% and 48% vs. 88%–100%). The false-positive rate has been reported to be 56%. A scoring system taking into account age at diagnosis, number of sebaceous neoplasms, and personal or family history of LS-associated cancers can be used to determine which patients with sebaceous neoplasms need IHC.[9]**
- **Metastatic CRC (liver, lymph node, and other metastases):[10]**
  - **There are data showing that the MSI and IHC results in primary tumors match the MSI and IHC results in metastatic tissue from the same tumor; therefore, this should be an acceptable alternative if the primary tumor is not available.**

[1] Pino MS, et al. Deficient DNA mismatch repair is common in Lynch syndrome-associated colorectal adenomas. J Mol Diagn 2009;11:238-247.
[2] Walsh MD, et al. Immunohistochemical testing of conventional adenomas for loss of expression of mismatch repair proteins in Lynch syndrome mutation carriers: a case series from the Australasian site of the colon cancer family registry. Mod Pathol 2012;25:722-730.
[3] Yurgelun MB, Goel A, Hornick JL, et al. Microsatellite instability and DNA mismatch repair protein deficiency in Lynch syndrome colorectal polyps. Cancer Prev Res (Phila) 2012;5:574-582.
[4] Vilkin A, Halpern M, Morgenstern S, et al. How reliable is immunohistochemical staining for DNA mismatch repair proteins performed after neoadjuvant chemoradiation? Hum Pathol 2014;45:2029-2036.
[5] Roberts ME, Riegert-Johnson DL, Thomas BC, et al. Screening for Muir-Torre syndrome using mismatch repair protein immunohistochemistry of sebaceous neoplasms. J Genet Couns 2013;22:393-405.
[6] Hampel H, Frankel WL, Martin E, et al. Feasibility of screening for Lynch syndrome among patients with colorectal cancer. J Clin Oncol 2008;26:5783-5788.
[7] Hampel H, Stephens JA, Pukkala E, et al. Cancer risk in hereditary nonpolyposis colorectal cancer syndrome: later age of onset. Gastroenterology 2005;129:415-421.
[8] Lindor NM, Burgart LJ, Leontovich O, et al. Immunohistochemistry versus microsatellite instability testing in phenotyping colorectal tumors. J Clin Oncol 2002;20:1043-1048.
[9] Roberts ME, Riegert-Johnson DL, Thomas BC, et al. A clinical scoring system to identify patients with sebaceous neoplasms at risk for the Muir-Torre variant of Lynch syndrome. Genet Med 2014;16:711-716.
[10] Haraldsdottir S, Roth R, Pearlman R, et al. Mismatch repair deficiency concordance between primary colorectal cancer and corresponding metastasis. Fam Cancer 2016;15:253-260.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PRINCIPLES OF IHC AND MSI TESTING FOR LYNCH SYNDROME

MSI

- **MSI-H in tumors refers to the tumor having a proportion of alterations in a predetermined panel of microsatellite repeat markers that indicates the loss of MMR activity. Its significance, use, and implications are similar to that of IHC, although the tests are slightly complementary.**
- **Laboratories vary in their approach in testing MSI. Dinucleotide markers may be less specific than mononucleotide markers of MSI.[11]**
- **There is a 5%–10% false-negative rate with MSI testing.**

Pros and Cons of Universal Tumor Screening with IHC and/or MSI for LS Using Colonoscopy-Based Biopsy Versus Surgical Resection Specimen[12,13]

Pre-surgical testing considerations

- **Pros**
  - **Informs surgical decision-making (subtotal vs. segmental resection)**
  - **For rectal tumors requiring neoadjuvant chemotherapy and RT, IHC is more reliable when done on pre-therapy specimens[14,15]**
- **Cons**
  - **Possibility of insufficient tissue for analysis**
  - **Screening could be done twice (once on biopsy and once on surgical resection), thereby decreasing cost-effectiveness**

Surgical testing considerations

- **Pros**
  - **Can perform MSI and/or IHC**
  - **Ensures test is only done once**
- **Cons**
  - **Cannot inform surgical decision-making**
  - **In rectal tumors exposed to neoadjuvant chemotherapy and RT, IHC may be less reliable, with the potential for false negative result (particularly *MSH6*)**

[11] Xicola RM, Llor X, Pons E. Performance of different microsatellite marker panels for detection of mismatch repair-deficient colorectal tumors. J Natl Cancer Inst 2007;99:244-252.
[12] Kumarasinghe AP, de Boer B, Bateman AC, Kumarasinghe MP. DNA mismatch repair enzyme immunohistochemistry in colorectal cancer: a comparison of biopsy and resection material. Pathology 2010;42:414-420.
[13] Shia J, Stadler Z, Weiser MR, et al. Immunohistochemical staining for DNA mismatch repair proteins in intestinal tract carcinoma: How reliable are biopsy samples? Am J Surg Pathol 2011;35:447-454.
[14] Bao F, Panarelli NC, Rennert H, et al. Neoadjuvant therapy induces loss of MSH6 expression in colorectal carcinoma. Am J Surg Pathol 2010;34:1798-1804.
[15] Radu OM, Nikiforova MN, Farkas LM, Krasinskas AM. Challenging cases encountered in colorectal cancer screening for Lynch syndrome reveal novel findings: nucleolar MSH6 staining and impact of prior chemoradiation therapy. Hum Pathol 2011;42:1247-1258.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

TUMOR TESTING RESULTS AND ADDITIONAL TESTING STRATEGIES[j]

Tumor Testing[a]							Plausible Etiologies	Additional Testing[d,e] NOTE: If younger than age 50 regardless of LS test results, consider genetic evaluation
IHC				MSI	*BRAF* V600E[b]	MLH1 Promoter Methylation		
MLH1	*MSH2*	*MSH6*	*PMS2*					
NL	NL	NL	NL	MSS/MSI-Low	N/A	N/A	1) Sporadic cancer 2) Other (not LS hereditary CRC syndrome)	1) None[c]
NL	NL	NL	NL	MSI-High	N/A	N/A	1) Germline pathogenic variant in any LS gene 2) Sporadic cancer	1) Germline LS genetic testing[f] 2) If germline testing negative, consider somatic MMR genetic testing[h]
N/A	N/A	N/A	N/A	MSI-High	N/A	N/A	1) Sporadic cancer 2) Germline pathogenic variant in any of the LS genes	1) Consider IHC analysis and additional testing depending on IHC results 2) If IHC not performed, consider germline LS genetic testing[f]
AB	NL	NL	AB	N/A	N/A	N/A	1) Sporadic cancer 2) Germline *MLH1* pathogenic variant or rarely *PMS2*	1) Consider *BRAF* pathogenic variant testing[b]/MLH1 promoter methylation 2) Germline LS genetic testing[f]
AB	NL	NL	AB	N/A	Positive	N/A	1) Sporadic cancer 2) Rarely germline *MLH1* pathogenic variant or constitutional *MLH1* epimutation	1) None, unless young age of onset or significant family history; then consider constitutional *MLH1* epimutation testing[g] and/or germline LS genetic testing[f]
AB	NL	NL	AB	N/A	Negative	Positive	1) Sporadic cancer 2) Rarely germline *MLH1* pathogenic variant or constitutional *MLH1* epimutation	
AB	NL	NL	AB	N/A	Negative	Negative	1) Germline *MLH1* pathogenic variant or rarely *PMS2* *2)* Sporadic cancer	1) Germline LS genetic testing[f] 2) If germline testing negative, consider somatic MMR genetic testing[h]
NL	AB	AB	NL	N/A	N/A	N/A	1) Germline *MSH2*/*EPCAM* pathogenic variant; or rarely germline *MSH6* pathogenic variant *2)* Sporadic cancer	
NL	NL	NL	AB	N/A	N/A	N/A	1) Germline *PMS2* pathogenic variant 2) Germline *MLH1* pathogenic variant 3) Sporadic cancer	
NL	AB	NL	NL	N/A	N/A	N/A	1) Germline *MSH2/EPCAM* pathogenic variant 2) Sporadic cancer	
NL	NL	AB	NL	N/A	N/A	N/A	1) Germline *MSH6* pathogenic variant 2) Germline *MSH2* pathogenic variant 3) Sporadic cancer/Treatment effect[i]	1) Germline LS genetic testing[f] 2) If applicable, consider MSI analysis or repeat IHC testing on nontreated tumor[i] 3) If germline testing negative, consider somatic MMR genetic testing[h]
AB	NL	NL	NL	N/A	N/A	N/A	1) Sporadic cancer; 2) Germline *MLH1* pathogenic variant; 3) Germline *PMS2* pathogenic variant; 4) Somatic *MLH1* or *PMS2* pathogenic variant	1) *BRAF* pathogenic variant testing[b]/MLH1 promoter methylation; 2) if *BRAF*/*MLH1* methylation testing normal, germline LS genetic testing including at least the *MLH1* and *PMS2* genes; 3) If germline testing negative, consider somatic MMR sequencing of the tumor DNA
AB	AB	AB	AB	N/A	N/A	N/A	1) Germline pathogenic variant in *any* LS gene 2) Sporadic cancer	1) Germline LS genetic testing[f] 2) If germline testing of *MLH1* negative, consider *BRAF*[b]/methylation studies 3) If germline testing negative, consider somatic MMR genetic testing[h]

N/A = Either testing was not done or results may not influence testing strategy; NL = Normal presence of positive protein staining; AB-= Abnormal/Absence (negative) protein staining

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Footnotes

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

TUMOR TESTING RESULTS AND ADDITIONAL TESTING STRATEGIES

Footnotes from LS-A 4 of 5

[a]Tumor testing strategies apply to colorectal and endometrial cancers. Limited data exist regarding the efficacy of tumor testing in other LS tumors.
[b]Testing is not appropriate for tumors other than CRC.
[c]If strong family history (ie, Amsterdam criteria) or additional features of hereditary cancer syndromes (multiple colon polyps) are present, additional testing may be warranted in the proband, or consider tumor testing in another affected family member due to the possibility of a phenocopy.
[d]Studies have shown that 45%–68% of cases with unexplained defective MMR (MSI-H and/or abnormal IHC with no evidence of *MLH1* promoter hypermethylation when indicated) have double somatic pathogenic variants (either two pathogenic sequence variants or one pathogenic sequence variant and loss of heterozygosity [LOH]) in the MMR genes. (Sourrouille I, Coulet F, Lefevre JH, et al. Fam Cancer 2013;12:27-33. Mensenkamp A, Vogelaar I, van Zelst-Stams W, et al. Gastroenterology 2014;146:643-646. Geurts-Giele W, Leenen C, Dubbink H, et al. J Pathol 2014;234:548-559. Haraldsdottir S, Hampel H, Tomsic J, et al. Gastroenterology 2014;147:1308-1316.) As a result, tumor sequencing may be helpful for individuals with tumor testing showing deficient MMR and no germline pathogenic variant detected. If double somatic pathogenic variants are identified or if the testing does not help clarify the result, *it is recommended that these patients and their close relatives be managed based on their family history and NOT as if they have LS.* If double somatic pathogenic variants are identified, LS is ruled out but there may still be some increased familial risk. If only one somatic pathogenic variant is found, the unidentified pathogenic variant could either be germline or somatic. If no somatic pathogenic variants are found, it is possible that the IHC results were incorrect (especially if the tumor was found to be microsatellite stable on tumor sequencing) or that none of the pathogenic variants (germline or somatic) are identifiable. In any of these cases, the patient and their close relatives still need to be managed based on their family history. Genetic consultation should be considered for interpretation of complex results.
[e]Prior to germline genetic testing, proper pre-test counseling should be done by an individual with expertise in genetics.
[f]Germline LS genetic testing may include testing of the gene(s) that are indicated (see “Plausible Etiologies” for possibilities on LS-A 4 of 5) by the abnormal tumor test results; or instead, multigene testing that includes *MLH1*, *MSH2*, *MSH6*, *PMS2*, and *EPCAM* concurrently may be performed.
[g]Evaluation for constitutional *MLH1* epimutation involves *MLH1* promoter hypermethylation studies on blood or other sources of normal tissue.
[h]Somatic MMR genetic testing of the corresponding gene(s) (see “Plausible Etiologies” for possibilities on LS-A 4 of 5) could be performed on tumor DNA to assess for somatic pathogenic variants that might explain the abnormal IHC and/or MSI results.
[i]Absent *MSH6* in rectal tumor tissue may be due to treatment effect (neoadjuvant chemoradiotherapy).
[j]These tumor testing results may also have implications for treatment in cases that are sporadic or hereditary. See The NCCN Guidelines for Colon Cancer for more information on pathologic review and the impact on management (COL-B 4 of 6). Consult with an expert if the scenario is not covered by this table.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Cancer Risks in Lynch Syndrome by Gene Compared to the General Population

	General Population Risk[1]	*MLH1*		*MSH2* (For EPCAM, see footnote 10)		*MSH6*		*PMS2*	
		Risk	Average age of diagnosis	Risk	Average age of diagnosis	Risk	Average age of diagnosis	Risk	Average age of diagnosis
Colorectal[1-6]	4.5%	46%–49%	43–45 years	43%–52%	44 years	15%–44%	51–63 years	12%–20%	47–66 years
Endometrial[1-6]	2.7%	43%–57%	49 years	21%–57%	47–48 years	17%–46%	53–55 years	0%–15%	49–56 years
Breast[2,3,7]	13%	12%–17%	53 years	12%	52 years	0%–13%	52 years	NE	
Ovarian[1,2,7]	1.3%	5%–20%	44–47 years	10%–38%	43–44 years	1%–11%	44–48 years	NE	
Gastric[1,2,7,8]	<1%	5%–7%	49–52 years	0.2%–16%	49–52 years	0%–5%	49–63 years	NE	
Pancreas[2]	1.5%	6%	52–57 years	NE		NE		NE	
Bladder[2,7,9]	2.5%	2%–4%	53–59 years	4%–17%	53–59 years	2%	53–71 years	NE	
Biliary tract[1,2]	<1%	2%–4%	50 years	0.02%	57 years	NE		NE	
Urothelial[1,2,7,9]	<1%	0.2%–5%	52-60 years	2%–18%	52–61 years	0.7%–7%	52–69 years	NE	
Small bowel[1,7]	<1%	0.4%–11%	46-47 years	1%–10%	46–48 years	0%–3%	46–54 years	NE	
Prostate[2,3,7,11]	11.6%	0%–17%	59 years	30%–32%	59 years	0%–5%	59 years	NE	
Brain/CNS[2]	<1%	NE		NE		Not reported	Not reported	NE	

NE = Not well established. The panel cautions that new data may confirm or change prior findings suggesting no increased risk, as more studies are needed to clarify lifetime risks for cancer in LS by mutation type.

Continued
Footnotes

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Cancer Risk Up to Age 70 Years in Individuals with Lynch Syndrome Compared to the General Population

Age-Specific Risks for Ovarian Cancer

	MLH1 Cumulative Risk by Age in Years, % (95% confidence interval)				Mean Age of Diagnosis
	40	50	60	70	
Ref. 1	0 (0-2)	4 (0-11)	15 (1-45)	20 (1-65)	44–47 years
Ref. 12	1 (0-3.6)	7 (2.2-11.2)	9 (2.9-12.2)	11 (3.2-19.8)	

	MSH2 Cumulative Risk by Age in Years, % (95% confidence interval)				Mean Age of Diagnosis
	40	50	60	70	
Ref. 1	1 (0-3)	4 (1-9)	11 (2-28)	24 (3-52)	43–44 years
Ref. 12	4 (0.0-8.9)	12 (4.2-20.2)	15 (5.5-24.4)	15 (5.5-24.4)	

	MSH6 Cumulative Risk by Age in Years, % (95% confidence interval)				Mean Age of Diagnosis
	40	50	60	70	
Ref. 1	0	0 (0-1)	1 (0-2)	1 (0-3)	44–48 years
Ref. 12	0 (-)	0 (-)	0 (-)	0 (-)	

	PMS2 Cumulative Risk by Age in Years, % (95% confidence interval)				Mean Age of Diagnosis
	40	50	60	70	
Ref. 5	†	†	†	†	NE
Ref. 12	0 (-)	0 (-)	0 (-)	0 (-)	

† The combined risk for renal pelvic, stomach, ovary, small bowel, ureter, and brain is 6% to age 70 (Senter L, Clendenning M, Sotamaa K, et al. The clinical phenotype of LS due to germ-line PMS2 mutations. Gastroenterology 2008;135:419-428). NE = Not well established.

[1] Bonadona, Bonaiti B, Olschwang S, et al. Cancer risks associated with germline mutations in MLH1, MSH2, and MSH6 genes in Lynch syndrome. JAMA 2011;305:2304-2310.
[2] Møller P, Seppälä TT, Bernstein I, et al. Cancer risk and survival in path_MMR carriers by gene and gender up to 75 years of age: a report from the Prospective Lynch Syndrome Database. Gut 2018;67:1306-1316.
[3] Baglietto L, Lindor NM, Dowty JG, et al. Risks of Lynch syndrome cancers for MSH6 mutation carriers. J Natl Cancer Inst 2010;102:193-201.
[4] Ten Broeke S, van der Klift H, Tops C, et al. Cancer risks for PMS2-associated Lynch syndrome. J Clin Oncol 2018;36:2961-2968.
[5] Senter L, Clendenning M, Sotamaa K, et al. The clinical phenotype of Lynch syndrome due to germ-line PMS2 mutations. Gastroenterology 2008;135:419-428.
[6] Ryan N, Morris J, Green K, et al. Association of mismatch repair mutation with age at cancer onset in Lynch syndrome: Implications for stratified surveillance strategies. JAMA Oncol 2017;3:1702-1706.
[7] Engel C, Loeffler M, Steinke V, et al. Risks of less common cancers in proven mutation carriers with lynch syndrome. J Clin Oncol 2012;30:4409-4415.
[8] Capelle LG1, Van Grieken NC, Lingsma HF, et al. Risk and epidemiological time trends of gastric cancer in Lynch syndrome carriers in the Netherlands. Gastroenterology 2010;138:487-492.
[9] Joost P, Therkildsen C, Dominguez-Valentin M, et al. Urinary tract cancer in Lynch syndrome; Increased risk in carriers of MSH2 mutations. Urology 2015;86:1212-1217.
[10] While EPCAM risks have not been separated out in this table, there has been an observation that based on the size of the pathogenic variant and how far it extends that the phenotype can be similar to MSH2. (Ligtenberg MJ, Kuiper RP, Geurts van Kessel A, Hoogerbrugge N. EPCAM deletion carriers constitute a unique subgroup of Lynch syndrome patients. Fam Cancer 2013;12:169-174. Huth C, Kloor M, Voigt AY, et al. The molecular basis of EPCAM expression loss in Lynch syndrome-associated tumors. Mod Pathol 2012;25:911-916.)
[11] A meta-analysis of studies (Ryan S, et al. Cancer Epidemiol Biomarkers Prev 2014;23:437-449) regarding prostate cancer risk in LS found that men with LS were 2.13–3.67 times more likely to develop prostate cancer. In addition, they found that 73% of prostate cancers in men with LS exhibit dMMR with IHC results consistent with the underlying germline pathogenic variant, suggesting that the prostate cancer was caused by the LS. Only one study includes absolute risks, which are presented here since they are in line with the relative risks reported thus far. There are too few prostate cancer cases among *PMS2* pathogenic variants carriers to know if they are at increased risk.
[12] Moller P, Seppala T, Bernstein I, et al. Cancer incidence and survival in Lynch syndrome patients receiving colonoscopic and gynaecological surveillance: first report from the prospective Lynch syndrome database. Gut 2017;66:464–472.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Adenomatous Polyposis Testing Criteria

NCCN Guidelines Index
Table of Contents
Discussion

ADENOMATOUS POLYPOSIS TESTING CRITERIA

- Personal history of ≥20 cumulative adenomas
- Known pathogenic variant in adenomatous polyposis gene in family[a]
- Consider testing if a personal history of a desmoid tumor, hepatoblastoma, cribriform-morular variant of papillary thyroid cancer, or multifocal/bilateral CHRPE, if individual meets criteria 1 or 3 for SPS (see SPS-1) with at least some adenomas or personal history of between 11–20 cumulative adenomas[b]

RISK STATUS

- Pathogenic variant(s) known
- No known pathogenic variants in any polyposis gene[c]

TESTING STRATEGY

- Pathogenic variant(s) known → Genetic testing for familial pathogenic variant
- No known pathogenic variants in any polyposis gene[c] → Germline multi-gene testing (preferred)[d] (See GENE-1) or Polyposis syndrome-specific testing (*APC* and/or *MUTYH*)[e]

RESULTS → TREATMENT/SURVEILLANCE

Genetic testing for familial pathogenic variant:

- Positive for familial *APC* pathogenic variant → To determine classical FAP vs. AFAP, see FAP/AFAP-1
- Positive for biallelic *MUTYH* pathogenic variant → See MAP-1
- Positive for known familial pathogenic variant in another polyposis gene → See GENE-7
- Genetic testing not done → Manage as if positive for the known familial pathogenic variant
- Negative for familial pathogenic variant → See NCCN Guidelines for Colorectal Cancer Screening- Additional testing may be indicated based on personal and family medical history
- One familal *MUTYH* pathogenic variant found[f] → See GENE-7

Germline multi-gene testing (preferred)[d] (See GENE-1) or Polyposis syndrome-specific testing (*APC* and/or *MUTYH*)[e]:

- Positive for familial *APC* pathogenic variant → To determine classical FAP vs. AFAP, see FAP/AFAP-1
- Positive for biallelic *MUTYH* pathogenic variant → See MAP-1
- Positive for pathogenic variant in another polyposis gene → See GENE-7
- No pathogenic variant found in any polyposis gene[f] → Tailored surveillance based on individual and family risk assessment (See CPUE-1 or See NCCN Guidelines for Colorectal Cancer Screening- Average risk)
- Genetic testing not done → See CPUE-1
- One *MUTYH* pathogenic variant found[f] → See GENE-7

[a] Irrespective of degree of relatedness.
[b] Age of onset, family history, personal history of colorectal cancer, and/or presence of other features may influence whether genetic testing is offered in these situations.
[c] There are clinically relevant yet rarer genes that can cause a polyposis that may be phenotypically indistinguishable from APC/MUTYH polyposis.
[d] Multi-gene panel should include all polyposis and colorectal cancer genes (Stanich P, et al. Clin Gastroenterol Hepatol 2018).
[e] When colonic polyposis is present in a single person with a negative family history, consider testing for a de novo APC pathogenic variant; if negative, follow with testing of MUTYH (targeted testing for the two common northern European founder pathogenic variants c.536A>G and c.1187G>A may be considered first followed by full sequencing if biallelic pathogenic variants are not found). When colonic polyposis is present only in siblings, consider recessive inheritance and test for MUTYH first. Order of testing for APC and MUTYH is at the discretion of the clinician. MUTYH genetic testing is not indicated based on a personal history of a hepatoblastoma, cribriform-morular variant of papillary thyroid cancer, or multifocal/bilateral CHRPE.
[f] Siblings of a patient with MAP are recommended to have site-specific testing for the familial pathogenic variants. Full sequencing of MUTYH may be considered in an unaffected parent when the other parent has MAP. If the unaffected parent is found to have one MUTYH pathogenic variant, testing the children for the familial MUTYH pathogenic variants is indicated. If the unaffected parent is not tested, comprehensive testing of MUTYH should be considered in the children. Testing for children of MUTYH heterozygotes should be offered if the other parent is also a heterozygote or could still be offered if the other parent is not a heterozygote and management would change (if they have an FDR affected with CRC) or inform reproductive risks (since their future children could be at-risk for MAP).

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

National Comprehensive Cancer Network®

NCCN Guidelines Version 3.2019
Familial Adenomatous Polyposis/AFAP

NCCN Guidelines Index
Table of Contents
Discussion

PHENOTYPE	RISK STATUS	
Classical FAP:[a] • Germline *APC* pathogenic variant • Presence of ≥100 cumulative adenomas[b] (sufficient for clinical suspicion of FAP) or fewer polyps at younger ages, especially in a family known to have FAP • Autosomal dominant inheritance[c] (except with de novo pathogenic variant) • Possible associated additional findings ▸ Congenital hypertrophy of retinal pigment epithelium (CHRPE) ▸ Osteomas, supernumerary teeth, odontomas ▸ Desmoids, epidermoid cysts ▸ Duodenal and other small bowel adenomas ▸ Gastric fundic gland polyps • Increased risk for medulloblastoma, papillary carcinoma of the thyroid (<2%), and hepatoblastoma (1%–2%, usually age ≤5 y) • Pancreatic cancers (<1%) • Gastric cancers (0.5%–1.3%) • Duodenal cancers (4%–12%)	Personal history of classical FAP	See Treatment and Surveillance (FAP-1)
	Family history of classical FAP, unaffected (no symptoms, findings, adenomas), family pathogenic variant known	See Genetic Testing and Surveillance (FAP-4)
AFAP:[d] • Germline *APC* pathogenic variant • Presence of 10–<100 cumulative adenomas (average of 30 polyps) • Frequent right-sided distribution of polyps • Adenomas and cancers at age older than classical FAP (mean age of cancer diagnosis >50 y) • Upper GI findings, thyroid and duodenal cancer risks are similar to classical FAP • Other extraintestinal manifestations, including CHRPE and desmoids, are unusual	Personal history of AFAP	See Treatment and Surveillance (AFAP-1)
	Family history of AFAP, unaffected (no symptoms, findings, adenomas), family pathogenic variant known	See Genetic Testing and Surveillance (AFAP-2)

[a] A clinical diagnosis of classical FAP is suspected when >100 polyps are present at a young age; however, genetic testing of *APC* and *MUTYH* is important to differentiate FAP from MAP or colonic polyposis of unknown etiology. Identification of a germline *APC* pathogenic variant confirms the diagnosis of FAP.
[b] Individuals with >100 polyps occurring at older ages (35–40 years or older) may be found to have AFAP.
[c] There is a 30% spontaneous new pathogenic variant rate; thus, family history may be negative. This is especially noteworthy if onset age <50 y.
[d] There is currently no consensus on what constitutes a clinical diagnosis of AFAP. AFAP is considered when >10–<100 adenomas are present and is confirmed when an *APC* pathogenic variant is identified. Genetic testing of *APC* and *MUTYH* is important to differentiate AFAP from MAP or colonic polyposis of unknown etiology.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

CLASSICAL FAP TREATMENT AND SURVEILLANCE: PERSONAL HISTORY

	TREATMENT	SURVEILLANCE[d,e] (POSTCOLECTOMY)	
Personal history of classical FAP	→ **Proctocolectomy or colectomy[a,b,c]**	→ **Colon cancer:** • **If patient had colectomy with ileorectal anastomosis, then endoscopic evaluation of the rectum every 6–12 mo depending on polyp burden.** • **If patient had total proctocolectomy (TPC) with ileal pouch-anal anastomosis (IPAA), then endoscopic evaluations of the ileal pouch every 1–3 y depending on polyp burden. Surveillance frequency should be increased to every 6 mo for large, flat polyps with villous histology and/or high-grade dysplasia.** • **If patient had an ileostomy, consider careful visualization and stoma inspection by ileoscopy to evaluate for polyps or malignancy every 1–3 years; evidence to support this recommendation is limited.** • **The use of chemoprevention is to facilitate management of the remaining rectum post-surgery. There are no FDA-approved medications for this indication at present. While there are data to suggest that sulindac is the most potent polyp regression medication, it is not known if the decrease in polyp burden decreases cancer risk.** **Extracolonic surveillance (See FAP-2)**	→ **In retained rectosigmoid, colectomy if dense polyposis or severe dysplasia** **If cancer found, see appropriate NCCN Guidelines for Treatment of Cancer by Site**

[a] *APC* genetic testing is recommended in a proband to confirm a diagnosis of FAP and allow for pathogenic variant-specific testing in other family members. Additionally, knowing the location of the pathogenic variant in the *APC* gene can be helpful for predicting severity of polyposis, rectal involvement, and desmoid tumors.
[b] See Surgical Options for Treating the Colon and Rectum in Patients with FAP (FAP-A).
[c] Timing of proctocolectomy in patients <18 y of age is not established since colon cancer is rare before age 18. In patients <18 y without severe polyposis and without family history of early cancer or severe genotype, the timing of proctocolectomy can be individualized. An annual colonoscopy is recommended if surgery is delayed.
[d] It is recommended that patients be managed by physicians or centers with expertise in FAP and that management be individualized to account for genotype, phenotype, and personal considerations.
[e] Other than colon cancer, screening recommendations are expert opinion rather than evidence-based.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

CLASSICAL FAP SURVEILLANCE: PERSONAL HISTORY

SURVEILLANCE[f,g] (POSTCOLECTOMY)

Extracolonic:

- **Duodenal or periampullary cancer: Upper endoscopy[h] (including complete visualization of the ampulla of Vater) starting at around age 20–25 y. Consider baseline upper endoscopy earlier, if colectomy before age 20 y.**
- **Gastric cancer: Fundic gland polyps occur in a majority of FAP patients, and focal low-grade dysplasia can occur but is typically non-progressive. The risk of gastric cancer in FAP patients appears to be increased in patients from geographic areas with high gastric cancer risk and may be elevated in the setting of certain endoscopic findings, including carpeting of fundic gland polyps, solitary polyps larger than 20 mm, and mounds of polyps. High-risk histologic features include tubular adenomas, polyps with high-grade dysplasia, and pyloric gland adenomas. Need for specialized surveillance or surgery should be considered in presence of high risk histologic features, preferably at a center of expertise.**
  - **Patients with high-risk lesions that cannot be removed endoscopically should be referred to a specialized center for consideration of gastrectomy.**
- **Thyroid cancer: Annual thyroid examination, starting in late teenage years. Annual thyroid ultrasound may be considered, though high-level evidence to support this recommendation is lacking.**
- **CNS cancer: An annual physical examination; due to limited data, no additional screening recommendation is possible at this time.**
- **Intra-abdominal desmoids: Annual abdominal palpation. If family history of symptomatic desmoids, consider abdominal MRI with and without contrast or CT with contrast within 1–3 y post-colectomy and then every 5–10 y. Suggestive abdominal symptoms should prompt immediate abdominal imaging. Data to support screening and treatment are limited.**
- **Small bowel polyps and cancer: High-level evidence to support routine small bowel screening distal to the duodenum is lacking. Consider adding small bowel visualization to CT or MRI for desmoids as outlined above, especially if duodenal polyposis is advanced.**
- **Hepatoblastoma: No recommendations have been made for FAP; however, there are other situations where the high risk for hepatoblastoma has been observed and the following recommendations have been considered:**
  - **Liver palpation, abdominal ultrasound, and measurement of AFP every 3–6 mo during the first 5 y of life. Screening in a clinical trial is preferred.**
- **Pancreatic cancer: Due to limited data, no screening recommendation is possible at this time.**

See Duodenoscopic Findings (FAP-3)

[f] It is recommended that patients be managed by physicians or centers with expertise in FAP and that management be individualized to account for genotype, phenotype, and personal considerations.

[g] Other than colon cancer, screening recommendations are expert opinion rather than evidence-based.

[h] Cap-assisted endoscopy may be adequate for visualization of the ampulla. (Kallenberg F, et al. Endoscopy 2017;49:181-185.)

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

DUODENOSCOPIC FINDINGS	SURVEILLANCE[i]
Stage 0, No polyposis	**Repeat endoscopy every 4 y**
Stage I, Minimal polyposis (1–4 tubular adenomas, size 1–4 mm)	**Repeat endoscopy every 2–3 y**
Stage II, Mild polyposis (5–19 tubular adenomas, size 5–9 mm)	**Repeat endoscopy every 1–3 y**
Stage III, Moderate polyposis (≥20 lesions, or size ≥1 cm)	**Repeat endoscopy every 6–12 mo**
Stage IV, Dense polyposis or high-grade dysplasia	**• Surgical evaluation • Expert surveillance every 3–6 mo • Complete mucosectomy or duodenectomy, or Whipple procedure if duodenal papilla is involved[j]**

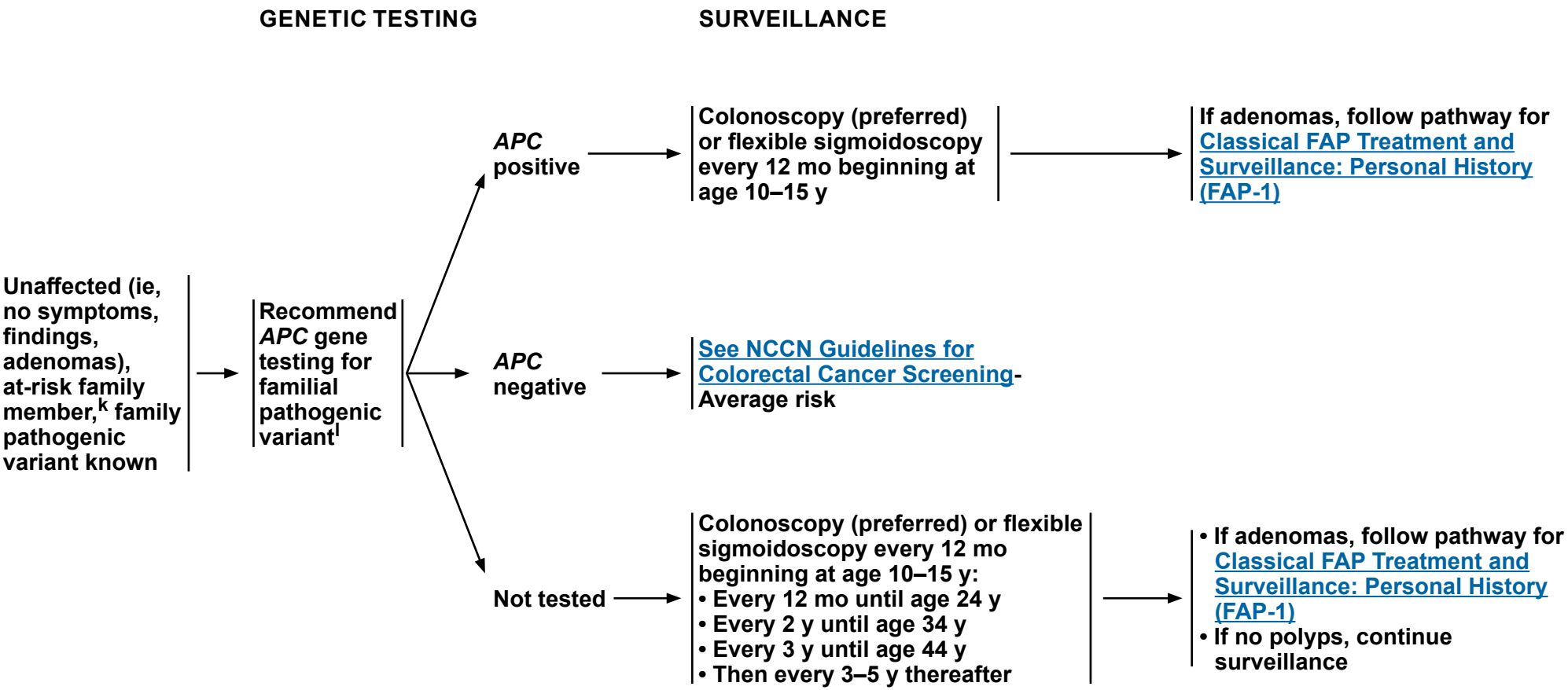
[i] Duodenal Surveillance:
It is recommended that patients be managed by physicians or centers with expertise in FAP and that management be individualized to account for genotype, phenotype, and personal considerations, including potential risks and benefits. Management that includes endoscopic treatment may require shorter intervals.
Recommend examination with side-viewing endoscope and use of Spigelman's or other standardized staging. More intensive surveillance and/or treatment is required in patients with large or villous adenomas, and with advancing age >50 y. Surgical counseling is advisable for patients with stage IV polyposis. (Spigelman AD, Williams CB, Talbot IC, et al. Upper gastrointestinal cancer in patients with familial adenomatous polyposis. Lancet 1989;2:783-785).
Endoscopic treatment options include endoscopic papillectomy in addition to excision or ablation of resectable large (>1 cm) or villous adenomas, as well as mucosectomy of resectable advanced lesions, including contained high-grade dysplasia, to potentially avert surgery while observing pathology guidelines for adequate resection.
Surgery is recommended for invasive carcinoma as well as for dense polyposis or high-grade dysplasia that cannot be managed endoscopically.
[j] Potentially higher risk adenomas involving the papilla (ie, large ≥1 cm adenomas or adenomas extending into papilla) should be referred to an expert center for evaluation and management.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

CLASSICAL FAP GENETIC TESTING AND SURVEILLANCE: FAMILY HISTORY OF CLASSICAL FAP PATHOGENIC VARIANT KNOWN



[k] If a first-degree relative is unavailable or unwilling to be tested, more distant relatives should be offered testing for the known pathogenic variant in the family.
[l] FAP genetic testing in children should be done by age 10 y when colon screening would be initiated. If there is intent to do hepatoblastoma screening, FAP genetic testing should be considered in infancy.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

SURGICAL OPTIONS FOR TREATING THE COLON AND RECTUM IN PATIENTS WITH FAP[a]

TAC/IRA is generally recommended for AFAP and TPC/IPAA is generally recommended for FAP.[b]

TOTAL ABDOMINAL COLECTOMY WITH ILEORECTAL ANASTOMOSIS (TAC/IRA)

- Indications:
  - The decision to remove the rectum is dependent on whether the polyps are amenable to endoscopic surveillance and resection.
- Contraindications:
  - Severe rectal disease (size or number of polyps)
  - Patient not reliable for follow-up surveillance of retained rectum
- Advantages:
  - Technically straightforward
  - Relatively low complication rate
  - Good functional outcome
  - No permanent or temporary stoma
  - Avoids the risks of sexual or bladder dysfunction and decreased fecundity that can occur following proctectomy
- Disadvantages
  - Risk of metachronous cancer in the remaining rectum

TOTAL PROCTOCOLECTOMY WITH END ILEOSTOMY (TPC/EI)

- Indications:
  - Very low, advanced rectal cancer
  - Inability to perform IPAA
  - Patient with IPAA with unacceptable function
  - Patient with a contraindication to IPAA
- Advantages:
  - Removes risk of CRC
  - One operation
- Disadvantages:
  - Risks of sexual or bladder dysfunction
  - Permanent stoma
  - May discourage family members from seeking evaluation for fear of permanent stoma

TOTAL PROCTOCOLECTOMY WITH ILEAL POUCH-ANAL ANASTOMOSIS (TPC/IPAA)

- Indications:
  - Severe disease in colon and/or rectum
  - After TAC/IRA with unstable rectum
  - Curable rectal cancer
  - Patient unreliable for follow-up after TAC/IRA
- Contraindications:
  - Intra-abdominal desmoid that would interfere with completion of surgery
  - Patient is not a candidate for IPAA (eg, concomitant Crohn's disease, anal sphincter dysfunction)
- Advantages:
  - Minimal risk of rectal cancer
  - No permanent stoma
  - Reasonable bowel function
- Disadvantages:
  - Complex operation
  - Usually involves temporary stoma
  - Risks of sexual or bladder dysfunction
  - Functional results are variable

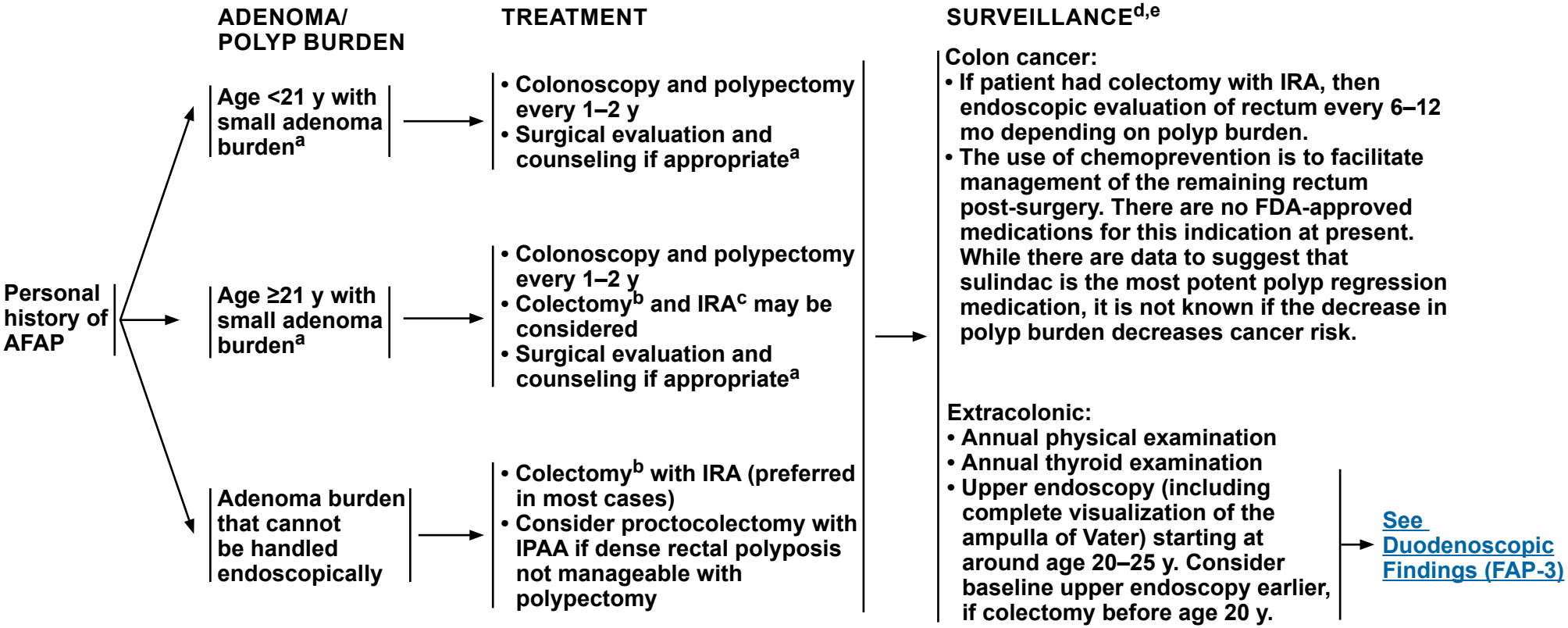
[a] It is recommended that patients be managed by physicians or centers with expertise in FAP and that management be individualized to account for genotype, phenotype, and personal considerations.
[b] In certain circumstances, such as AFAP with mainly proximal polyps, the extent of colectomy may be modified based on the burden of adenoma distribution and number.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

ATTENUATED FAP TREATMENT AND SURVEILLANCE: PERSONAL HISTORY



[a] Small adenoma burden is defined (somewhat arbitrarily) as fewer than 20 adenomas, all <1 cm in diameter, and none with advanced histology, so that colonoscopy with polypectomy can be used to effectively eliminate the polyps. Colectomy may be indicated before this level of polyp burden, especially if colonoscopy is difficult and polyp control is uncertain. Surgery could be considered when polyp burden is >20 at any individual examination, when polyps have been previously ablated, when some polyps have reached a size >1 cm, or when advanced histology is encountered in any polyp.

[b] See Surgical Options for Treating the Colon and Rectum in Patients with FAP (FAP-A).

[c] Earlier surgical intervention should be considered in noncompliant patients.

[d] It is recommended that patients be managed by physicians or centers with expertise in FAP/AFAP and that management be individualized to account for genotype, phenotype, and personal considerations.

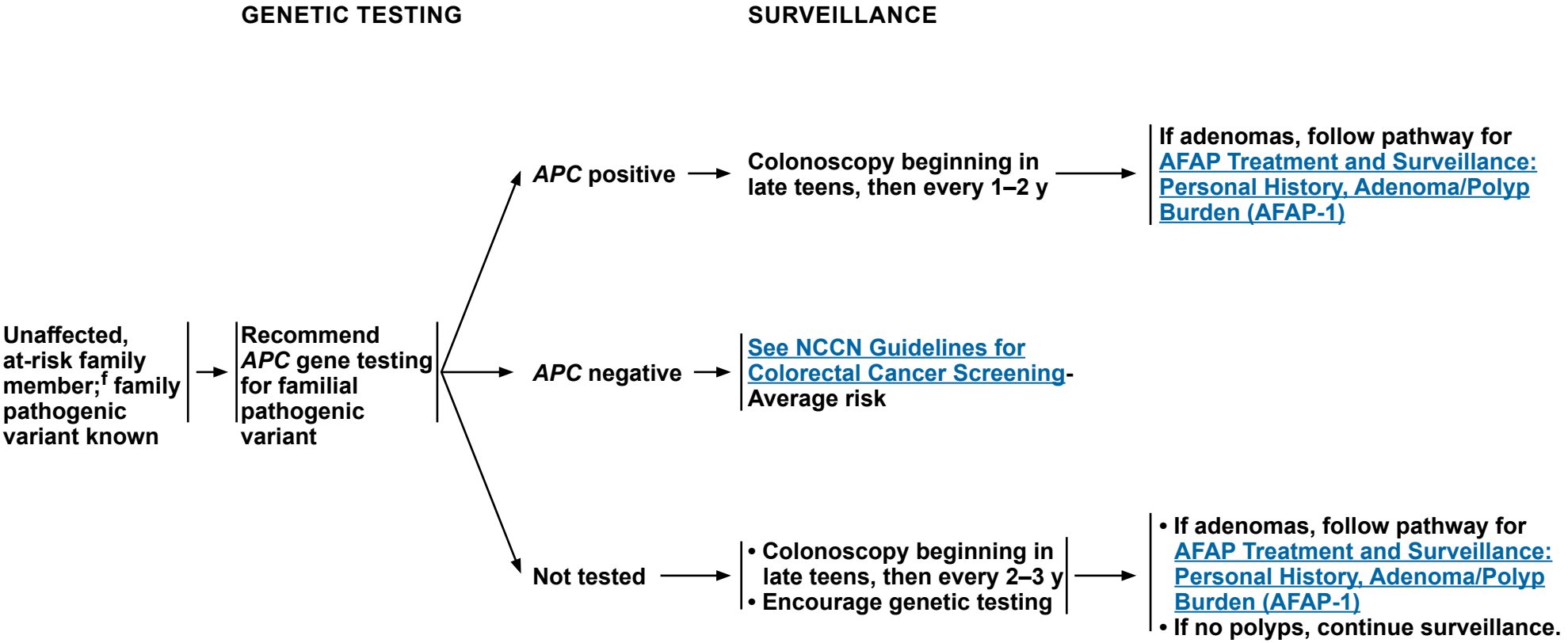
[e] Surveillance for upper GI findings for AFAP is similar to classical FAP.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

ATTENUATED FAP GENETIC TESTING AND SURVEILLANCE: FAMILY HISTORY OF ATTENUATED FAP PATHOGENIC VARIANT KNOWN



[f] If a first-degree relative is unavailable or unwilling to be tested, more distant relatives should be offered testing for the known pathogenic variant in the family.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

NCCN Guidelines Version 3.2019
MUTYH-Associated Polyposis

NCCN Guidelines Index
Table of Contents
Discussion

PHENOTYPE

- **Biallelic *MUTYH* pathogenic variants**
- **Polyposis or colon cancers consistent with autosomal recessive inheritance (ie, parents unaffected, siblings affected)**
- **Consanguinity**
- **Fewer than 100 adenomas[a] (range 0–100s and uncommonly >1000)**
- **Adenomas and CRC at age older than classical FAP (median CRC age >50 y)**
- **Duodenal cancer (5%)**
- **Duodenal polyps**

RISK STATUS

Personal history of MAP → **See Treatment and Surveillance (MAP-2)**

Unaffected, at-risk family member; family pathogenic variant known → **See Genetic Testing and Surveillance (MAP-3)**

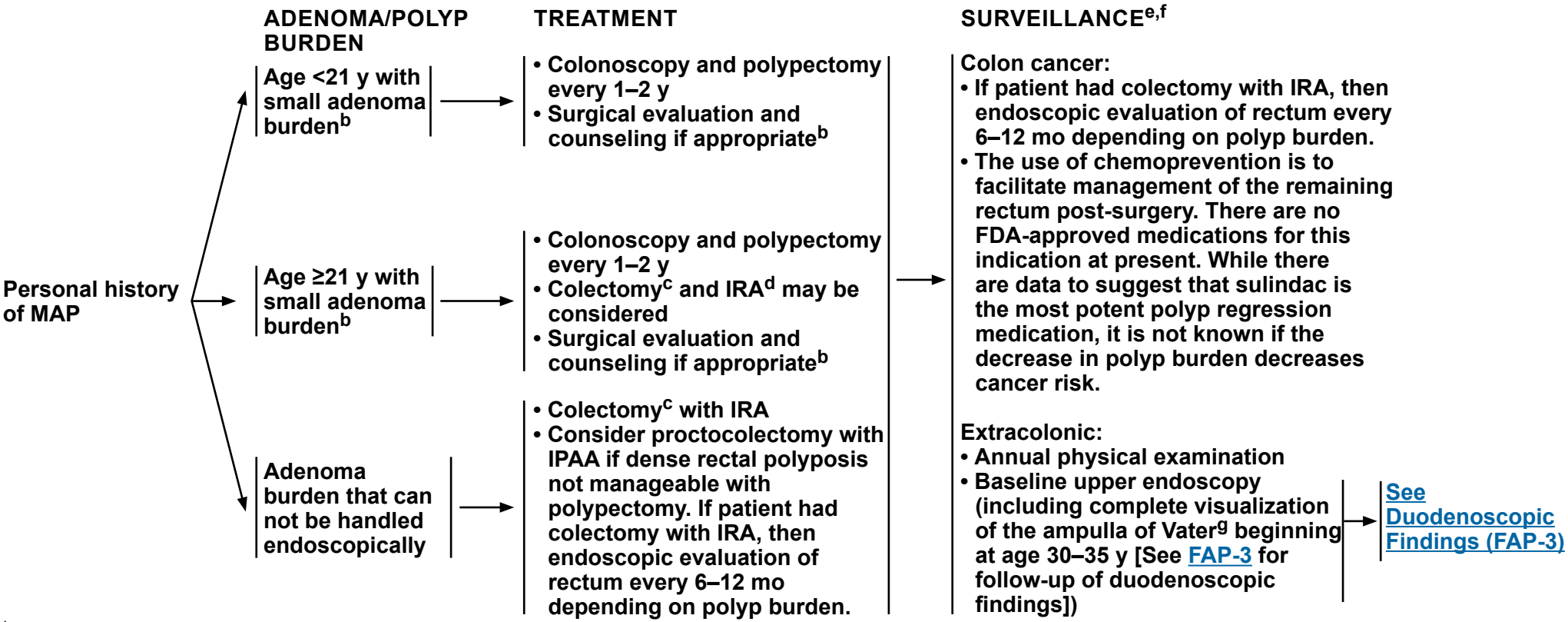
[a] Multiple serrated polyps (hyperplastic polyps, sessile serrated polyps, and traditional serrated adenomas) may also be seen in patients with MAP polyposis. Patient with MAP may also meet criteria for SPS.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MAP TREATMENT AND SURVEILLANCE: PERSONAL HISTORY



[b] Small adenoma burden is defined (somewhat arbitrarily) as fewer than 20 adenomas, all <1 cm in diameter, and none with advanced histology, so that colonoscopy with polypectomy can be used to effectively eliminate the polyps. Colectomy may be indicated before this level of polyp burden, especially if colonoscopy is difficult and polyp control is uncertain. Surgery could be considered when polyp burden is >20 at any individual examination, when polyps have been previously ablated, when some polyps have reached a size >1 cm, or when advanced histology is encountered in any polyp. Extent of colectomy may be modified based on the burden and distribution of adenomas.

[c] See Surgical Options for Treating the Colon and Rectum in Patients with FAP (FAP-A).

[d] Earlier surgical intervention should be considered in noncompliant patients.

[e] It is recommended that patients be managed by physicians or centers with expertise in MAP and that management be individualized to account for genotype, phenotype, and personal considerations.

[f] Surveillance for upper GI findings for MAP is similar to classical FAP.

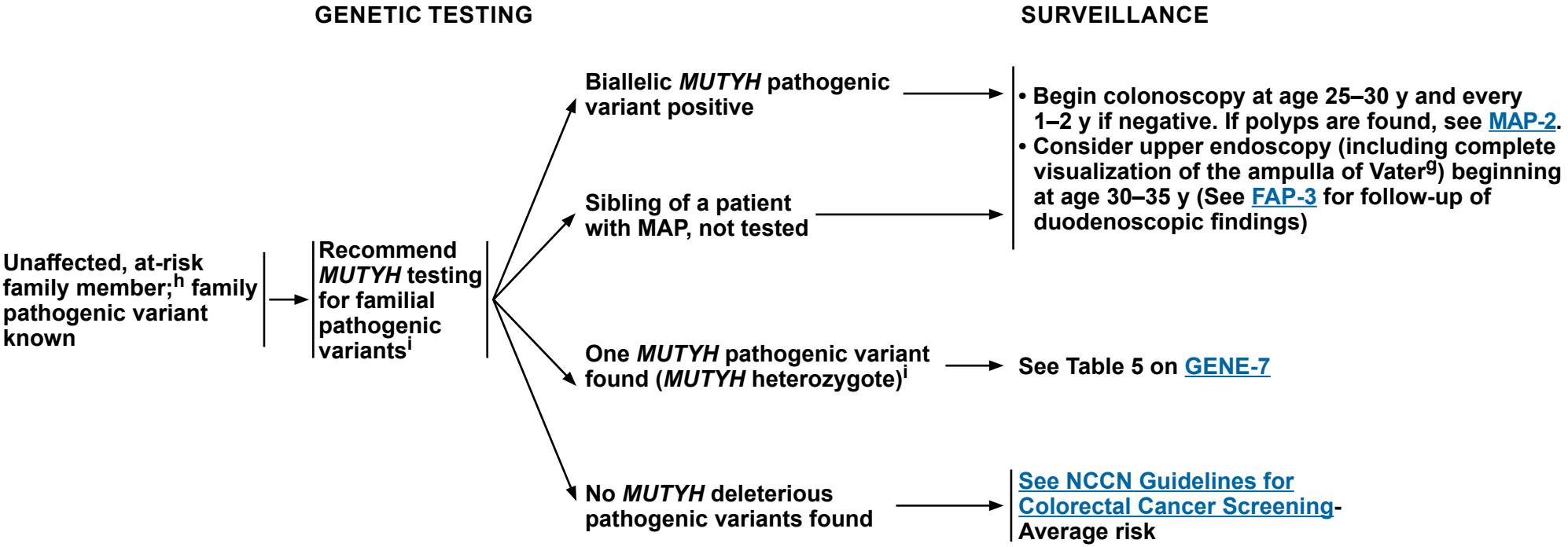
[g] Cap-assisted endoscopy may be adequate for visualization of the ampulla. (Kallenberg F, et al. Endoscopy 2017;49:181-185.)

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MAP TREATMENT AND SURVEILLANCE: FAMILY HISTORY OF MAP PATHOGENIC VARIANT KNOWN



[g] Cap-assisted endoscopy may be adequate for visualization of the ampulla. (Kallenberg F, et al. Endoscopy 2017;49:181-185.)
[h] An at-risk family member can be defined as a sibling of an affected individual and/or proband. Other individuals in a family may also be at risk of having MAP or a monoallelic *MUTYH* pathogenic variant.
[i] Siblings of a patient with MAP are recommended to have site-specific testing for the familial pathogenic variants. Full sequencing of *MUTYH* may be considered in an unaffected parent when the other parent has MAP. If the unaffected parent is found to not have a *MUTYH* pathogenic variant, genetic testing in the children is not necessary to determine MAP status. If the unaffected parent is not tested, comprehensive testing of *MUTYH* should be considered in the children. If the unaffected parent is found to have one *MUTYH* pathogenic variant, testing the children for the familial *MUTYH* pathogenic variants is indicated.
[j] There are no specific data available to determine screening recommendations for a patient with a heterozygous *MUTYH* pathogenic variant and a second-degree relative affected with CRC. See the NCCN Guidelines for Colorectal Cancer Screening.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

PJS diagnosis:[a,b]

- **The majority of cases occur due to pathogenic variants in the *STK11 (LKB1)* gene. Clinical genetic testing is available.**
- **A clinical diagnosis of PJS can be made when an individual has two or more of the following features:**
  - **Two or more Peutz-Jeghers-type hamartomatous polyps of the GI tract**
  - **Mucocutaneous hyperpigmentation of the mouth, lips, nose, eyes, genitalia, or fingers**
  - **Family history of PJS**

Surveillance considerations:

- **Referral to a specialized team is recommended and participation in clinical trials is especially encouraged.**
- **Surveillance should begin at the approximate ages on PJS-2 if symptoms have not already occurred, and any early symptoms should be evaluated thoroughly.**
- **The surveillance guidelines (See PJS-2) for the multiple organs at risk for cancer are provisional, but may be considered in view of the cancer risks in PJS and the known utility of the tests. There are limited data regarding the efficacy of various screening modalities in PJS.**

See Cancer Risk and Surveillance Guidelines (PJS-2)

[a] Tomlinson IP, Houlston RS. Peutz-Jeghers syndrome. J Med Genet 1997;34:1007-1011.
[b] Due to the rarity of the syndrome and complexities of diagnosing and managing individuals with PJS, referral to a specialized team or centers with expertise is recommended.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Peutz-Jeghers Syndrome: Cancer Risk and Surveillance Guidelines

Site	% Lifetime Risk[c]	Screening Procedure and Interval	Initiation Age (y)
Breast (women)	32%–54%	• Mammogram and breast MRI annually[d] • Clinical breast exam every 6 mo	~ 25 y
Colon	39%	• Colonoscopy every 2–3 y	~ Late teens
Stomach	29%	• Upper endoscopy every 2–3 y	~ Late teens
Small intestine	13%	• Small bowel visualization (CT or MRI enterography or video capsule endoscopy baseline at age 8–10 y with follow-up interval based on findings but at least by age 18 y, then every 2–3 y, though this may be individualized. Repeat small intestinal exam is also indicated at any time based on symptoms.)	~ 8–10 y
Pancreas	11%–36%	• Magnetic resonance cholangiopancreatography with contrast or endoscopic ultrasound every 1–2 y (both ideally performed at center of expertise)	~ 30–35 y[e]
Ovary (typically benign sex cord/Sertoli cell tumors) Cervix (typically cervical adenoma malignum) Uterus	18%–21% 10% 9%	• Pelvic examination and Pap smear annually	~ 18–20 y
Testes (typically sex cord/ Sertoli cell tumors)	9%	• Annual testicular exam and observation for feminizing changes	~ 10 y
Lung	7%–17%	• Provide education about symptoms and smoking cessation • No other specific recommendations have been made	

[c] Hearle N, Schumacher V, Menko FH, et al. Frequency and spectrum of cancers in the Peutz-Jeghers syndrome. Clin Cancer Res 2006;12:3209-3215; Giardiello FM, Brensinger JD, Tersmette AC, et al. Very high risk of cancer in familial Peutz-Jeghers syndrome. Gastroenterology 2000;119:1447-1453.

[d] See NCCN Guidelines for Genetic/Familial High-Risk Assessment: Breast/Ovarian (BRCA-A) for further breast screening recommendations regarding mammogram and breast MRI screening. High-quality breast MRI limitations include having: a need for a dedicated breast coil, the ability to perform biopsy under MRI guidance, experienced radiologists in breast MRI, and regional availability. Breast MRI performed preferably days 7–15 of menstrual cycle for premenopausal women. The appropriateness of imaging modalities and scheduling is still under study. Lowry KP, et al. Cancer 2012; 118:2021-2030.

[e] Based on clinical judgment, early initiation age may be considered, such as 10 y younger than the earliest age of onset in the family.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

JPS definition:[a,b]

- **A clinical diagnosis of JPS is considered in an individual who meets at least one of the following criteria:**
  - **≥5 juvenile polyps of the colon**
  - **Multiple juvenile polyps found throughout the GI tract**
  - **Any number of juvenile polyps in an individual with a family history of JPS**

Genetic testing:

- **Clinical genetic testing is recommended with approximately 50% of JPS cases occurring due to pathogenic variants in the *BMPR1A* or *SMAD4*[c] genes. If there is a known *SMAD4* pathogenic variant in the family, genetic testing should be performed within the first 6 months of life due to hereditary hemorrhagic telangiectasia (HHT) risk.**

Surveillance considerations:

- **Referral to a specialized team is recommended and participation in clinical trials is especially encouraged.**
- **Surveillance should begin at the approximate ages listed below, if symptoms have not already occurred. Any early symptoms should be evaluated thoroughly.**
- **The following surveillance guidelines for the multiple organs at risk for cancer may be considered. Limited data exist regarding the efficacy of various screening modalities in JPS.**

Juvenile Polyposis Syndrome: Risk and Surveillance Guidelines

Site	% Lifetime Risk	Screening/Surveillance Procedure and Interval	Initiation Age (y)
Colon	40%–50%	Colonoscopy: repeat annually if polyps are found and if no polyps, repeat every 2–3 years[d]	~ 15 y
Stomach	21% if multiple polyps	Upper endoscopy: repeat annually if polyps are found and if no polyps, repeat every 2–3 years[d,e]	~ 15 y
Small intestine	Rare, undefined	No recommendations have been made	
Pancreas	Rare, undefined	No recommendations have been made	
HHT	Undefined	In individuals with *SMAD4* pathogenic variants, screen for vascular lesions associated with HHT[c]	Within first 6 mo of life

[a] Due to the rarity of the syndrome and complexities of diagnosing and managing individuals with JPS, referral to a specialized team is recommended.

[b] Syngal S, Brand R, Church J, et al. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. Am J Gastroenterol 2015;110:223-262.

[c] Faughnan M, Palda V, Garcia-Tsao G, et al. HHT Foundation International - Guidelines Working Group. International guidelines for the diagnosis and management of hereditary haemorrhagic telangiectasia. J Med Genet 2011;48:73-87.

[d] In families without an identified genetic pathogenic variant, consider substituting endoscopy every 5 y beginning at age 20 and every 10 years beginning at age 40 y in patients in whom no polyps are found.

[e] There may be management issues related to anemia from giant confluent polyps. If anemia develops requiring blood transfusion due to many stomach polyps, gastrectomy can be considered in severe cases.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

NCCN Guidelines Version 3.2019
Serrated Polyposis Syndrome

NCCN Guidelines Index
Table of Contents
Discussion

Serrated polyposis syndrome (previously known as hyperplastic polyposis) definition:[a,b,c]

- **A clinical diagnosis of serrated polyposis is considered in an individual who meets at least one of the following empiric criteria:[d]**
  - **1) At least 5 serrated polyps[e] proximal to the sigmoid colon with 2 or more of these being >10 mm**
  - **2) Any number of serrated polyps[e] proximal to the sigmoid colon in an individual who has a first-degree relative with serrated polyposis**
  - **3) ≥20 serrated polyps of any size, but distributed throughout the colon[f]**
- **Occasionally, more than one affected case of serrated polyposis is seen in a family.[g]**
- **For the majority of patients with SPS, no causative gene is identifiable. Pathogenic variants in *RNF43* have been identified as a rare cause of serrated polyposis.**
- **The risk for colon cancer in this syndrome is elevated, although the precise risk remains to be defined.**

Surveillance recommendations for individuals with serrated polyposis:

- **Colonoscopy with polypectomy until all polyps ≥5 mm are removed, then colonoscopy every 1 to 3 years depending on number and size of polyps. Clearing of all polyps is preferable but not always possible.**
- **Consider surgical referral if colonoscopic treatment and/or surveillance is inadequate or if high-grade dysplasia occurs.**

Surveillance recommendations for individuals with a family history of serrated polyposis:

- **The risk of CRC in first-degree relatives of individuals with serrated polyposis is elevated.**
- **First-degree relatives are encouraged to have colonoscopy at the earliest of the following:**
  - **Aged 40 years**
  - **Same age as youngest diagnosis of serrated polyposis if uncomplicated by cancer**
  - **Ten years earlier than earliest diagnosis in family of CRC complicating serrated polyposis**
- **Following baseline exam, repeat every 5 years if no polyps are found. If proximal serrated polyps or multiple adenomas are found, consider colonoscopy every 1–3 years.**

[a] The Serrated Polyposis Syndrome Guidelines are based on expert opinion on the current data available.

[b] Snover DC, Ahnen DJ, Burt RW, Odze RD. Serrated polyps of the colon and rectum and serrated polyposis. In: Bosman FT, Carneiro F, Hruban RH, Theise ND, eds. WHO Classification of Tumours of the Digestive System: LYON: IARC, 2010:160-165.

[c] The final classification of SPS awaits more definitive genetic/epigenetic molecular characterization. These lesions are considered premalignant. Until more data are available, it is recommended that they be managed similarly to adenomas.

[d] There may be other clinical senarios (eg, patient has between 5–10 serrated polyps, <1 cm) that increase colon cancer risk and may require additional evaluation per clinical judgment (Egoavil C, Juárez M, Guarinos C, et al. Increased risk of colorectal cancer in patients with multiple serrated polyps and their first-degree relatives. Gastroenterology 2017;153:106-112.)

[e] Serrated polyps include hyperplastic polyps, sessile serrated adenomas/polyps, and traditional serrated adenomas.

[f] Multiple hyperplastic polyps localized to the rectum and sigmoid are unlikely to contribute to SPS. Such distal polyps should not be counted toward the “qualifying” burden unless they a) are >10 mm; or b) display additional characteristics of serrated polyps (serrations extending to base of crypt, with widened or “boot”-shaped crypt base).

[g] Boparai KS, Reitsma JB, Lemmens V, et al. Increased colorectal cancer risk in first-degree relatives of patients with hyperplastic polyposis syndrome. Gut 2010;59:1222-1225.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

COLONIC ADENOMATOUS POLYPOSIS OF UNKNOWN ETIOLOGY

The following are surveillance/management recommendations for colonic adenomatous polyposis without a known pathogenic variant in a polyposis gene.[a]

Phenotype	Management/Surveillance
Personal history of ≥100 adenomas	Manage as FAP (See FAP-1)
Personal history of >20–<100 adenomas: Small adenoma burden manageable by colonoscopy and polypectomy	• Colonoscopy and polypectomy every 1–2 years ▸ Clearing of all polyps is recommended. Repeat at short interval if residual polyps are present. • Consider baseline upper endoscopy (including complete visualization of the ampulla of Vater[d]) at baseline and repeat following duodenal surveillance guidelines on page FAP-3
Personal history of >20–<100 adenomas: Dense polyposis or large polyps not manageable by polypectomy	• Subtotal colectomy • Consider proctocolectomy if there is dense rectal polyposis not manageable by polypectomy. • Consider baseline upper endoscopy (including complete visualization of the ampulla of Vater[d]) at baseline and repeat following duodenal surveillance guidelines on page FAP-3
Personal history of 11–20 adenomas	Manage based on clinical judgment
Family history of ≥100 adenomas in a first-degree relative[b,c] at age <40 y	• Consider colonoscopy beginning at age 10–15 y ▸ then every 1 y until age 24 y, ▸ every 2 y from 24–34 y, ▸ every 3 y from 34–44 y, ▸ then every 3–5 y thereafter • If polyposis is detected, follow pathway for Classical FAP Treatment and Surveillance: Personal History (See FAP-1)
Family history of >20–<100 adenomas in a first-degree relative[b,c]	Consider colonoscopy and polypectomy every 3–5 y[e] starting at the same age as the youngest diagnosis of polyposis in the family if uncomplicated by cancer or by age 40, whichever is earliest
Family history of ≥100 adenomas diagnosed at age ≥40 in a first-degree relative[b,c]	Consider colonoscopy and polypectomy every 2–3 y[e] starting at age 40 y if uncomplicated by cancer

[a] Gene mutations associated with adenomatous polyposis include, but are not limited to monoallelic mutations in APC, GREM1, POLE, POLD1, and AXIN2, and biallelic mutations in MUTYH, NTHL1, and MSH3.
[b] Recommend genetic testing (See POLYP-1) in family member affected with polyposis.
[c] There are limited data to suggest definitive recommendations for when to initiate screening or the interval of screening.
[d] Cap-assisted endoscopy may be adequate for visualization of the ampulla. (Kallenberg F, et al. Endoscopy 2017;49:181-185.)
[e] If multiple polyps are found, then colonoscopy every 1–3 years depending on type, number, and size of polyps.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MULTI-GENE TESTING

Overview

- **The introduction of multi-gene testing for hereditary forms of cancer has rapidly altered the clinical approach to testing at-risk patients and their families. Based on next-generation sequencing technology, these tests simultaneously analyze a set of genes that are associated with a specific family cancer phenotype or multiple phenotypes. Given relative novelty of multi-gene testing, terminology and associated definitions used in this section of the guidelines are outlined in Table 1. Pros and cons of multi-gene testing are outlined in Table 2, and Table 3 provides examples of clinical scenarios for which multi-gene testing may be considered. Table 4 provides a list of genes that may be found on commercially available multi-gene panels with the strength of evidence, risk level, and phenotypic association, and Table 5 provides current recommendations for surveillance, based on gene pathogenic variant type.**
- **When more than one gene can explain an inherited cancer syndrome, then multi-gene testing may be more efficient and/or cost-effective than single-gene testing.**
- **There is also a role for multi-gene testing in individuals who have tested negative (indeterminate) for a single syndrome, but whose personal or family history remains strongly suggestive of an inherited susceptibility.**
- **When multi-gene testing is performed, there is an increased likelihood of finding variants of uncertain significance.**
- **Chances of finding a variant of uncertain significance or pathogenic variant with uncertain clinical management increase as the number of genes included in the multi-gene panel increase.**
  - **Reclassification of variants of uncertain significance is commonplace.[a,b] Historically, over 91% of variants of uncertain significance in hereditary cancer testing have been downgraded to benign or likely benign categories.[a,b] Nonetheless, clinical phenotypic correlation is warranted with further discussion with the testing laboratory if there is evidence supporting variant pathogenicity. Patient and provider guidelines and policies for follow-up of variants of uncertain significance have been developed.[c,d]**
- **As commercially available tests differ in the specific genes analyzed (as well as classification of variants, reclassification procedures, and many other factors), choosing the specific laboratory and test panel is important.**
- **Multi-gene testing can include "intermediate" penetrant (moderate-risk) genes. For many of these genes, there are limited data on the degree of cancer risk and there are no clear guidelines on risk management for carriers of pathogenic variants. Not all genes included on available multi-gene tests are necessarily clinically actionable.**
- **As is the case with high-risk genes, it is possible that the risks associated with moderate-risk genes may not be entirely due to that gene alone, but may be influenced by gene/gene or gene/environment interactions. In addition, certain pathogenic variants in a gene may pose higher or lower risk than other pathogenic variants in that same gene. Therefore, it may be difficult to use a known pathogenic variant alone to assign risk for relatives.**
- **In many cases the information from testing for moderate penetrance genes does not change risk management compared to that based on family history alone.**
- **It is for these and other reasons that multi-gene testing is ideally offered in the context of professional genetic expertise for pre- and post-test counseling. Individuals with the recommended expertise include certified genetic counselors, as well as clinicians who have had extensive training and/or experience in identification and management of hereditary syndromes.**

[a] Mersch J, Brown N, Pirzadeh-Miller S, et al. Prevalence of variant reclassification following hereditary cancer genetic testing. JAMA 2018;320:1266-1274.
[b] Slavin T, Van Tongeren L, Behrendt C, et al. Prospective study of cancer genetic variants: Variation in rate of reclassification by ancestry. J Natl Cancer Inst 2018;110(10):1059-1066.
[c] Slavin T, Manjarrez S, Pritchard C, et al. The effects of genomic germline variant reclassification on clinical cancer care. Oncotarget 2019;10:417-423.
[d] David K, Best R, Brenman L, et al. Patient re-contact after revision of genomic test results: points to consider-a statement of the American College of Medical Genetics and Genomics (ACMG). Genet Med 2019;21:769-771.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MULTI-GENE TESTING

Table 1: Multi-Gene Testing Definitions

TERM	DEFINITION
Multi-gene panel	Laboratory test that includes testing for pathogenic variants of more than one gene.
Syndrome-specific panel	Panel that only tests for one syndrome (eg, LS, adenomatous polyposis).
Cancer-specific panel	Panel that tests for more than one gene associated with a specific type of cancer.
"Comprehensive" cancer panel	Panel that tests for more than one gene associated with multiple cancers or multiple cancer syndromes.
Actionable pathogenic variant	Pathogenic variant that results in a recommendation for a change in clinical management.
Variant of uncertain significance	Genetic test result indicating a sequence variant in a gene that is of uncertain significance. Variants are generally not clinically actionable, and most (but not all) are ultimately re-classified as benign.[a,b]

Table 2: Pros and Cons of Multi-Gene Testing for Hereditary Colorectal Syndromes[e]

PROS	CONS
• More efficient testing when more than one gene may explain presentation and family history. • Higher chance of providing proband with possible explanation for cause of cancer. • Competitive cost relative to sequentially testing single genes.	• Higher chance of identifying pathogenic variants for which clinical management is uncertain. Estimates suggest that 3%–4% (Gastroenterology 2015 Sep;149:604-13.e20; Clin Genet 2014: 86:510–520) of pathogenic variants identified are not clearly clinically actionable, such as finding a pathogenic variant in a moderate-risk gene for which management is unclear. • Higher chance of identifying variants of uncertain significance that are not actionable; reported rates of finding variants of uncertain significance range from 17%–38%. • Higher chance that patient will mistakenly receive overtreatment and overscreening if variants of uncertain significance or pathogenic variants for which clinical management is uncertain are incorrectly interpreted.

[a] Mersch J, Brown N, Pirzadeh-Miller S, et al. Prevalence of variant reclassification following hereditary cancer genetic testing. JAMA 2018;320:1266-1274.
[b] Slavin T, Van Tongeren L, Behrendt C, et al. Prospective study of cancer genetic variants: Variation in rate of reclassification by ancestry. J Natl Cancer Inst 2018;110(10):1059-1066.
[e] Hall MJ, et al. Gene panel testing for inherited cancer risk. J Natl Compr Canc Netw 2014;12:1339-1346.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MULTI-GENE TESTING

Table 3: Examples of Clinical Scenarios for Which Multi-Gene Testing Should and Should Not Be Used

Examples of clinical scenarios for which multi-gene testing should be considered: • Personal medical and/or family cancer history meets criteria for more than one hereditary cancer syndrome (ie, family meets both *BRCA*-related breast and/or ovarian cancer and LS clinical criteria or family history of young-onset CRC and oligopolyposis) • Colonic polyposis with uncertain histology • Family cancer history does not meet established testing guidelines, but consideration of inherited cancer risk persists and an appropriate panel is available • Individuals concerned about cancer predisposition for whom family cancer history is limited or unknown • Second-line testing for inherited cancer risk when first-line testing has been inconclusive • Adenomatous polyposis
Examples of clinical scenarios for which multi-gene testing SHOULD NOT be considered:[f] • An individual from a family with a known pathogenic variant and no other reason for multi-gene testing • As first-line testing when the family history is strongly suggestive of a known hereditary syndrome

[f]Syndrome-specific panels may be appropriate.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MULTI-GENE TESTING

Table 4: Evaluation of CRC Genes Commonly Included on Multi-Gene Panels[g]

GENE	STRENGTH OF EVIDENCE	RISK LEVEL*	ASSOCIATION	REFERENCE
APC	Well-established	High	FAP & AFAP	See *APC* and *MUTYH* Genetic Testing Criteria (APC/MUTYH-1)
APC I1307K pathogenic variant	Well-established	Moderate	Increased frequency in Ashkenazi Jewish individuals; increased risk for CRC	Boursi B, et al. Eur J Cancer 2013;49:3680-3685. Liang J, et al. Am J Epidemiol 2013;177:1169-1179.
ATM	Not well-established	Unclear – moderate at most	Increased risk for breast cancer and CRC	Thompson D, et al. J Natl Cancer Inst 2005;97:813-822. Olsen JH, et al. Br J Cancer 2005;93:260-265.
AXIN2	Not well-established	Uncertain – presumed high risk from limited case reports	Polyposis and oligodontia	Lammi L, et al. Am J Hum Genet 2004;74:1043-50. Marvin ML, et al. Am J Med Genet A 2011;155A:898-902. Rivera B, et al. Eur J Hum Genet 2014;22:423-426. Lejuene S, et al. Hum Mutat 2006;27:1064. Wong S, et al. Arch Oral Biol 2014;59:349-353.
BLM heterozygotes	Not well-established	Uncertain – none to low	Possible increased risk for CRC	Cleary SP, et al. Cancer Res 2003;3:1769-1771. Baris HN, et al. Isr Med Assoc J 2007;9:847-850. Laitman Y, et al. Cancer Genet 2016;209:70-74.
BMPR1A	Well-established	High	JPS	See Juvenile Polyposis Syndrome Guidelines (JPS-1)
CHEK2	Not well-established	Moderate	Increased risk for breast, colon, and other cancers	Xiang HP, et al. Eur J Cancer 2011;47:2546-2551. Liu C, et al. Asian Pac J Cancer Prev 2012;13:2051-2055. Gronwald J, et al. Br J Cancer 2009;100:1508-1512.
EPCAM	Well-established	High	LS	See Lynch Syndrome Guidelines (LS-1)

*Risk level is based on panel consensus.

[g]*RPS20* is an emerging gene that is potentially linked to CRC, and there are not enough data at present to include *RPS20* on this list.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MULTI-GENE TESTING

Table 4: Evaluation of CRC Genes Commonly Included on Multi-Gene Panels[g] (continued)

GENE	STRENGTH OF EVIDENCE	RISK STATUS*	ASSOCIATION	REFERENCE
GALNT12	Not well-established	Uncertain – moderate at most	Increased risk for colorectal cancer	Guda K, et al. Proc Natl Acad Sci U.S.A. 2009;106:12921-12925. Clarke E, et al. Hum Mutat 2012;33:1056-1058. Segui N, et al. Hum Mutat 2014;35:50-52.
GREM1	Not well-established	Uncertain – presumed high risk from limited case reports	Hereditary mixed polyposis syndrome due to a 40kb duplication upstream of *GREM1* in Ashkenazi Jewish ancestry only	Jaeger E, et al. Nat Genet 2012; 44:699-703.
MLH1	Well-established	High	LS	See Lynch Syndrome Guidelines (LS-1)
MSH2	Well-established	High	LS	
MSH6	Well-established	High	LS	
MSH3 biallelic pathogenic variants	Not well-established	Uncertain – presumed high risk from limited case reports	Polyposis	Adam R, et al. Am J Hum Genet 2016;99:337-51.
MUTYH biallelic pathogenic variants	Well-established	High	MAP	See *APC* and *MUTYH* Genetic Testing Criteria (APC/MUTYH-1)
MUTYH heterozygotes	Not well-established	Uncertain – moderate at most	Possible increased risk for CRC	Win AK, et al. Gastroenterology 2014;146:1208-1211.

*Risk level is based on panel consensus.

[g]*RPS20* is an emerging gene that is potentially linked to CRC, and there are not enough data at present to include *RPS20* on this list.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

MULTI-GENE TESTING

Table 4: Evaluation of CRC Genes Commonly Included on Multi-Gene Panels[g] (continued)

GENE	STRENGTH OF EVIDENCE	RISK STATUS*	ASSOCIATION	REFERENCE
NTHL1 **biallelic pathogenic variants**	**Not well-established**	**Uncertain – presumed high from limited case reports**	**Polyposis**	**Weren RD, et al. Nat Genet 2015;47:668-671. Rivera B, et al. N Engl J Med 2015;373:1985-1986. Broderick P, et al. BMC Cancer 2006:6:243.**
POLD1	**Not well-established**	**Uncertain – presumed high risk from limited case reports**	**Polymerase proofreading-associated polyposis**	**Palles C, et al. Nat Genet 2013; 45:136-144. Spier I, et al. Int J Cancer 2015;137:320-331. Bellido F, et al. Genet Med 2016;18:325-332.**
POLE	**Not well-established**	**Uncertain – presumed high risk from limited case reports**	**Polymerase proofreading-associated polyposis**	**Bellido F, et al. Genet Med 2016;18:325-332.**
PMS2	**Well-established**	**High**	**LS**	**See Lynch Syndrome Guidelines (LS-1)**
PTEN	**Well-established**	**Moderate-High**	**Cowden syndrome/PTEN hamartoma syndrome**	**See NCCN Guidelines for Genetic/Familial High-Risk Assessment: Breast and Ovarian**
SMAD4	**Well-established**	**High**	**JPS**	**See Juvenile Polyposis Syndrome Guidelines (JPS-1)**
STK11	**Well-established**	**High**	**PJS**	**See Peutz-Jeghers Syndrome Guidelines (PJS-1)**
TP53	**Well-established**	**High**	**Li-Fraumeni syndrome**	**See NCCN Guidelines for Genetic/Familial High-Risk Assessment: Breast and Ovarian**

*Risk level is based on panel consensus.

[g]*RPS20* is an emerging gene that is potentially linked to CRC, and there are not enough data at present to include *RPS20* on this list.

Continued

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Table 5: Recommended Management for Patients with Pathogenic Variants in Genes That May Confer a Risk for Colorectal Cancer

GENE	RECOMMENDATION
APC	See NCCN Guidelines for Familial Adenomatous Polyposis (FAP-1)
BMPR1A	See NCCN Guidelines for Juvenile Polyposis Syndrome (JPS-1)
LS genes *(MLH1, MSH2, MSH6, PMS2, EPCAM)*	See NCCN Guidelines for Lynch Syndrome (LS-2)
MUTYH biallelic pathogenic variants	See NCCN Guidelines for *MUTYH*-Associated Polyposis (MAP-1)
PTEN	See NCCN Guidelines for Genetic/Familial High-Risk Assessment: Breast and Ovarian
STK11	See NCCN Guidelines for Peutz-Jeghers Syndrome (PJS-1)
SMAD4	See NCCN Guidelines for Juvenile Polyposis Syndrome (JPS-1)
TP53	See NCCN Guidelines for Genetic/Familial High-Risk Assessment: Breast and Ovarian
GREM1[h]	• Begin colonoscopy at age 25–30 y and every 2–3 y if negative. If polyps are found, colonoscopy every 1–2 y with consideration of surgery if the polyp burden becomes unmanageable by colonoscopy. • Surgical evaluation if appropriate.
POLD1[h]	
POLE[h]	
AXIN2[h]	
NTHL1 biallelic pathogenic variants[h]	
MSH3 biallelic pathogenic variants[h]	
APC I1307K pathogenic variant[h] *CHEK2*[h,i]	• For probands with CRC and one of these pathogenic variants: ▸ See surveillance recommendations for post-CRC resection: ◊ NCCN Guidelines for Colon Cancer ◊ NCCN Guidelines for Rectal Cancer • For probands unaffected by CRC with a first-degree relative with CRC: ▸ Colonoscopy screening every 5 y, beginning at age 40 or 10 y prior to age of first-degree relative's age at CRC diagnosis. • For probands unaffected by CRC and no first-degree relative with CRC: ▸ Colonoscopy screening every 5 y, beginning at age 40.
MUTYH heterozygotes[h]	• For probands unaffected by CRC with a first-degree relative with CRC: ▸ Colonoscopy screening every 5 y, beginning at age 40 y or 10 y prior to age of first-degree relative's age at CRC diagnosis. See screening recommendations in NCCN Guidelines for Colorectal Cancer Screening. • There are no specific data available to determine screening recommendations for a patient with an *MUTYH* heterozygous pathogenic variant and a second-degree relative affected with CRC. See NCCN Guidelines for Colorectal Cancer Screening. • For probands unaffected by CRC with NO family history of CRC: ▸ Data are unclear as to whether specialized screening is warranted for *MUTYH* monoallelic carriers unaffected by CRC with no family history of CRC.[j]

See footnotes on GENE-8.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

[h]The panel recognizes that data to support the surveillance recommendations for these particular genes are evolving at this time. Caution should be used when implementing final colonoscopy surveillance regimens in context of patient preferences and new knowledge that may emerge.
[i]Heterogeneity in CRC risk may exist based on type of pathogenic *CHEK2* variant (Han F, Guo C, Liu L. The effect of CHEK2 variant I157T on cancer susceptibility: evidence from a meta-analysis. DNA Cell Biol 2013;32:329-335; Liu C, Wang Q, Wang Y. The CHEK2 I157T variant and colorectal cancer susceptibility: a systematic review and meta-analysis. Asian Pac J Cancer Prev 2012;13:2051-2055; Xiang H, Geng X, Ge W, Li H. Meta-analysis of CHEK2 1100delC variant and colorectal cancer susceptibility. Eur J Cancer 2011;47:2546-2551.); some patients may elect for less aggressive screening based on shared decision-making. One model has suggested that earlier screening than the average risk initiation may be justified for *CHEK2* 1100delC and I157T carriers based on reaching same risk for CRC at an earlier age than observed among average-risk persons initiating screening at age 50 (Katona B, Yurgelun M, Garber J, et al. A counseling framework for moderate-penetrance colorectal cancer susceptibility genes. Genet Med 2018;20:1324-1327.).
[j]Katona B, Yurgelun M, Garber J, et al. A counseling framework for moderate-penetrance colorectal cancer susceptibility genes. Genet Med 2018;20:1324-1327.

Note: All recommendations are category 2A unless otherwise indicated.
Clinical Trials: NCCN believes that the best management of any patient with cancer is in a clinical trial. Participation in clinical trials is especially encouraged.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Discussion

NCCN Categories of Evidence and Consensus

Category 1: Based upon high-level evidence, there is uniform NCCN consensus that the intervention is appropriate.

Category 2A: Based upon lower-level evidence, there is uniform NCCN consensus that the intervention is appropriate.

Category 2B: Based upon lower-level evidence, there is NCCN consensus that the intervention is appropriate.

Category 3: Based upon any level of evidence, there is major NCCN disagreement that the intervention is appropriate.

All recommendations are category 2A unless otherwise indicated.

Table of Contents

Overview MS-2

Literature Search Criteria and Guidelines Update Methodology MS-2

Assessment for Hereditary CRC Syndrome (HRS-1) MS-3

- Criteria for the Evaluation of Lynch Syndrome (HRS-3) MS-3
- Management After Diagnosis with a Genetic Syndrome MS-4

Lynch Syndrome (LS-1) MS-4

- Strategies for Evaluating for Lynch Syndrome in Individuals Meeting Criteria for the Evaluation of Lynch Syndrome (LS-1) MS-5
- Lynch Syndrome Management (LS-2) MS-9
- Lynch Syndrome Colonoscopy Surveillance Findings and Follow-up (LS-5) MS-12
- Adenomatous Polyposis Testing Criteria (POLYP-1) MS-13

Familial Adenomatous Polyposis (FAP/AFAP-1) MS-14

- Diagnosis: Classical vs. Attenuated FAP MS-15
- Management of FAP and AFAP MS-15
- Surveillance Following Surgery for FAP (FAP-1) MS-19
- Surveillance After Surgery for AFAP (AFAP-1) MS-21

MUTYH-Associated Polyposis (MAP-1) MS-22

- Preoperative and Surgical Management of MAP (MAP-2/-3) MS-24
- Postoperative Surveillance in MAP (MAP-2) MS-24

Peutz-Jeghers Syndrome (PJS-1) MS-24

- Management of Peutz-Jeghers Syndrome (PJS-2) MS-25

Juvenile Polyposis Syndrome (JPS-1) MS-25

- Management of Juvenile Polyposis Syndrome MS-26

Serrated Polyposis Syndrome (SPS-1) MS-26

- Management of Serrated Polyposis (SPS-1) MS-27

Colonic Adenomatous Polyposis of Unknown Etiology (CPUE-1) MS-27

Multi-Gene Testing (GENE-1) MS-28

References MS-33

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Overview

Colorectal cancer (CRC) is the fourth most frequently diagnosed cancer and the second leading cause of cancer death in the United States. In 2019, an estimated 101,420 new cases of colon cancer and 44,180 new cases of rectal cancer will occur in the United States. During the same year, it is estimated that 51,020 people will die from colon and rectal cancer.[1] Importantly, the incidence of CRC per 100,000 decreased from 60.5 in 1976 to 46.4 in 2005.[2] The incidence rate for CRC reported by the CDC for 2011 is 40.0 per 100,000 persons.[3] In addition, mortality from CRC decreased by almost 35% from 1990 to 2007,[4] and by 53% from 1970 to 2016.[1] These improvements in incidence of and mortality from CRC are thought in part to be a result of cancer prevention and earlier diagnosis through screening and better treatment modalities.

Despite the observed improvements in the overall CRC incidence rate, a retrospective cohort study of the SEER colorectal cancer registry found that the incidence of CRC in patients younger than 50 years has been increasing.[5] The authors estimate that the incidence rates for colon and rectal cancers will increase by 90.0% and 124.2%, respectively, for patients 20 to 34 years of age by 2030. The cause of this trend is currently unknown.

CRC often occurs sporadically, but familial cancer syndromes are also common in this disease. Genetic susceptibility to CRC includes well-defined inherited syndromes such as Lynch syndrome (also known as hereditary nonpolyposis colorectal cancer, or HNPCC), familial adenomatous polyposis (FAP), and MutY human homolog (*MUTYH*)-associated polyposis (MAP). Other entities include Muir-Torre, Turcot, Gardner, Cowden, Bannayan-Riley-Ruvalcaba, Peutz-Jeghers, juvenile polyposis, and serrated polyposis syndromes.[6-8]

These NCCN Guidelines for Genetic/Familial High-Risk Assessment: Colorectal provide recommendations for the management of patients with high-risk syndromes, including Lynch syndrome, FAP, MAP, Peutz-Jeghers syndrome (PJS), juvenile polyposis syndrome (JPS), serrated polyposis syndrome (SPS), and other high-risk syndromes associated with CRC risk (Li-Fraumeni syndrome [LFS] and Cowden syndrome/PTEN hamartoma tumor syndrome [PHTS]).

Literature Search Criteria and Guidelines Update Methodology

Prior to the update of this version of the NCCN Guidelines® for Genetic/Familial High-Risk Assessment: Colorectal, an electronic search of the PubMed database was performed to obtain key literature in the field of high-risk CRC published since the previous Guidelines update, using the following search terms: (lynch syndrome) or (hereditary nonpolyposis colorectal cancer) or (familial adenomatous polyposis) or (MUTYH polyposis) or (Peutz-Jeghers syndrome) or (polyposis syndrome) or (familial colon cancer) or (familial rectal cancer) or (familial colorectal cancer) or (hereditary colon cancer) or (hereditary rectal cancer) or (hereditary colorectal cancer). The PubMed database was chosen because it remains the most widely used resource for medical literature and indexes only peer-reviewed biomedical literature.[9]

The search results were narrowed by selecting studies in humans published in English. Results were confined to the following article types: Clinical Trial, Phase II; Clinical Trial, Phase III; Clinical Trial, Phase IV; Guideline; Practice Guidelines; Randomized Controlled Trials; Meta-Analysis; Systematic Reviews; and Validation Studies.

The data from key PubMed articles and articles from additional sources deemed as relevant to these Guidelines and discussed by the panel have been included in this version of the Discussion section (eg, e-publications ahead of print, meeting abstracts). Recommendations for which high-level

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

evidence is lacking are based on the panel's review of lower-level evidence and expert opinion.

The complete details of the Development and Update of the NCCN Guidelines are available on the NCCN website (available at www.NCCN.org).

Assessment for Hereditary CRC Syndrome (HRS-1)

Genetic susceptibility to CRC includes well-defined inherited syndromes such as Lynch syndrome, FAP, MAP, and other less common syndromes. Many approaches have been proposed for identifying individuals with hereditary CRC syndromes. NCCN recommends a stepwise approach. First, if an individual has a personal or family history of a known pathogenic variant in a colorectal polyposis or cancer gene, further evaluation and management appropriate for established hereditary CRC syndromes is warranted. A pathogenic variant in this case includes likely pathogenic variants.[10] Second, if there is no known personal or family history of a known pathogenic variant in a colorectal polyposis or cancer gene, the patient's personal history of any of the following should be determined:

- >10 adenomatous polyps, or
- ≥2 hamartomatous polyps, or
- ≥5 serrated polyps proximal to the sigmoid colon

NCCN recommends that individuals meeting any of the above criteria have detailed risk assessment and potential genetic evaluation to rule out polyposis syndromes (HRS-2). The presence of >10 adenomas may be linked to FAP, attenuated FAP (AFAP), MAP, or rare genetic causes of multiple adenomatous polyps including genetic mutations in *AXIN2*, *GREM1*, *NTHL1*, *POLE*, *POLD1*, or *MSH3*; ≥2 hamartomatous polyps may be associated with PJS, JPS, or Cowden syndrome/PHTS (see the NCCN Guidelines for Genetic/Familial High-Risk Assessment: Breast and Ovarian); and ≥5 serrated polyps proximal to the sigmoid colon may be associated with SPS.

Third, if the patient's personal history is not suspicious for a polyposis syndrome, personal and family history of Lynch syndrome-associated cancers should be elicited. Lynch syndrome-associated cancers include: colorectal, endometrial, gastric, ovarian, pancreatic, ureter and renal pelvis, brain (usually glioblastoma), biliary tract, and small intestinal cancers, as well as sebaceous adenomas, sebaceous carcinomas, and keratoacanthomas as seen in Muir-Torre syndrome. Those with a personal or family history of Lynch syndrome-related cancers should undergo further evaluation (See *Criteria for the Evaluation of Lynch Syndrome*).

Individuals not meeting any of the above criteria may be considered average risk for CRC, and follow the NCCN Guidelines for Colorectal Cancer Screening, unless other significant personal or family history indicate increased risk for a hereditary cancer syndrome. Increased risk warranting genetic evaluation may be indicated by, but not restricted to personal or family history of congenital hypertrophy of the retinal pigment epithelium, osteomas, supernumerary teeth, desmoid tumor, cribriform variant of papillary thyroid cancer, brain cancer (typically medulloblastoma), and hepatoblastoma.

Criteria for the Evaluation of Lynch Syndrome (HRS-3)

If an individual has a personal or family history of a Lynch syndrome-related cancer, the panel has summarized criteria under three domains that can be used to select patients for the evaluation of Lynch syndrome:

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Personal history of colorectal, endometrial, or other Lynch syndrome-associated cancer:

- An individual with colorectal or endometrial cancer at any age with tumor showing evidence of mismatch repair (MMR) deficiency, either by microsatellite instability (MSI) or loss of MMR protein expression
- Known Lynch syndrome pathogenic variant in the family
- An individual with CRC or endometrial cancer and any of the following:
  - Diagnosed <50 years
  - Another synchronous or metachronous Lynch syndrome-related cancer
  - ≥1 first-degree or second-degree relative with Lynch syndrome-related cancer diagnosed <50 years
  - ≥2 first-degree or second-degree relatives with Lynch syndrome-related cancers regardless of age
- An individual with colorectal tumor with MSI high (MSI-H) histology (ie, presence of tumor-infiltrating lymphocytes, Crohn's-like lymphocytic reaction, mucinous/signet ring differentiation, medullary growth pattern)

Family history of any of the following:

- ≥1 first-degree relative with CRC or endometrial cancer diagnosed <50 years
- ≥1 first-degree relative with CRC or endometrial cancer and another synchronous or metachronous Lynch syndrome-related cancer
- ≥2 first-degree or second-degree relatives with Lynch syndrome-related cancer; including ≥1 diagnosed <50 years
- ≥3 first-degree or second-degree relatives with Lynch syndrome-related cancers, regardless of age

Increased model-predicted risk for Lynch syndrome:

- An individual with a ≥5% risk of having an MMR gene pathogenic variant based on predictive models (PREMM5,[11] MMRpro, MMRpredict)

The panel recommends tumor screening for MMR deficiency for all CRC and endometrial cancers regardless of age at diagnosis; however, germline genetic testing is generally reserved for patients diagnosed at an early age, with positive family history, or abnormal tumor testing results including MSI or loss of MMR protein expression. A study evaluating the performance of the PREMM5 prediction model relative to a previous model, PREMM$_{1,2,6}$, found that PREMM5 quantified the risk of an individual with MMR gene mutations at a threshold of ≥2.5%, suggesting that testing can be reasonably performed based on this score.[11] It is worth noting that at a threshold of ≥2.5%, there is an increase in sensitivity, but a decrease in specificity, and it is unknown how this applies to the general population of unaffected individuals. For these reasons, the panel did not issue an unqualified recommendation to utilize a threshold of ≥2.5% to trigger testing. The threshold will be revisited, including if data on specificity of this approach when applied to the general population of unaffected individuals become available.

Management After Diagnosis with a Genetic Syndrome

Following evaluation, those with Lynch syndrome, FAP, MAP, and other syndromes are managed as described in the following sections.

Lynch Syndrome (LS-1)

Lynch syndrome is the most common form of genetically determined colon cancer predisposition, accounting for 2% to 4% of all CRC cases,[12-15] and a consensus is emerging across medical specialty societies and expert groups regarding the best strategies for identifying patients with this

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

condition. Lynch syndrome results from a germline mutation in 1 of 4 DNA MMR genes (*MLH1*, *MSH2*, *MSH6*, or *PMS2*).[16] Additionally, deletions in the *EPCAM* gene, which lead to hypermethylation of the *MSH2* promoter and subsequent *MSH2* silencing, cause Lynch syndrome.[17,18] Identification of Lynch syndrome is important both for individuals with cancer, because of high personal risk for metachronous Lynch syndrome cancers (ie, endometrial cancer after CRC or vice versa; second CRC), and for their families because of autosomal dominant inheritance and potentially high penetrance. After identification of Lynch syndrome, surveillance (particularly for first or metachronous CRC) offers an opportunity for early detection and perhaps even prevention of cancer among mutation carriers. Further, cancer site-specific evaluation and heightened attention to symptoms is also advised for other cancers that occur with increased frequency in affected persons, including colorectal, endometrial, gastric, ovarian, pancreatic, ureter and renal pelvis, biliary tract, brain (glioblastoma), and small intestinal cancers, as well as sebaceous gland adenomatous polyps and keratoacanthomas.

Strategies for Evaluating for Lynch Syndrome in Individuals Meeting Criteria for the Evaluation of Lynch Syndrome (LS-1)

Deleterious Lynch syndrome pathogenic variant in family is known: When a known Lynch syndrome pathogenic variant exists in the family, the individual should be tested for the familial pathogenic variant. If the test is positive or if testing is not performed for any reason, the individual should follow surveillance or prevention strategies for Lynch syndrome outlined below (See *Lynch Syndrome Management*). In addition, genetic testing should be offered to at-risk family members. However, the recommendation to manage patients in whom genetic testing was not done is category 2B. Individuals who test negative for the familial Lynch syndrome pathogenic variant, or who do not have a family history of a Lynch syndrome-related cancer are considered to be at average risk for CRC and should follow the NCCN Guidelines for Colorectal Cancer Screening. Additional testing may be indicated based on personal family and medical history.

No known Lynch syndrome pathogenic variant in proband or family: The traditional approach to identifying individuals at risk for Lynch syndrome has generally employed a 2-step screening process. First, patients meeting clinical criteria based on family history, personal history of cancer, and/or pathologic characteristics are identified, followed by additional application of screening with a molecular test.

The Amsterdam II criteria outline increased risk for Lynch syndrome in a family with a proband affected by CRC or any other Lynch syndrome-associated cancer (ie, endometrial, small bowel, ureter, or renal-pelvic cancers), and three relatives with a Lynch syndrome-associated cancer provided the following family criteria are met:

- One relative should be a first-degree relative of the other two
- At least two successive generations should be affected
- At least one Lynch syndrome-associated cancer should have been diagnosed before age 50 years

Additionally, the Amsterdam II criteria stipulate that FAP should be excluded, and tumors should be verified through pathologic examination.[19] Approximately 50% of families meeting the Amsterdam II criteria have a mutation in an MMR gene.[20] These criteria are very stringent, however, and miss as many as 68% of patients with Lynch syndrome.[21]

The Bethesda Guidelines were later developed and updated to provide broader clinical criteria for Lynch syndrome screening.[22] Updated Bethesda criteria are as follows:[23]

- CRC diagnosed in a patient younger than age 50 years

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

- Synchronous, metachronous, colorectal, or other tumor associated with Lynch syndrome
- CRC with MSI-H histology (ie, presence of tumor-infiltrating lymphocytes, Crohn's-like lymphocytic reaction, mucinous/signet-ring differentiation, medullary growth pattern) in a patient younger than 60 years
- CRC in a patient with a family history of cancer diagnosed earlier than age 50 years and associated with Lynch syndrome. If more than one relative was diagnosed with a Lynch syndrome-associated cancer, then the age criterion is not needed.

One study reported that *MLH1* and *MSH2* mutations were detected in 65% of patients with MSI of colon cancer tissue who met the Bethesda criteria.[24] Another study reported on the accuracy of the revised Bethesda criteria, concluding that the guidelines were useful for identifying patients who should undergo further testing.[25] Patients fulfilling the revised Bethesda criteria had an odds ratio (OR) for carrying a germline mutation in *MLH1* or *MSH2* of 33.3 (95% confidence interval [CI], 4.3–250; *P* = .001). Still, a considerable number of patients with Lynch syndrome fail to meet even the revised Bethesda Guidelines.[14]

Statistical models that predict risk for carrying a mutation in a DNA MMR gene are an additional commonly applied clinical approach to identifying individuals at risk for Lynch syndrome.[21,26-28] These models give probabilities of mutations and/or of the development of future cancers based on family and personal history. The PREMM5 model can be used online at http://premm.dfci.harvard.edu/ and the MMRpredict model is available for online use at http://hnpccpredict.hgu.mrc.ac.uk/. MMRpro is available for free download at http://www4.utsouthwestern.edu/breasthealth/cagene/. Using a cut-off of 5%, one study suggests that both PREMM5 and MMRpredict are effective at predicting an individual's risk of carrying MMR mutations, but they may be less effective at identifying individuals with *PMS2* mutations.[29]

Overall, for individuals without a previously known Lynch syndrome-associated pathogenic variant, the panel recommends additional evaluation for Lynch syndrome based on clinical criteria (see *Criteria for the Evaluation of Lynch Syndrome*), including for individuals with no known Lynch syndrome pathogenic variant who meet the Amsterdam II criteria or Bethesda Guidelines, have a cancer diagnosis prior to age 50 years, or have a predicted risk for Lynch syndrome >5% on one of the following prediction models: MMRpro, PREMM5,[11] or MMRpredict.

A problem with nearly all clinically based criteria for identifying individuals with Lynch syndrome is suboptimal sensitivity. This has led several groups to study an alternative strategy, referred to as "universal screening," in which all individuals newly diagnosed with CRC have either MSI or immunohistochemistry (IHC) testing for absence of 1 of the 4 DNA MMR proteins. This approach provides a sensitivity of 100% (95% CI, 99.3%–100%) and a specificity of 93.0% (95% CI, 92.0%–93.7%) for identifying individuals with Lynch syndrome.[30] An alternative approach is to test all patients with CRC diagnosed prior to age 70 years plus patients diagnosed at older ages who meet the Bethesda Guidelines.[30] This approach gave a sensitivity of 95.1% (95% CI, 89.8%–99.0%) and a specificity of 95.5% (95% CI, 94.7%–96.1%). This alternative approach had improved sensitivity compared to the revised Bethesda criteria, and improved specificity compared to universal screening regardless of age.

Cost-effectiveness of universal screening has been established and has been endorsed by the Evaluation of Genomic Applications in Practice and Prevention (EGAPP) working group at the Centers for Disease Control and Prevention (CDC), the U.S. Multi-Society Task Force on Colorectal Cancer, and the European Society for Medical Oncology (ESMO).[31-35]

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

The panel recommends universal screening of all CRCs, in order to maximize sensitivity for Lynch syndrome detection and simplify care processes.[30,36,37] The panel emphasizes that great care must be taken in implementing system-level universal testing to avoid loss of follow-up for patients with abnormal tests and to avoid misinterpretation of the molecular screening tests, and accordingly recommends that an infrastructure needs to be in place to handle the screening results.[38] The panel also suggests that counseling by an individual with expertise in genetics is not required prior to routine tumor testing, but strongly recommends follow-up with a provider with expertise in genetics following a positive screen (see below).

Initial Tumor Testing Methodologies

Screening for Lynch syndrome currently requires performance of 1 of 2 molecular tests (see *Principles of IHC and MSI Testing for Lynch Syndrome* in algorithm), either after the aforementioned clinical criteria are met, or as part of a universal screening strategy with: 1) IHC for abnormal absence of MMR protein expression; or 2) MSI analysis to evaluate for MSI-H on a tumor specimen.[39] Greater than 90% of Lynch syndrome tumors are MSI-H and/or lack expression of at least one of the MMR proteins by IHC.

IHC analysis has the advantage of predicting which gene is most likely to be mutated (the gene for the affected protein or its corresponding dimer partner) and thus the first candidate(s) for germline sequencing.[39] Interpretation of IHC test reports can sometimes be confusing; when “positive” IHC is reported, care should be taken to ensure that “positive” means abnormal absence of MMR protein expression, as opposed to normal presence of expression.

MSI testing panels may consist of mononucleotide and dinucleotide markers.[40] In a study including 1058 patients with CRC, detection of MMR deficiency by a panel including both mononucleotide and dinucleotide markers (BAT26, BAT25, D5S346, D2S123, and D17S250) was compared to that of a panel including only mononucleotide markers (BAT26, BAT25, NR21, NR22, and NR24).[41] Sensitivity and positive predictive value of the panel including only mononucleotide markers (95.8% and 88.5%, respectively) were better, compared to the panel including both mononucleotide and dinucleotide markers (76.5% and 65.0%, respectively).

Some studies have shown that both IHC and MSI are cost-effective and useful for selecting high-risk patients who may have *MLH1*, *MSH2*, and *MSH6* germline mutations.[33,42,43] However, conclusive data are not yet available that establish which strategy is optimal.[16,25,44-47] A review showed that the sensitivities of MSI and IHC testing are 77% to 89% and 83%, respectively; specificities are 90% and 89%, respectively[33]; as mentioned previously, even higher sensitivity has been reported for screening with MSI and IHC in context of “universal screening”. An analysis of 5591 unrelated CRC probands undergoing both MSI and IHC testing showed a concordance rate of 97.5%.[30] Some experts advocate for using both methods when possible.[48] However, the panel recommends using only one test initially. If normal results are found and Lynch syndrome is strongly suspected, then the other test may be carried out. Alternatively, emerging studies suggest a role for next-generation sequencing (NGS) panels in Lynch syndrome tumor testing.[49-51]

Where genetic testing is recommended, the panel recommends consultation with an individual with expertise in genetics, and germline testing to exclude presence of Lynch-associated mutations. The approach to mutation testing is evolving. Previously, a sequential approach in which 1 or 2 genes were sequenced guided by either disease prevalence or IHC results, followed by additional testing of other genes was followed. Recognition of scenarios in which IHC results were not available also allowed for syndrome-specific testing of the panel of genes that cause

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Lynch syndrome (*MLH1*, *MSH2*, *MSH6*, *PMS2*, and *EPCAM*) simultaneously. Reductions in cost of sequencing, and recognition that some patients meeting Lynch syndrome testing criteria may have germline mutations not associated with Lynch syndrome have led to growing use of so called “multi-gene” panels in clinical practice. These panels test not only for Lynch syndrome-associated genes, but also for additional mutations. The panel recommends that for patients or families where colorectal or endometrial tumor is available, one of three options should be considered for workup: 1) tumor testing with IHC or MSI; 2) comprehensive tumor NGS panel (that includes, at minimum, the 4 MMR genes and *EPCAM*, *BRAF*, MSI, and other known familial cancer genes); or 3) germline multi-gene testing that includes the four MMR genes and *EPCAM*. The panel recommends tumor testing with IHC and/or MSI be used as the primary approach for pathology-lab–based universal screening. If no tumor is available, tumor material is insufficient, or the affected relative is unavailable, germline multi-gene testing may be considered that includes the four MMR genes and *EPCAM*. Multi-gene testing may be preferred, particularly for patients with a strong family history or if the age of diagnosis is less than 50 years.[52,53]

Follow-up Testing of Individuals with Increased Risk Based on Screening
If abnormal MSI or IHC for one of the DNA MMR proteins is identified within a colorectal or endometrial cancer, then a differential diagnosis must be considered. For example, 10% to 15% of CRCs have MSI or abnormal IHC (particularly in the case of absent *MLH1* expression) due to sporadic development of cancer, rather than an underlying inherited (germline) genetic mutation. *Tumor Testing Results and Additional Testing Strategies* in the algorithm identifies a range of test result scenarios, the differential diagnosis, and recommended follow-up. In some scenarios, such as with absent *MSH2* expression by IHC, follow-up germline testing for indicated genes is directly recommended. In other scenarios, additional testing of tumor tissue is recommended. For example, for the common scenario of absent *MLH1* expression by IHC, the panel recommends additional tumor testing for presence of *MLH1* hypermethylation and/or *BRAF* V600E mutation, either of which would be consistent with sporadic, rather than Lynch syndrome-associated, cancer.[35,39,54,55]

Follow-up of Genetic Test Results
If a pathogenic variant for familial Lynch syndrome is found, the panel recommends that Lynch syndrome management guidelines be followed (See *Lynch Syndrome Management*).

If no pathogenic variant for familial Lynch syndrome is found, clinicians are advised to confirm that testing for large rearrangements and deletions of MMR genes were performed by the lab test provider. If still no pathogenic variant or a variant of uncertain significance (VUS) is identified, the panel recommends tailored surveillance based on individual and family risk assessment. Notably, some individuals with abnormal MSI and/or IHC tumor results and no germline mutation detected in the corresponding gene(s) may still have undetected Lynch syndrome. At this time, no consensus has been reached as to whether these patients (sometimes referred to as having “Lynch-like syndrome”) should be managed as having Lynch syndrome or managed based on personal/family history. Growing evidence suggests a subset of these individuals may have double somatic mutations/changes in the MMR genes.[56] Although the efficacy of the approach has not yet been proven, genetic testing of the corresponding gene(s) could be performed on tumor DNA to assess for somatic mutations. Individuals found to have double somatic mutations/changes in the MMR genes are unlikely to have Lynch syndrome, though double somatic mutations might also be due to non-Lynch germline mutations. Thus, management should be based on personal/family history until further research on Lynch-like syndrome emerges. Additionally, germline testing may be normal despite a strong family history (ie, Amsterdam criteria) or additional features of hereditary

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

cancer syndromes (multiple colon polyps) being present. In these cases, additional testing may be warranted in the proband (such as expanded multi-gene testing), or tumor testing in an affected family member could be considered due to the possibility of a phenocopy.

Newly Identified Lynch Syndrome

When a Lynch syndrome pathogenic variant is found in the family, it offers an opportunity to provide predictive testing for at-risk family members. If a first-degree relative is unavailable or unwilling to be tested, more distant relatives should be offered testing for the known family mutation.

There are many other issues involved in the genetic counseling process of individuals for presymptomatic testing for cancer susceptibility. Some individuals elect not to undergo testing, and it is important to counsel these individuals so they continue with increased surveillance.

Lynch Syndrome Management (LS-2)

The NCCN Panel carefully considered surveillance schemes for individuals with Lynch syndrome. Compared to the general population, these patients are at increased lifetime risk for CRC (46%–49% vs. 4.5%), endometrial cancer (43%–57% vs. 2.7%), and other cancers including of the stomach and ovary.[57-62] Within Lynch syndrome carriers, risk may vary by specific type of DNA MMR gene mutation. For example, individuals with *PMS2* mutations have a 12% to 20% risk for colon cancer, while those with *MLH1* mutations have a 46% to 49% risk. The panel recognizes that there continues to be controversy regarding whether mutation-specific risks should guide differential management.[63] The panel's current approach is to offer uniform recommendations for cancer surveillance and prevention, recognizing that, in some clinical scenarios, delaying initiation of surveillance (eg, later starting age for colonoscopy surveillance among *PMS2* carriers) may be appropriate, pending availability of large cohort studies of risk among specific mutation carriers.

Existing data on screening refer primarily to colon and endometrial cancers. More data are needed to evaluate the risks and benefits of extracolonic and extra-endometrial cancer screening, and recommendations are based mainly on expert opinion. The panel recognizes that there are limited population-based studies on lifetime risk for most of the cancers related to each of these genes. Although there are some mutation-specific data available, a generalized screening approach is suggested for many surveillance strategies. Screening and the option of risk-reducing surgeries should be individualized after risk assessment and counseling.

Colon Cancer Surveillance

If Lynch syndrome is confirmed, colonoscopy is advised to start between the ages of 20 to 25 or 2 to 5 years younger than the youngest diagnosis age in the family, whichever comes first, and should be repeated every 1 to 2 years.[34,35,54,55,64,65] Some patients may benefit from a shorter 1-year versus a longer 2-year screening interval.[66] Factors that may favor a 1-year interval may include: being male, age >40 years, having *MLH1*/*MSH2* pathogenic variants, or having a history of CRC or adenomas.[66,67] For *MSH6* mutation carriers, consider a later age of onset for colonoscopy initiation, such as at age 30 years or 10 years younger than age of any relative with CRC.[60,68] Due to limited data for the *PMS2* gene, the panel is not able to make a specific recommendation regarding later age of onset for colonoscopy.[64] There is some uncertainty regarding best age to initiate colonoscopic surveillance. For example, the results of a meta-analysis in which CRC risk in 1114 Lynch syndrome families (*MLH1* and *MSH2* mutation carriers) was examined showed that 5-year CRC risk for those aged 20 to 29 years is about 1%, with the risk for those aged 30 to 39 years being 3% to 5%, with greater risk in men.[69] The investigators argued that annual colonoscopy in patients aged 25 to 29 years may be an overly aggressive recommendation that is not cost-effective (ie, 155 men and 217 women in this age group would need to be screened to prevent one

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

CRC death). However, the panel concluded that more evidence was needed in order to understand best age of initiation of screening.

Chromoendoscopy may also be used during colonoscopy in which dye spray is used to enhance visualization. A systematic review of four studies indicated that chromoendoscopy is a promising technique for improving detection of lesions and flat adenomas in patients with Lynch syndrome.[70] Only one of these studies was a prospective randomized trial, however, and this trial was limited by a small sample of patients who had already undergone colonoscopy and inadequate statistical power to detect clinically meaningful effects.[71] Chromoendoscopy may be considered in patients with Lynch syndrome, but larger prospective randomized trials are needed to better understand its role in Lynch syndrome.

Endometrial Cancer Surveillance (LS-3)

Women with Lynch syndrome are at heightened risk for endometrial cancer.[58,64,72,73] With a lifetime risk of up to 60%, endometrial cancer is the second most common cancer in women with Lynch syndrome.[72] Education that enhances recognition and prompt reporting of relevant symptoms (ie, dysfunctional uterine bleeding or postmenopausal bleeding) is advised in order to promote early endometrial cancer detection. The evaluation of these symptoms should include an endometrial biopsy. Endometrial cancer screening does not have proven benefit in women with Lynch syndrome. However, endometrial biopsy is highly sensitive and specific as a diagnostic procedure. Screening through endometrial biopsy every 1 to 2 years may be considered.[74-79] Routine transvaginal ultrasound to screen for endometrial cancer in postmenopausal women has not been shown to be sufficiently sensitive or specific to warrant a positive recommendation,[75-80] but may be considered at the clinician's discretion. However, transvaginal ultrasound is not recommended as a screening tool in premenopausal women due to the wide range of endometrial strip thickness throughout the normal menstrual cycle. Total abdominal hysterectomy has not been shown to reduce endometrial cancer mortality, but is an option that may be considered for risk reduction in women who have completed childbearing and carry a *MLH1*, *MSH2*, *EPCAM*, *PMS2*, or *MSH6* mutation.[54,65,74,76,81,82] The timing of a hysterectomy can also be individualized based on comorbidities, family history, and Lynch syndrome gene mutation, as risks for endometrial cancer vary by mutated gene. An observational study showed that hormonal contraceptive use is associated with lower risk for endometrial cancer in carriers of MMR mutations (HR, 0.39; 95% CI, 0.23–0.64, $P < .001$).[83] However, prospective data are needed before hormonal contraceptives are recommended for prevention of gynecologic cancers in patients with Lynch syndrome. In general, risk reduction agents should be considered, with detailed discussion between the physician and patient outlining the associated risks and benefits.

Ovarian Cancer Surveillance (LS-3)

Women with Lynch syndrome are also at a heightened risk for ovarian cancer, which varies based on affected MMR gene and age.[58,61,64,67,72,73] There are circumstances where clinicians may find screening helpful; however, the data do not support routine ovarian cancer screening for Lynch syndrome. Transvaginal ultrasound and serum CA-125 testing to screen for ovarian cancer in postmenopausal women has not been shown to be sufficiently sensitive or specific to warrant a routine recommendation,[75-80] but may be considered at the clinician's discretion. Since there is no effective screening for ovarian cancer, women should be educated on the symptoms that may be associated with the development of ovarian cancer, such as pelvic or abdominal pain, bloating, increased abdominal girth, difficulty eating, early satiety, or increased urinary frequency or urgency. Symptoms that persist for several weeks and are a change from a woman's baseline should prompt evaluation by her physician. Bilateral salpingo-oophorectomy (BSO) may reduce the incidence of ovarian cancer.[54,65,74,76,81,82] The decision and timing of BSO as an option may be considered and individualized based on whether

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

childbearing is complete, menopausal status, comorbidities, family history, and Lynch syndrome gene, as risks for ovarian cancer vary by mutated gene. There is insufficient evidence to recommend risk-reducing salpingo-oophorectomy (RRSO) in *MSH6* and *PMS2* mutation carriers. Similar to endometrial cancer management, risk reduction agents should be considered, with detailed discussion between the physician and patient outlining the associated risks and benefits.

Surveillance for Other Cancers (LS-2)

The lifetime risk for gastric cancer varies widely between individuals with Lynch syndrome in different populations, from 2% to 4% in the Netherlands to 30% in Korea.[64,84] Most cases occur after age 40 years, and males have a stronger predisposition. Lynch syndrome is also associated with a 3% to 6% risk for small bowel cancer.[58,61,85-88] There is no clear evidence to support screening for gastric, duodenal, and more distal small bowel cancer in patients with Lynch syndrome.[89] Select individuals with a family history of gastric, duodenal, or more distal small bowel cancer may have an increased risk[90] and benefit from surveillance. Individuals of Asian descent or other countries with high background incidence of gastric cancer may have an increased risk for stomach cancer and may benefit from surveillance. For individuals with *MLH1*, *MSH2*, or *EPCAM* mutations who have an increased risk, physicians may consider upper esophagogastroduodenoscopy (EGD) extended to the distal duodenum or into the jejunum every 3 to 5 years starting at age 40 years.[59,91] Infection with Helicobacter pylori (H. pylori) is a cause of gastric cancer.[92,93] Given the increased risk for gastric cancer in patients with Lynch syndrome, testing and treating for H. pylori, if detected, should be considered. This is consistent with recommendations by ASCO and ESMO.[34,54]

Risk for urinary tract cancer in patients with Lynch syndrome varies and ranges from less than 1% to 18%, with greater risk among carriers of *MSH2* mutations (ranging from 2%–18%), relative to *MLH1* (ranging from 0.2%–5%) and *MSH6* (ranging from 0.7%–7%) mutation carriers.[58,59,94,95] There is insufficient evidence to recommend a particular surveillance strategy, but surveillance may be considered in selected individuals—including those with a family history of urothelial cancer or individuals with *MSH2* pathogenic variants (especially males), as they appear to be at higher risk. These groups may benefit from annual urinalysis starting at age 30 to 35 years.

Risk for pancreatic cancer and brain cancer is also elevated in Lynch syndrome.[72,73,88,96] However, the panel is unable to make a screening recommendation for pancreatic cancer at this time. If screening is performed for pancreatic cancer, the panel recommends that it should be considered at high-volume centers with multidisciplinary teams and preferably in the context of research protocols. The International Cancer of the Pancreas Screening (CAPS) Consortium recommends that patients with Lynch syndrome and one first-degree relative with pancreatic cancer should be considered for screening.[97] An annual physical and neurologic examination starting at age 25 to 30 years may be considered for central nervous system (CNS) cancers, but data to support this practice are lacking.

In addition, there have been suggestions of an increased risk for breast cancer in the Lynch syndrome population;[57,59,94,98] however, there is not enough evidence to support increased screening above average-risk breast cancer screening recommendations or those based on personal or family history of breast cancer (see NCCN Guidelines for Breast Cancer Screening and Diagnosis).[54,65] A study of 188 men with Lynch syndrome also showed a 5-fold increase in risk for prostate cancer.[99] However, there is insufficient evidence to recommend earlier or more frequent prostate cancer screening among males with Lynch syndrome.[54,65] Men with Lynch syndrome should be encouraged to participate in prostate cancer

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

screening as recommended in the NCCN Guidelines for Prostate Cancer Early Detection.

Reproductive Options (LS-4)

Patients of reproductive age should be advised regarding their options for prenatal diagnosis and assisted reproduction, including pre-implantation genetic diagnosis. This discussion should include known risks, limitations, and benefits of these technologies. If both partners are a carrier of a mutation(s) in the same MMR gene or *EPCAM* (eg, if both partners carry a mutation in the *PMS2* gene), then they should also be advised about the risk for constitutional MMR deficiency (CMMRD) syndrome, a rare recessive syndrome.[100]

Lynch Syndrome Colonoscopy Surveillance Findings and Follow-up (LS-5)

If there are no pathologic findings, continued surveillance every 1 to 2 years is recommended. If the patient is not a candidate for routine surveillance, subtotal colectomy may be considered, though generally extended surgery is limited to patients following CRC diagnosis. After subtotal colectomy, endoscopic surveillance of the rectum is required, at similar intervals as described above.

Patients with confirmed adenocarcinoma should be treated following the appropriate NCCN Guidelines for Treatment of Cancer by Site (available at www.NCCN.org). For patients with colorectal adenocarcinoma, depending on the clinical scenario, segmented or extended colectomy should be considered.

For patients with adenomatous polyps, recommendations include endoscopic polypectomy with a follow-up colonoscopy every 1 to 2 years. If the adenomas have high-grade dysplasia, recommendations include colonoscopic surveillance every 1 to 2 years, if an en bloc complete excision was performed, or surgery (segmental or extended colectomy), if resection was not en bloc or if dysplasia is involved in the resection margin. If an adenomatous polyp cannot be completely resected endoscopically, then segmental or extended colectomy may be done. Post-colectomy patients should be followed with lower endoscopic exams every 1 to 2 years.

The option of segmental or extended segmental colectomy for patients with confirmed adenocarcinoma and/or adenomatous polyps is based on individual considerations and discussion of risks. For example, the U.S. Multi-Society Task Force on Colorectal Cancer recommends that surgery in those older than 60 to 65 years and those with underlying sphincter dysfunction should potentially be less extensive.[35] Surgical principles for polyps are similarly controversial. Practically, a patient who is unable or unlikely to comply with frequent colonoscopy should be considered for more extensive colectomy, especially if young. Post-colectomy patients should be followed with examination of all remaining colonic mucosa every 1 to 2 years.

Chemoprevention in Lynch Syndrome

In the randomized CAPP2 trial, 861 participants with Lynch syndrome took either daily aspirin (600 mg) or placebo for up to 4 years; the primary endpoint was the development of CRC.[101] After a mean follow-up of 55.7 months, participants taking daily aspirin for at least 2 years had a 63% reduction in the incidence of CRC (incidence rate ratio [IRR], 0.37; 95% CI, 0.18–0.78; P = .008). These participants also saw protection from all Lynch syndrome cancers (IRR, 0.42; 95% CI, 0.25–0.72; P = .001). There was no protection seen for participants who completed <2 years of the intervention. Subgroup analyses from this trial showed that the association between obesity and CRC in patients with Lynch syndrome may be attenuated by taking daily aspirin.[102] However, limitations of the CAPP2 trial highlight the need for larger and long-term randomized trials in this area.[103,104] In an observational study including 1858 patients from the

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Colon Cancer Family Registry who have Lynch syndrome, aspirin use was associated with reduced risk for CRC, both for patients who took aspirin for 5 or more years (HR, 0.25; 95% CI, 0.10–0.62; *P* = .003) and patients who took aspirin between 1 month and 4.9 years (HR, 0.49; 95% CI, 0.27–0.90; *P* = .02), compared to those who took aspirin for less than 1 month.[105]

At this time, the panel suggests that aspirin may be used to prevent cancer in patients with Lynch syndrome, but it is emphasized that the optimal dose and duration of therapy are currently unknown. The CAPP2 trial used a dose of 600 mg per day,[101] though many clinicians who prescribe daily aspirin as chemoprevention in patients with Lynch syndrome utilize a lower dose. The CAPP3 randomized double-blind trial is currently examining the effects of low, moderate, and high doses of daily aspirin on Lynch syndrome-associated cancer incidence (NCT02497820), but results are not yet available. The panel's recommendation to consider aspirin for chemoprevention is consistent with the stance of the American Gastroenterological Association.[55] Due to limited mature data,[101,106] the American College of Gastroenterology does not recommend standard use of aspirin for chemoprevention.[65]

Adenomatous Polyposis Testing Criteria (POLYP-1)

Genetic testing for adenomatous polyposis is recommended when an individual has a personal history of ≥20 cumulative adenomas. Some studies have suggested genetic testing with a threshold of ≥10 cumulative adenomas.[71,107] Genetic testing is also recommended when an individual has a personal history of a known pathogenic variant in polyposis genes in the family.

Testing may also be considered if: 1) there is a personal history of a desmoid tumor, hepatoblastoma,[108] cribriform-morular variant of papillary thyroid cancer,[109,110] or multifocal/bilateral congenital retinal pigment epithelial hypertrophy (CHRPE);[65] or 2) the individual meets one of the following criteria for SPS: a) ≥5 serrated polyps proximal to the sigmoid colon with ≥2 of these being >10 mm; or b) ≥20 serrated polyps of any size distributed throughout the colon; or 3) the individual has a personal history of between 11 and 20 cumulative adenomas. Age of onset, family history, and/or presence of other features may influence whether genetic testing is offered in these situations.

A cross-sectional study of >7000 individuals found that the prevalence of pathogenic *APC* mutations was 80%, 56%, 10%, and 5% for those with ≥1000 adenomas, 100 to 999 adenomas, 20 to 99 adenomas, and 10 to 19 adenomas, respectively.[111] For the same groups, the prevalence of biallelic *MUTYH* mutations was 2%, 7%, 7%, and 4%. Notably, these prevalence estimates may be overestimates since data from this study were taken from a convenience sample of individuals referred for genetic testing to a testing provider, and not from consecutive patients with multiple adenomas. In a cross-sectional study of 3789 individuals with at least 10 colorectal polyps who underwent multi-gene panel testing, the prevalence of mutations in adenomatous polyposis genes decreased with increasing age in all polyp count groups (*P* < .001 for 10–19, 20–99, and ≥100 polyps).[107] In addition, the prevalence of mutations in all genes of interest remained above 5% in all age and polyp cohorts.[107] These data provide the rationale for recommending genetic testing for individuals with ≥20 cumulative lifetime adenomas, and considering genetic testing for those with >10 cumulative lifetime adenomas.

When colonic polyposis is present only in siblings, consider recessive inheritance. For example, MAP follows a recessive pattern of inheritance, so *MUTYH* testing can be performed prior to *APC* testing if a recessive pattern is apparent in the pedigree (eg, when family history is positive only for a sibling). If, on the other hand, a clear autosomal dominant inheritance pattern is observed, *MUTYH* testing is unlikely to be informative. In

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

addition, *MUTYH* testing is not indicated based solely on a personal history of a desmoid tumor, hepatoblastoma, or cribriform-morular variant of papillary thyroid cancer. Overall, the decision to order *APC*, *MUTYH*, or germline multi-gene testing including these genes should be at the discretion of the clinician.

If pathogenic variant(s) in the family is known, genetic testing for familial pathogenic variant is recommended. If there is no known pathogenic variant in any polyposis gene in the family, germline multi-gene testing is preferred, and the panel should include all polyposis and colorectal cancer genes.[107] Alternatively, when colonic polyposis is present in a single person with a negative family history, the panel recommends polyposis syndrome-specific testing (eg, for *de novo APC* or *MUTYH* pathogenic variants).

When a familial mutation is known (ie, deleterious *APC* pathogenic variant, monoallelic or biallelic *MUTYH* pathogenic variant, or other known pathogenic variant in another polyposis gene), genetic testing can be considered for at-risk family members. An at-risk family member can be defined as a sibling of an affected individual and/or proband. Siblings of a patient with MAP are recommended to have site-specific testing for the familial mutations. Other individuals in a family may also be at risk of having MAP or a monoallelic *MUTYH* pathogenic variant. Full sequencing of *MUTYH* may be considered in an unaffected parent when the other parent has MAP. If the unaffected parent is not tested, then comprehensive testing of *MUTYH* should be considered in the children. If the unaffected parent is found to have one *MUTYH* pathogenic variant, then testing the children for the familial *MUTYH* mutations is clinically indicated. Testing of children of *MUTYH* heterozygotes should be offered if the other parent is also a heterozygote or could still be offered if the other parent is not a heterozygote and management would change, if they have a first-degree relative affected with CRC, or to inform reproductive risks, since their future children could be at risk for MAP.

Among patients with concern for a polyposis syndrome, if no pathogenic variant is detected, further management should be based on family and personal history of CRC and polyps. Patients with no family and lower cumulative polyp burden (eg, fewer than 20 adenomas) may follow the NCCN Guidelines for Colorectal Cancer Screening. Individuals with either a family history of CRC or polyposis and/or a higher cumulative polyp burden may require additional testing based on personal family and medical history, and specialized management, such as described in a subsequent section (see *Colonic Adenomatous Polyposis of Unknown Etiology*). If genetic testing is not done, the individuals should also be surveilled and managed based on family and personal history of CRC and polyps and as described in a subsequent section (see *Colonic Adenomatous Polyposis of Unknown Etiology*).

Counseling should be provided for at-risk individuals so that they are able to make informed decisions about the implications involved in genetic testing, as well as the implications for their own management. Genetic testing in these individuals should be considered before or at the age of screening. The age for beginning screening should be based on the patient's symptoms, family phenotype, and other individual considerations. Fatal CRC is rare before the age of 18 years. If an individual at risk is found not to carry the mutation responsible for familial polyposis in the family, screening as an average-risk individual is recommended.

Surveillance and treatment recommendations depend on the performance and findings of genetic testing, as outlined below.

Familial Adenomatous Polyposis (FAP/AFAP-1)

Classical FAP and AFAP are autosomal dominant conditions characterized by a germline mutation in the *APC* gene, located on

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

chromosome 5q21.[112,113] Truncating mutation of the *APC* gene is detectable in about 80% of patients with FAP using protein-truncating tests.[114,115] Although FAP accounts for less than 1% of all CRC, it has been recognized as a paradigm for treating individuals at increased risk for cancer.

Diagnosis: Classical vs. Attenuated FAP

A clinical diagnosis of classical FAP is suspected with the early onset of at least 100 cumulative adenomas in the large bowel. Fewer than 100 adenomas may be observed in younger ages, especially in patients with a family history of FAP.[112] However, at older ages, patients often exhibit hundreds to thousands of colonic adenomas. The lifetime risk for cancer in individuals with classic FAP approaches 100% by the age of 50. Most of the resulting cancers occur in the left colon. Individuals with FAP also have an increased risk for other cancers, including duodenal cancer (4%–12%), hepatoblastoma (1%–2%, usually by age 5 years), thyroid cancer (<2%), and gastric cancer (0.5%–1.3%).[116,117] FAP is associated with increased malignancy risk in cribriform-morular variant, a rare form of papillary thyroid carcinoma.[109] Other possible associated findings of patients with FAP include desmoid tumors, which occur more frequently in patients with distal *APC* mutations, and CHRPE, which occurs in patients with mutations in the central portion of the gene.[108,118-120] Increasingly, family members are diagnosed at adolescence through genetic testing for their specific familial mutation or through sigmoidoscopic screening in the second decade of life.[121]

AFAP is a recognized variant of FAP characterized by a later onset of disease and fewer cumulative adenomas than observed with FAP, typically ranging from 10 to less than 100.[112,113] These adenomas are more prone to occur in the right colon and may take the form of diminutive sessile adenomatous polyps.[122] Phenotypic expression is often variable within families. The onset of CRC is typically delayed compared to patients with FAP,[123] but the incidence of cancer rises sharply after the age of 40 years and approaches 70% by age 80 years. Upper GI findings and thyroid and duodenal cancer risks are similar to that found in classical FAP.

However, there is currently no consensus on what constitutes a clinical diagnosis of AFAP and some patients may present with more than 100 adenomas. To confirm the diagnosis of FAP or AFAP, a germline mutation in *APC* must be identified (see *Adenomatous Polyposis Testing Criteria*, above). A family history may be negative, since approximately 30% of individuals have *de novo APC* germline mutations.[124,125]

Management of FAP and AFAP

It is recommended that physicians or centers with expertise in FAP should manage patients, and the management should be individualized based on genotype, phenotype, and other personal considerations. The surveillance interval should be adjusted according to the actual adenoma burden. Management of FAP includes early screening and colectomy or proctocolectomy after the onset of polyposis. Because cancer incidence in FAP rises dramatically early in the third decade of life, prophylactic proctocolectomy is usually indicated in the second decade of life. Management of AFAP includes early screening, with colectomy or proctocolectomy when the adenoma burden becomes significant and no longer manageable by polypectomy. Post-colectomy chemoprevention can also be considered (see below).

Preoperative surveillance schedules, surgical options, and surveillance following resection are discussed in more detail below.

Preoperative Surveillance for Individuals with a Family History of Classical FAP (FAP-4)

Management of individuals with a family history of FAP depends on whether the familial mutation is known or unknown (also see

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Adenomatous Polyposis Testing Criteria, above). When the mutation is unknown, an affected family member should have genetic counseling and testing, followed by counseling and testing of at-risk family members. If affected family members are unavailable, testing of at-risk individuals can be considered. When the familial mutation is known, genetic counseling and testing of at-risk family members is indicated. Genetic testing for FAP in children should be done by age 10 years when colon screening would be initiated. If there is intent to do hepatoblastoma screening, FAP genetic testing should be considered in infancy. Preoperative surveillance for at-risk individuals with a family history of FAP depends on genetic testing results, as described below.

Negative genetic testing: If an individual at risk is found not to carry the *APC* pathogenic variant responsible for familial polyposis in the family, screening as an average-risk individual is recommended.

Positive genetic testing: If an *APC* pathogenic variant is found, colonoscopy (preferred option) or flexible sigmoidoscopy every 12 months, beginning at 10 to 15 years of age, is recommended. If adenomas develop, surgical options should be reviewed (see below).

No genetic testing: Some people who undergo genetic counseling are determined to have a high risk for FAP, but decide, for a variety of reasons, not to undergo genetic testing, which influences how their screening is managed. These individuals are considered to be potentially at risk and should be offered annual colonoscopy (preferred option) or flexible sigmoidoscopy beginning at 10 to 15 years of age until the age of 24 years. If results continue to be negative, the following surveillance intervals are recommended: every 2 years for patients >24 to ≤34 years of age; every 3 years for patients >34 to ≤44 years of age; and every 3 to 5 years for patients older than 44 years of age.

There are several reasons why surveillance is recommended so often for these individuals. First, adenomas may begin to develop in adolescence. Most people with classic FAP present with adenomas before the age of 25 years, so annual surveillance with sigmoidoscopy will detect the majority of patients with FAP. Less often, people with FAP will not develop adenomas until a later age. The probability of FAP in a person without any adenomas on annual surveillance begins to decrease with age around this time, so that surveillance does not need to be as frequent between the ages of 24 and 34 years, and can be even less frequent between the ages of 34 and 44 years. This recommended schedule is more rigorous than screening guidelines for the general population, because serial negative examinations up to age 35 years do not exclude the diagnosis of FAP. It is important to recognize that individuals with attenuated polyposis may not present until a later age and may have fewer adenomas than those with classic FAP, yet enhanced surveillance is still warranted in these individuals. Notably, the lack of data to support precise intervals for surveillance in individuals from families with FAP is one key reason to pursue genetic testing of an affected individual within the family, since identification of a pathogenic mutation can allow for surveillance to rule in and rule out disease in unaffected relatives.

No familial mutation found: In some families, mutations cannot be found with available testing technology. The sensitivity to identify *APC* pathogenic variants is currently only about 70% to 90%.[126] Evaluating asymptomatic individuals at risk in these families presents a difficult problem. By far the best approach in this situation is additional attempts to identify the *APC* or *MUTYH* pathogenic variants in an affected family member, even if the available person is not a first-degree relative. If a mutation is found, then the at-risk individual should be managed similarly to those with known familial mutations. FAP can be excluded in a person at risk whose genetic testing results indicate no mutation is found when a

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

mutation has been previously identified in an affected family member (a "true negative" test result).

If, however, a familial mutation is still not identified, genetic testing of at-risk individuals can be considered. Certainly, a positive test in an asymptomatic person is informative even when the familial mutation has not been previously identified. However, interpreting a test in which "no mutation is found" in an asymptomatic person is not the same as a "negative test." This particular issue is often a source of confusion and misinterpretation. Thus, it is critical that patients receive appropriate genetic counseling to avoid false-negative interpretations of test results.[127] Surveillance for these at-risk individuals for whom no mutation is found is identical to that for untested individuals with known familial mutation (see section above). Again, if polyposis is detected, patients should be managed in the same way as those with a personal history of classical FAP.

Preoperative Surveillance for Individuals with a Personal History of AFAP (AFAP-1)

Treating patients with a personal history consistent with AFAP varies depending on the patient's age and adenoma burden. For young patients younger than age 21 years with a small adenoma burden (defined as fewer than 20 adenomas, all <1 cm in diameter and none with advanced histology), colonoscopy and polypectomy are recommended every 1 to 2 years with surgical evaluation and counseling if appropriate. In patients aged 21 years and older with a small adenoma burden, colectomy and ileorectal anastomosis (IRA) are alternative treatment options to colonoscopy and polypectomy that may be considered (see *Surgical Options in FAP and AFAP* below for further description of colectomy and IRA). Earlier surgical intervention should be considered in patients who are unlikely to be able to comply with frequent surveillance colonoscopy.

If adenoma burden is endoscopically unmanageable, colectomy with IRA is preferred in most cases. When polyposis becomes too significant to be managed by polypectomy (ie, when polyps number >20 at any individual examination or when a polyp ≥1 cm in diameter or with advanced histology is identified), proctocolectomy with ileal pouch anal anastomosis (IPAA) may be considered (see *Surgical Options in FAP and AFAP* below for further description).

Preoperative Surveillance for Individuals with a Family History of AFAP (AFAP-2)

Similar genetic counseling, testing, and surveillance considerations discussed previously for patients with a classical FAP family history apply to patients with a family history of AFAP, except for the endoscopy approach. It is important to recognize that individuals with attenuated polyposis may not present until a later age and may have fewer adenomas than those with classical FAP. However, enhanced surveillance is still warranted for these patients.

Negative genetic testing: If an individual at risk is found not to carry the *APC* pathogenic variant responsible for polyposis in the family, screening as an average-risk individual is recommended.

Positive genetic testing, no genetic testing, or no familial pathogenic variant found: In an individual at risk who is found to carry the APC pathogenic variant, colonoscopy surveillance should begin in the late teens, with repeat examinations every 1 to 2 years. If adenomas are detected, surveillance recommendations are as described for individuals with a personal history of AFAP. In the absence of a true negative genetic test result or if the individual is has not undergone genetic testing, an individual with a family history of AFAP should begin colonoscopy surveillance in the late teens, with repeat examinations every 2 to 3 years. Thus, the late onset and right colon involvement is accommodated in contrast to classical FAP. Individuals should continue with surveillance

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

until adenomas are found, at which point they should be managed as patients with a personal history of AFAP.

Surgical Options in FAP and AFAP (FAP-A)

Three different surgical options are available for individuals with classical FAP and AFAP: total proctocolectomy with IPAA (TPC/IPAA) (recommended for FAP), total abdominal colectomy with IRA (TAC/IRA) (recommended for AFAP), and TPC with permanent end ileostomy (TPC/EI).[128] The prime factors to consider when choosing an operation for FAP and AFAP are the personal and familial phenotype, including the rectal polyp burden (ie, distribution and number) and whether colon or rectal cancer is present at diagnosis. In patients presenting with the classical FAP phenotype, TPC/IPAA is generally recommended because it prevents both colon and rectal cancers. For patients with AFAP, TAC/IRA is generally recommended; TPC/IPAA can also be considered in cases of dense rectal polyposis not manageable with polypectomy. Surgery is performed either at the onset of polyposis or later, depending on the severity of the familial phenotype and genotype, the extent of polyposis at diagnosis, individual considerations, and local practices and expertise. Proper post-surgical surveillance should be followed as outlined in the sections below. In patients who are younger than 18 years without severe polyposis and without a family history of early cancers or severe genotype, the timing of proctocolectomy can be individualized. If surgery is delayed, then annual colonoscopy is recommended. Patients should be managed by physicians or centers with expertise in FAP, and management should be individualized to account for genotype, phenotype, and personal considerations.

Total Proctocolectomy with Ileal Pouch Anal Anastomosis:

TPC/IPAA, usually with a temporary loop ileostomy, is offered to patients with classical FAP, patients with AFAP with severe phenotypes resulting in carpeting of the rectum, patients with curable rectal cancer complicating the polyposis, and patients who underwent IRA and now have an unstable rectum in terms of polyp number, size, or histology. The operation is generally not offered to patients with incurable cancer, those with an intra-abdominal desmoid that may interfere with the completion of surgery, or patients who have an anatomic, physiologic, or pathologic contraindication to an IPAA. The advantages of this operation are that the risks of developing rectal cancer are negligible and a permanent stoma is not needed. The disadvantages are that it is a complex operation, a temporary stoma is usually needed, and it carries a small risk of bladder and sexual dysfunction after proctectomy. Functional results are variable. Bowel function, although usually reasonable, is also somewhat unpredictable. The ileal pouch requires surveillance, and the area of the IPAA should still be examined due to the imperfect nature of mucosectomy.

Total Abdominal Colectomy with Ileorectal Anastomosis: A TAC/IRA has an overall low morbidity rate. It generally results in good bowel function. Most patients have three to four bowel movements per day, and the risk of urgency or fecal incontinence is low. Without proctectomy, there should be no risk of problems with bladder or sexual function, or decreased fertility, and even a temporary stoma is obviated. The main disadvantages of TAC/IRA are increased risk for developing metachronous rectal cancer, associated morbidity and mortality, and the need to undergo subsequent proctectomy due to severe rectal polyposis.[129-131] A review of 659 patients in the Dutch-Scandinavian collaborative national polyposis registries who underwent colectomy with IRA found a high rate of advanced and fatal rectal cancers even though 88% of the patients underwent a diagnostic proctoscopy within 18 months of presentation. It was estimated that 12.5% of patients undergoing this procedure would die of rectal cancer by age 65 even if compliant with endoscopic screening.[131] The authors concluded that proctocolectomy is the preferred procedure for most patients with the classical FAP

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

phenotype, though some controversy remains regarding this choice. They and others also observed that patients could not reliably be selected for colectomy based on genotype alone. However, subsequent studies have reported that the risk for rectal cancer associated with TAC/IRA has declined since the 1980s when IPAA first became available for high-risk patients with severe polyposis.[132,133]

The choice of TAC/IRA versus TPC/IPAA centers on the issues of the relative quality of life.[134-139] A modest reduction in life expectancy is expected in patients with classical FAP with rectal preservation.[129,140] The decision to remove the rectum is dependent on whether the polyps are amenable to endoscopic surveillance and resection. Proctoscopic examination of a retained rectum is indicated annually. IRA is the surgery of choice for the majority of patients with AFAP who either have rectal sparing or endoscopically manageable rectal polyposis. In certain cases, such as AFAP with mainly proximal polyps, the extent of colectomy may be modified based on the burden of adenoma distribution and number. It is not recommended for patients with extensive rectal polyposis. Patients and families must be absolutely reliable for follow-up endoscopic examinations. The risk to the rectal stump rises considerably after age 50 years. If the rectum becomes unstable, a proctectomy with either an IPAA or EI is recommended.[141]

Total Proctocolectomy with Permanent End Ileostomy: A TPC/EI is rarely indicated as a prophylactic procedure because good options are available that do not involve a permanent stoma, which has implications for the patient and the family. Fear of a permanent stoma may make family members reluctant to undergo screening. The operation removes all risk for colon and rectal cancer, but is associated with the risk of bladder or sexual function disorders. This operation may be offered to patients with a low, locally advanced rectal cancer, patients who cannot have an ileal pouch due to a desmoid tumor, patients with a poorly functioning ileal pouch, and patients who have a contraindication to an IPAA (eg, concomitant Crohn's disease, poor sphincter function).

TPC with continent ileostomy is offered to patients who are motivated to avoid EI because they are either not suitable for TPC/IPAA or they have a poorly functioning IPAA. This is a complex operation with a significant risk for reoperation.

Surveillance Following Surgery for FAP (FAP-1)

Colorectal Cancer

Patients with retained rectum should undergo endoscopic rectal examination every 6 to 12 months, depending on polyp burden. If the entire colorectal tract has been removed, the ileal pouch should be evaluated endoscopically every 1 to 3 years, depending on polyp burden; this should be increased to every 6 months if large flat polyps with villous histology and/or high-grade dysplasia are found. If the patients had an ileostomy, consider careful visualization and stoma inspection by ileoscopy to evaluate for polyps or malignancy every 1 to 3 years, although the panel notes that evidence to support this recommendation is limited. Chemoprevention may also be considered (see discussion of *Chemoprevention in FAP and AFAP* below).

Duodenal or Periampullary Cancer (FAP-2)

A major component of surveillance in patients with a personal history of FAP or AFAP after surgery relates to the upper GI tract. Duodenal adenomatous polyps develop in greater than 90% of patients with FAP. These adenomatous polyps are classified into stages 0 to IV, as defined by Spigelman based on macroscopic and histologic criteria (FAP-3).[142] Duodenal cancer is uncommon before age 40 years, and rare before age 30 years. The cumulative lifetime risk of developing severe duodenal polyposis (stage IV) has been estimated to be approximately 35% (95%

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

CI, 25%–45%).[143] The risk for duodenal cancer increases dramatically with Spigelman stage IV disease.

Surveillance should be done with upper endoscopy (including complete visualization of the ampulla of Vater) and use of Spigelman's or other standardized staging, though efficacy of surveillance of these sites has not been demonstrated. Cap-assisted endoscopy may be adequate for visualization of the ampulla.[144] More intensive surveillance and/or treatment is required in patients older than 50 years with large or villous adenomatous polyps. The panel recommends that surveillance begin at approximately 20 to 25 years of age. If colectomy was done before age 20 years, then an earlier baseline upper endoscopy could be considered.

The appropriate period for follow-up endoscopy relates to the burden of polyps, varying from every 4 years if no polyps are found to every 3 to 6 months for Spigelman's stage IV polyposis. Surgical evaluation and counseling are recommended for stage IV polyps, invasive carcinoma, and high-grade dysplasia or dense polyposis that cannot be managed endoscopically. If surgery is deferred, expert surveillance endoscopy every 3 to 6 months is recommended. Endoscopic treatment options, when feasible, include endoscopic papillectomy in addition to excision or ablation of resectable large or villous adenomatous polyps and mucosectomy of resectable advanced lesions to potentially avert surgery (FAP-3). Potentially higher risk adenomas involving the papilla, including adenomas ≥1 cm in size or adenomas extending into the papilla, should be referred to an expert center for evaluation and management.

Other Cancers (FAP-2)

Fundic gland polyps (FGPs) of the stomach also occur in the majority of patients with FAP and AFAP and often are too numerous to count. In FAP, FGPs usually have biallelic inactivation of the *APC* gene, and often display foci of dysplasia or microadenomatous polyps of the foveolar epithelium.[145] However, malignant progression in FGPs is uncommon and the lifetime risk for gastric cancer in patients with FAP in Western countries is reported to be in the range of 0.5% to 1.3%.[116,117] The risk of gastric cancer in FAP patients appears to be increased in patients from geographic areas with high gastric cancer risk and may be elevated in the setting of certain endoscopic findings including carpeting of FGPs, solitary polyps >20 mm, and mounds of polyps.[116,146] High-risk histologic features include tubular adenomas, polyps with high-grade dysplasia, and pyloric gland of polyps.[147] In light of this, the panel recommends that the need for specialized surveillance or surgery should be considered in the presence of described high-risk histologic features,[65] preferably at a center of expertise. Patients with high-risk lesions that cannot be removed endoscopically should be referred to a specialized center for gastrectomy.

Patients with classical FAP also have elevated risk for developing other extracolonic cancers that may warrant surveillance (FAP-2).[148] Several studies suggest that there is an increased lifetime risk for developing thyroid cancer in FAP patients when compared to the general population, with incidence ranging from approximately 1% to 12%.[149-153] The mean age of diagnosis in these patients ranges from 29 to 33 years.[151,153] In addition, thyroid cancers in FAP are most commonly papillary and occur predominantly in women.[148,151,152,154] Although there is currently no high-level evidence to support thyroid cancer screening in FAP patients, some studies have found that screening with thyroid ultrasound has potential to detect thyroid cancers.

A retrospective analysis of 51 patients with a proven diagnosis of FAP demonstrated that out of 28 patients who had at least one screening ultrasound, 2 (7%) had papillary thyroid carcinoma.[151] Another study performed thyroid ultrasounds on FAP patients during their annual colonoscopy and found that out of 205 patients screened, 38% had thyroid cancer.[154] A concern regarding thyroid surveillance is potential for high

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

rates of benign thyroid nodule detection. In the aforementioned series, rates of thyroid nodule detection ranged from 51.7% to 79%.[151 154] Thus, the benefit of regular surveillance for thyroid cancer is uncertain and more studies may be necessary to develop optimal management. Currently the panel recommends an annual thyroid physical examination starting in the late teenage years. Annual thyroid ultrasound may be considered to supplement physical examination, although supportive data are lacking.

FAP is also associated with an increased risk for intra-abdominal desmoid tumors, the majority of which present within 5 years of colectomy. Since significant morbidity and mortality are associated with advanced desmoid tumors, early diagnosis is likely of benefit.[155] Although data to support screening and treatment are limited,[156,157] annual abdominal palpation during physical examination is advised. If family history of symptomatic desmoids is present, consider abdominal CT with contrast or MRI with or without contrast within 1 to 3 years post-colectomy, and then every 5 to 10 years. Immediate abdominal imaging is warranted if suggestive abdominal symptoms are present. For small bowel polyps and cancer, adding small bowel visualization to CT or MRI for desmoids as outlined above can be considered, especially if duodenal polyposis is advanced. However, there is a lack of high-level evidence to support routine small bowel screening distal to the duodenum. The risk for hepatoblastoma is much higher in young children with FAP.[108] Although the absolute risk is about 1.5%, given the lethality of the disease (25% mortality), active screening by liver palpation, abdominal ultrasound, and alpha-fetoprotein (AFP) measurements every 3 to 6 months during the first 5 years of life may be considered.

Medulloblastoma accounts for most of the brain tumors found in patients with FAP, predominantly in females younger than age 20 years.[158] The incidence of pancreatic cancer in FAP is not well defined and is likely very low. Giardiello and colleagues reported 4 retrospective cases (histology not documented) out of 1391 FAP-related subjects.[150] More studies are needed to elucidate the risk and benefit of screening for brain and pancreatic cancers, and there is no additional screening recommendation other than annual physical exam.

Surveillance After Surgery for AFAP (AFAP-1)

After surgery for AFAP, annual physical and thyroid examinations are recommended as for FAP. Surveillance of a retained rectum and the upper GI tract is similar to that for classical FAP.

Chemoprevention in FAP and AFAP (FAP-1/AFAP-1)

Aspirin has been shown to reduce the incidence and recurrence of colorectal adenomatous polyps in the general population.[159-164] Nonsteroidal anti-inflammatory drugs (NSAIDs) and aspirin have been shown in clinical trials to reduce recurrence of colorectal adenomatous polyps.

Cyclooxygenase-2 (COX-2) has been shown to be overexpressed in colorectal adenomatous polyps and cancers. The COX-2 inhibitor celecoxib is another NSAID that has been studied for its role in the chemoprevention of colorectal adenomatous polyps in the general population.[161,163,165-168] Results from the Prevention of Colorectal Sporadic Adenomatous Polyps (PreSAP) trial showed that the use of celecoxib significantly reduced the occurrence of colorectal adenomatous polyps within 3 years after polypectomy.[165] Similarly, the Adenoma Prevention with Celecoxib trial (APC trial) showed that in patients at high risk for CRC who had their polyps removed, celecoxib significantly lowered the formation of adenomatous polyps during a 3-year period.[168] Five-year safety and efficacy results of the APC trial showed that compared to placebo, the reduction in the incidence of advanced adenomatous polyps over 5 years was 41% for those who received the lower dose of celecoxib and 26% for patients who received the higher dose compared to the

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

control arm (both $P < .0001$).[169] However, due to the increased risk of cardiovascular events associated with their use, COX-2 inhibitors are not recommended routinely for sporadic adenomatous polyps.[170,171]

NSAIDs have also been studied for their role in chemoprevention in patients with FAP and AFAP. In a randomized, double-blind, placebo-controlled study, the NSAID sulindac did not prevent the development of colorectal adenomatous polyps in persons with FAP prior to surgical intervention.[172] In addition, a randomized controlled trial failed to show a strong benefit of chemoprevention with aspirin in young patients with FAP prior to surgical intervention, despite non-significant trends in reduced colorectal polyp size and number.[173] Thus, NSAIDs and aspirin may be as effective for chemoprevention in FAP. Some evidence suggests utility for NSAIDs when used in combination with other agents. Preclinical studies have demonstrated an association between COX-2 and the epidermal growth factor receptor (EGFR) signaling pathways and the development of intestinal tumorigenesis.[174-176] A double blind, randomized, placebo-controlled trial examined the effect of sulindac and erlotinib, an EGFR inhibitor, on duodenal adenomas in patients with FAP.[177] Participants with FAP were randomized to receive placebo (n = 46) or 150 milligrams (mg) of sulindac twice a day and 75 mg of erlotinib once a day (n = 46) for 6 months.[177] Over the course of 6 months, the median duodenal polyp burden increased in the placebo group and decreased in the sulindac/erlotinib group, with a net difference of -19.0 mm between the groups (95% CI, -32.0 to -10.9; $P < .001$).[177]

Chemoprevention with NSAIDs has also been studied following initial prophylactic surgery for both classical FAP and AFAP as an adjunct to endoscopic surveillance and to reduce the rectal polyp burden. Long-term use of sulindac may be effective in polyp regression and preventing recurrence of higher-grade adenomatous polyps in the retained rectal segment of patients with FAP.[178] In a randomized, double-blind, placebo-controlled study of 77 patients with FAP who had not had their entire colon and rectum removed, patients treated twice daily with 400 mg of celecoxib for 6 months had a 28% reduction in polyp number ($P = .003$) and a 31% decrease in sum of polyp diameters ($P = .001$), whereas patients receiving placebo had 4.5% and 4.9% reductions in those parameters, respectively.[179] It should be noted, however, that the FDA indication for use of celecoxib in FAP was removed in 2011 due to the lack of phase IV (follow-up) data.

A pilot study looked at a possible similar postoperative chemopreventive role in FAP and AFAP for the omega-3 polyunsaturated fatty acid, eicosapentaenoic acid (EPA).[180] Patients receiving EPA demonstrated a significant 22.4% decrease in polyp number and a significant 29.8% decrease in sum polyp diameter after 6 months of treatment, while patients in the placebo arm saw a worsening of global polyp burden during this time. However, the evidence is insufficient to recommend routine use, and a meta-analysis of N-3 polyunsaturated fatty acids intake and risk of CRC—not limited to FAP patients—did not show a clear protective association.

Overall, the panel notes that there are no FDA-approved medications for chemoprevention to facilitate management of the remaining rectum after surgery. While data suggest that sulindac, alone or combined with the EGFR inhibitor, erlotinib, may be a potent polyp-regression strategy,[172,177] addition studies with longer follow-up are needed to determine if the decrease in polyp burden decreases cancer risk.

MUTYH-Associated Polyposis (MAP-1)

MAP is an autosomal recessive hereditary syndrome that predisposes individuals to attenuated adenomatous polyposis and CRC.[181-183] It is caused by biallelic germline pathogenic variants in the *MUTYH* gene. *MUTYH* encodes the A/G-specific adenine DNA glycosylase excision

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

repair protein (also called hMYH), which is responsible for excising adenine nucleotides mismatched with 8-oxoguanine, a product of oxidative damage to DNA. Dysfunctional hMYH protein can thus result in G:C to T:A transversions during DNA replication. Adenomatous polyposis is thought to result from such transversions occurring within the *APC* gene. Individuals with MAP also have an increased risk for extracolonic tumors including duodenal cancer.[184]

Monoallelic carriers of *MUTYH* mutations may have a modest or slightly increased risk for CRC.[183,185-187] A study of 2332 relatives of patients with CRC with monoallelic *MUTYH* mutations showed that carriers have an estimated 2.5-fold increased risk for CRC, relative to the general population.[186] The estimated CRC risks, up to 70 years of age, were 7.2% for male carriers of monoallelic *MUTYH* mutations (95% CI, 4.6%–11.3%) and 5.6% for female carriers of monoallelic *MUTYH* mutations (95% CI, 3.6%–8.8%), irrespective of family history.[186] The risks for CRC were higher for carriers of monoallelic *MUTYH* mutations with a first-degree relative with CRC.[186] A study of 852 monoallelic *MUTYH* mutation carriers who were relatives of patients with CRC showed an increase in risk for CRC, relative to the general population (standardized incidence ratio [SIR], 2.04; 95% CI, 1.56–2.70; $P < .001$).[185] Another study evaluated the frequency of monoallelic *MUTYH* mutations and colorectal adenomas, and found that 13 of 72 individuals with CRC were monoallelic *MUTYH* mutation carriers, and 11 of the 13 had a family history of cancer in first or second-degree relatives.[188] In contrast, a population-based analysis of 198 monoallelic *MUTYH* mutation carriers showed that a monoallelic *MUTYH* mutation does not significantly increased CRC risk (OR, 1.07; 95% CI, 0.87–1.31; $P = .55$).[189] In addition, a meta-analysis of 945 articles investigating the associations between genetic variants and CRC risk determined that there is no substantial evidence supporting monoallelic *MUTYH* mutations and increased CRC risk.[190]

Given conflicting evidence regarding CRC risk associated with having a monoallelic MUTYH pathogenic variant,[186] the NCCN Panel recommends specialized screening for CRC mainly based on family history (GENE-7). Specifically, the panel recommends that monoallelic *MUTYH* carriers unaffected by CRC with a first-degree relative with CRC receive colonoscopy screening every 5 years beginning at age 40 or 10 years prior to first-degree relative's age at CRC diagnosis. Notably, these are consistent with standard NCCN recommendations based on having a first-degree relative with CRC alone (see NCCN Guidelines for Colorectal Cancer Screening). There are no specific data available to determine screening recommendations for a patient with one *MUTYH* pathogenic variant and a second-degree relative affected with CRC (see NCCN Guidelines for Colorectal Cancer Screening). Data are unclear as to whether specialized screening is warranted for *MUTYH* monoallelic carriers unaffected by CRC with no family history of CRC.[191] For monoallelic *MUTYH* carries with CRC, it is recommended that colonoscopy screenings occur at 1 year post-CRC resection. If an advanced adenoma is found, repeat annual screening. If there are no advanced adenomas detected, repeat at 3 years and then every 5 years. These recommendations are consistent with standard NCCN recommendations for surveillance of sporadic CRC (see NCCN Guidelines for Colon Cancer and the NCCN Guidelines for Rectal Cancer).

Most individuals with MAP generally have fewer than 100 adenomas, although a minority can present with greater than 1000. Hyperplastic polyps, sessile serrated polyps (SSPs), and traditional serrated adenomas may also be seen in this setting. In fact, some patients with MAP may also meet the criteria for SPS. The lifetime risk for CRC for patients with MAP may be very high.[192] The median age of presentation is approximately 45 to 59 years. While duodenal polyposis is reported less frequently in MAP than in FAP, duodenal cancer occurs in about 5% of patients with MAP. In

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

addition, individuals with MAP generally require colectomy at a later age than those with FAP.

Preoperative and Surgical Management of MAP (MAP-2/-3)

Genetic counseling and testing is recommended for individuals with a family history of MAP and known *MUTYH* pathogenic variants (see *Adenomatous Polyposis Testing Criteria*, above). With positive genetic testing (biallelic *MUTYH* pathogenic variants) or no testing in such individuals, surveillance colonoscopy should begin at age 25 to 30 years and should be repeated every 1 to 2 years if negative. If polyps are found, these patients should be managed as those with a personal history of MAP (see below). Upper endoscopy (including complete visualization of the ampulla of Vater) can also be considered beginning at age 30 to 35 years, with follow-up as described above for patients with a personal history of FAP.

Genetic counseling and testing is recommended for patients with multiple adenomatous polyps (see *Adenomatous Polyposis Testing Criteria*, above). Such individuals who have a negative test for *MUTYH* pathogenic variant should be managed individually as patients with FAP.

Individuals younger than 21 years of age with confirmed biallelic *MUTYH* pathogenic variants and small adenoma burden are followed with colonoscopy and complete polypectomy every 1 to 2 years. Surgical evaluation and counseling are also recommended if appropriate. Colectomy and IRA may be considered as the patient gets older. Surgery in the form of colectomy with IRA is recommended in most cases of significant polyposis not manageable by polypectomy. Proctocolectomy with IPAA can be considered in cases of dense rectal polyposis not manageable by polypectomy. Extent of colectomy may be modified based on adenoma burden (distribution and number).

Postoperative Surveillance in MAP (MAP-2)

After colectomy with IRA, endoscopic evaluation of the rectum every 6 to 12 months is recommended, depending on polyp burden. The use of chemoprevention can facilitate management of the remaining rectum postsurgery, although there are no FDA-approved medications for this indication at the present time. While there are data suggesting that sulindac is the most potent polyp-regression medication,[172] it is not known if the decrease in polyp burden decreases cancer risk.

In addition to evaluation of the rectum, an annual physical exam is recommended, with baseline upper endoscopy (including complete visualization of the ampulla of Vater) beginning at age 30 to 35 years. Cap-assisted endoscopy may be adequate for visualization of the ampulla.[144] Follow-up of duodenoscopic findings is as described above for patients with FAP (see FAP-3).

Peutz-Jeghers Syndrome (PJS-1)

PJS is an autosomal dominant condition mainly characterized by hamartomatous gastrointestinal (GI) polyps.[193] PJS polyps tend to be large and pedunculated, and have a characteristic histology showing broad bands of smooth muscle fibers (often in a tree-like configuration), chronic inflammation, edema, and fibrosis within the lamina propria and dilated glands.[194] Medical treatment if often sought due to complications that arise from the polyps (eg, obstruction, bleeding). PJS polyps tend to be accompanied with freckling or hyperpigmentation on the lips, buccal mucosa, vulva, fingers, and toes, which appears early in life but tends to fade during adulthood.[193] Besides being associated with an increased risk for CRC, PJS is also associated with increased risk for cancers of the breast, pancreas, ovary, and gallbladder.[195-198] A study of 33 patients with PJS in the United Kingdom showed that the risk of developing any cancer by age 65 years is 37% (95% CI, 21%–61%).[199] In a study of 72 patients with PJS, 12.5% had a GI malignancy.[198] The majority of PJS cases occur

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

due to pathogenic variants in the *STK11 (LKB1)* gene.[200,201] However, other genetic mutations may be involved, as an estimated half of patients with PJS do not have detectable *STK11/LKB1* mutations.[199]

A PJS clinical diagnosis is made when an individual has at least two of the following: two or more PJS-type polyps of the GI tract; mucocutaneous hyperpigmentation of the mouth, lips, nose, eyes, genitalia, or fingers; or family history of PJS. This is consistent with the statement from the American College of Gastroenterology regarding genetic testing and management of hereditary syndromes associated with CRC.[65] Since PJS is rare, referral to a specialized team or centers with expertise is recommended.

Management of Peutz-Jeghers Syndrome (PJS-2)

As there are limited data regarding the efficacy of various screening modalities in PJS, panel recommendations were made while taking into consideration cancer risk in PJS and the known utility of the specific screening modalities. Individuals with PJS should receive a colonoscopy every 2 to 3 years, beginning in the late teens.[202] To screen for breast cancer, a mammography and breast MRI should be done annually with a clinical breast exam conducted every 6 months, beginning at approximately age 25 years. For surveillance for gastric cancer, upper endoscopy should be done every 2 to 3 years beginning in the late teen years. For small intestinal cancers, small bowel visualization should be performed with CT or MRI enterography or video capsule endoscopy baseline at ages 8 to 10 years with follow-up interval based on findings but at least by age 18 years. Repeat imaging may then occur every 2 to 3 years (though this may be individualized). A repeat small intestinal exam is also indicated at any time based on symptoms. To monitor for cancer of the pancreas, magnetic resonance cholangiopancreatography with contrast (ideally performed at a center of expertise) or endoscopic ultrasound should be done every 1 to 2 years beginning in one's early 30s. Based on clinical judgment, an earlier age of initiation may be considered, such as 10 years younger than the earliest age of onset in the family. To monitor for gynecologic cancer, a pelvic exam and Pap smear should be done annually, beginning around ages 18 to 20 years. In males, annual testicular exam and observation for feminizing changes should be done beginning at around age 10 years. For lung cancer, education should be provided about symptoms and smoking cessation, if necessary. No other specific recommendations have been made for lung cancer. The panel's recommendations for screening of extracolonic cancers in patients with PJS reflect recommendations from the American College of Gastroenterology.[65]

Juvenile Polyposis Syndrome (JPS-1)

JPS is an autosomally dominant condition that is characterized by multiple hamartomatous polyps of the colon and rectum that usually manifests during childhood. Colonic polyps tend to be right-sided,[203] and 90% of patients present with bleeding and/or anemia.[204] Histologically, polyps from patients with JPS are exophytic and eroded, and contain marked edema and inflammation within the lamina propria, cystic glands filled with thick mucin, and some degree of smooth muscle proliferation.[194] Though patients with JPS are usually diagnosed during adolescence, it is a heterogeneous condition in that symptom intensity and age at diagnosis vary across patients.[205] About 50% to 64% of JPS cases occur due to mutations in the *BMPR1A* and *SMAD4* genes.[202,203] If there is a known *SMAD4* mutation in the family, genetic testing should be done within the first 6 months of life due to risk of hereditary hemorrhagic telangiectasia.[206] In a retrospective review of 44 patients with JPS from a polyposis registry in the United Kingdom, 9% had telangiectasia or vascular abnormalities.[203] Family history of juvenile polyposis is present in about half of patients with JPS.[204] Though lifetime risk for CRC has been difficult to estimate, a review of a large JPS kindred (117 members) provided an estimate of a 50% risk of GI malignancy.[207] The large number of polyps often found in

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

JPS increases the risk of malignancy.[204] In a separate review of 218 patients with juvenile polyposis, malignancy developed in 17% of patients.[204] The mean age of cancer diagnosis in this sample was 33.5. Out of the 36 malignancies that developed, 4 were not resectable, 7 were poorly differentiated, and 4 were metastatic.

A clinical diagnosis is made if at least one of three criteria is met: 1) at least five juvenile polyps of the colon; 2) multiple juvenile polyps found throughout the GI tract; and 3) at least one polyp in an individual with a family history of JPS.[65,208,209]

Management of Juvenile Polyposis Syndrome

Since JPS is rare, referral to a specialized team is recommended. Further, there are limited data regarding the efficacy of various screening modalities in JPS, so panel recommendations were made while taking into consideration cancer risk in JPS and the known utility of the specific screening modalities.

CRC screening via colonoscopy should begin around age 15 years, since the mean age of a juvenile polyp undergoing adenomatous changes is 18.6 years.[204] If polyps are found, colonoscopy should be repeated annually. If no polyps are found, then colonoscopy is recommended every 2 to 3 years. Screening for stomach cancer should also begin at age 15 years. An upper endoscopy screening schedule should match that of the colonoscopy screening schedule (ie, annually if polyps are found, every 2–3 years if no polyps are found). In families without an identified genetic mutation, consider substituting endoscopy every 5 years beginning at age 20 and every 10 years beginning at age 40 in patients in whom no colon or stomach polyps are found. In patients with gastric polyps, management issues related to anemia from giant confluent polyps may occur. In severe cases, if anemia develops requiring blood transfusion due to many gastric polyps, gastrectomy can be considered. The panel has made no recommendations regarding surveillance of the small intestine and the pancreas, since cancer of these organs in patients with JPS is rare and/or undefined, though the American College of Gastroenterology recommends screening of the small intestine.[65]

Serrated Polyposis Syndrome (SPS-1)

Serrated polyps include hyperplastic polyps, sessile serrated adenomas/polyps, and traditional serrated adenomas.[210] SSPs are flat or slightly raised and usually occur on the right side, while traditional serrated adenomas are generally polyploid.[211] Serrated polyps are more difficult to detect during colonoscopy and account for a disproportionate amount of interval cancers.[212] These polyps are considered premalignant, may account for as many as a third of CRCs, and should be managed similarly to adenomas.[212] Serrated polyps are thought to progress to cancer via pathways that are different from those in adenomas and to have an unfavorable prognosis.[211,213-215]

A clinical diagnosis of SPS (previously known as hyperplastic polyposis) is considered if at least one of the following criteria previously established by the WHO are met: 1) at least 5 serrated polyps proximal to the sigmoid colon, 2 or more greater than 10 mm; 2) at least one serrated polyp proximal to the sigmoid colon and a first-degree relative with serrated polyposis; or 3) at least 20 serrated polyps throughout the colon.[210] At the time of publication of the 2019 Discussion update, WHO issued updated criteria for diagnosis of SPS as follows: 1) ≥5 serrated lesions/polyps proximal to the rectum, all being ≥5 mm in size, with ≥2 being ≥10mm in size; 2) >20 serrated lesions/polyps of any size distributed throughout the large bowel, with ≥5 being proximal to the rectum.[216] There may be other clinical scenarios (eg, patient has between 5–10 serrated polyps or polyps are <1 cm) that increase CRC risk and may require additional evaluation per clinical judgment.[217] Individuals with SPS have an increased risk for colon cancer, though data on CRC risk for patients with SPS are

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

limited.[218,219] One retrospective study found that 35% of patients developed CRC during a mean follow-up period of 5.6 years (0.5–26.6 years).[218] In 6% of the patients, CRC was found during surveillance in diminutive polyps (4–16 mm) after a median interval of 11 months. In a retrospective cohort study examining 52 individuals who met criteria for serrated polyposis, 82% had colorectal adenomas, 16% had a personal history of CRC, and 37% had a family history of CRC.[220] Another retrospective analysis of 64 patients with serrated polyposis showed an SIR of 18.72 (95% CI, 6.87–40.74) for CRC.[221] For the majority of patients with SPS, no causative gene is identifiable. Emerging evidence links pathogenic variants in *RNF43*, a regulator of *ATM*/*ATR* (ataxia-telangiectasia mutated/ataxia-telangiectasia and Rad3-related protein) DNA damage response, to SPS.[222-225] Whole exome sequencing of 20 unrelated individuals with multiple sessile serrated adenomas (16 who fulfilled WHO criteria of SPS) led to the identification of nonsense mutations in *RNF43* in two individuals.[222] The *RNF43* mutations were associated with multiple serrated polyps (OR, 3.0; 95% CI, 0.9–8.9; $P = .04$).[222] One study identified a germline *RNF43* mutation in 1 out of 4 families with serrated polyposis, but more research is needed to understand prevalence of RNF43 mutations in patients with SPS.[225]

Management of Serrated Polyposis (SPS-1)

Colonoscopy with polypectomy is recommended for all polyps ≥5 mm, every 1 to 3 years depending on size and number of polyps, consistent with recommendations by the American College of Gastroenterology.[65] It may not always be possible to remove all polyps. Colonoscopic surveillance with consideration of surgical referral is recommended if colonoscopic treatment and/or surveillance is inadequate or if high-grade dysplasia occurs.[65]

Management of First-Degree Relatives (SPS-1)

The risk for CRC is elevated in first-degree relatives of individuals with SPS.[226-228] One study that compared CRC incidence in 347 first-degree relatives of patients with SPS to that in the general population (Eindhoven Cancer Registry) found 27 cases compared to an expected 5 cases (rate ratio [RR], 5.4; 95% CI, 3.7–7.8; $P < .001$).[226] In addition, this study found that four first-degree relatives satisfied the criteria for serrated polyposis (projected RR, 39; 95% CI, 13–121), suggesting a hereditary basis in some cases. Another multinational retrospective study found a similar increase in risk for CRC in both first- and second-degree relatives of patients with SPS.[228] In addition, an increased risk for pancreatic cancer was observed. In a prospective study, 76% of first-degree relatives of patients with SPS were found to have SPS at colonoscopy.[229]

The panel considers it reasonable to screen first-degree relatives at the youngest age of onset of SPS diagnosis, 10 years earlier than earliest diagnosis of CRC in the family, or by age 40 years, whichever is earliest. Subsequent screening is per colonoscopic findings or every 5 years if no polyps are found.

Colonic Adenomatous Polyposis of Unknown Etiology (CPUE-1)

When genetic testing in an individual with colonic adenomatous polyposis does not diagnose a pathogenic variant in a polyposis gene, surveillance should be tailored based on individual and family risk assessment. If the patient has a history of ≥100 adenomas, the panel recommends that the patient be managed as described above for patients with a personal history of classical FAP.

If the patient has a history of 11 to 20 adenomas, management should be based on clinical judgment, taking into account number, size, and type of polyps, as well as family history.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

If the patient has a history of >20 but <100 adenomas, and the adenoma burden is small and considered to be manageable by colonoscopy and polypectomy, the panel recommends colonoscopy and polypectomy every 1 to 2 years. Clearing of all polyps is recommended and can be repeated at short intervals if residual polyps are present. An upper endoscopy (including complete visualization of the ampulla of Vater) at baseline may be considered,[230,231] and repeated following duodenal surveillance guidelines as described above for patients with FAP (see FAP-3). Cap-assisted endoscopy may be adequate for visualization of the ampulla.[144]

If the patient has a history of >20 but <100 adenomas, but the adenoma burden is dense and considered unmanageable by polypectomy, the panel recommends a subtotal colectomy. A proctocolectomy may be considered if there is a dense rectal polyposis that cannot be managed by polypectomy.

In patients with a family history of ≥100 adenomas diagnosed at age <40 years in a first-degree relative, there are limited data to support recommendations for when to initiate screening or the interval of screening. The panel suggests consideration for colonoscopy screenings to begin at age 10 to 15 years with the following intervals post initial screen: every 1 year until age 24 years, every 2 years from 24 to 34 years, every 3 years from 34 to 44 years, and then every 3 to 5 years thereafter. If polyposis is detected, the panel recommends that patients be managed as described above for patients with a personal history of classical FAP. In addition, the panel recommends genetic testing for family members affected with polyposis.

In patients with a family history of >20 to <100 adenomas in a first-degree relative, there are limited data to suggest definitive recommendations for when to initiate screening or the interval of screening. The panel suggests considering colonoscopy screenings and polypectomy every 3 to 5 years starting at the same age as the youngest diagnosis of polyposis in the family if uncomplicated by cancer or by age 40 years, whichever is earliest. If multiple polyps are found during screenings, the interval for colonoscopies should occur every 1 to 3 years, depending on the type, number, and size of polyps. As described above, the panel recommends genetic testing for family members affected with polyposis.

In patients with a family history of >100 adenomas diagnosed at age ≥40 years in a first-degree relative, there are limited data to support recommendations for when to initiate screening or the interval of screening. The panel suggests considering colonoscopy screenings and polypectomy every 2 to 3 years starting at age 40 years if uncomplicated by cancer. If multiple polyps are found during screenings, the interval for colonoscopies should occur every 1 to 3 years, depending on the type, number, and size of polyps. As described above, the panel recommends genetic testing for family members affected with polyposis.

Multi-Gene Testing (GENE-1)

NGS allows for the sequencing of multiple genes simultaneously. This is referred to as multi-gene testing. The introduction of multi-gene testing for hereditary forms of cancer has rapidly altered the clinical approach to testing at-risk patients and their families. Multi-gene testing simultaneously analyzes a set of genes that are associated with a specific family cancer phenotype or multiple phenotypes. Multi-gene testing may include syndrome-specific tests (ie, panels that test for only one syndrome like Lynch syndrome, adenomatous polyposis), cancer-specific tests (ie, panels that test for more than one gene associated with a specific type of cancer like CRC), and comprehensive cancer panels (ie, panels that test for more than one gene associated with multiple cancers or cancer syndromes).

Multi-gene testing can include only high-penetrance genes associated with a specific cancer, or both high- and moderate-penetrance genes.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Comprehensive cancer risk panels, which include a large number of genes associated with a variety of cancer types, are also available.[232] The decision to use multi-gene testing for patient care should be no different than the rationale for testing a single gene known to be associated with the development of a specific type of cancer. Testing is focused on identifying a mutation known to be clinically actionable; that is, whether the management of an individual patient is altered based on the presence or absence of a mutation. Multi-gene testing may be most useful when more than one gene can explain a patient's clinical and family history. In these cases where more than one gene mutation could potentially influence a condition, multi-gene testing may be more efficient and/or cost-effective.[232] Multi-gene testing with panels that include genes associated with Lynch syndrome, as well as other highly penetrant genes associated with CRC, may be cost-effective,[233] and this approach may detect mutations not found in single-gene testing.[234] Multi-gene testing may also be considered for those who tested negative (indeterminate) for one particular syndrome, but whose personal and family history is strongly suggestive of an inherited susceptibility.[232,235]

A major dilemma regarding multi-gene testing is that there are limited data and a lack of clear guidelines regarding degree of cancer risk associated with some of the genes assessed in multi-gene testing, and how to communicate and manage risk for carriers of these genes.[235-237] This issue is compounded by the low incidence rates of hereditary disease, leading to a difficulty in conducting adequately powered studies.[236] Some multi-gene tests may include low- or moderate-penetrance genes, for which there are little available data regarding degree of cancer risk and guidelines for risk management.[232,237-240] Further, it is possible that the risks associated with these genes may not be due entirely to that gene only, but may be influenced by gene/gene or gene/environment interactions. Multi-gene tests also increase the likelihood of detecting VUS,[232,235,237,240-243] with likelihood rates ranging from 17% to 38%.[238,240,241,244] The considerable possibility of detecting a VUS adds to the complexity of counseling following multi-gene testing. However, as multi-gene testing is increasingly used, the frequency of a VUS being detected is expected to decrease. In addition, many VUS previously identified through hereditary cancer testing have been reclassified and downgraded to benign or likely benign categories.[10,245] Nonetheless, clinical phenotypic correlation is warranted with further discussion with the testing laboratory if evidence supports potential pathogenicity of a VUS. Patient and provider guidelines for follow-up of VUS have been developed.[246,247]

There are other issues to consider regarding multi-gene testing. First, commercially available tests may differ significantly on a number of factors, such as number of genes analyzed, turnaround time, and insurance coverage, among others. Tests requiring a longer turnaround time may not be suitable for patients who need rapid results. The specific laboratory and multi-gene test should be chosen carefully.[232] Second, in some cases, NGS may miss some mutations that would have been detected with traditional single-gene analysis.[232] Third, mutations identified for more than one gene add complexity that may lead to difficulty in making risk management recommendations.[235] A management plan should only be developed for identified gene mutations that are clinically actionable; care should be taken to ensure that overtreatment or over-screening does not occur due to findings for which clinical management is uncertain, or findings that are incorrectly interpreted due to lack of evidence.

Multi-gene testing is a new and rapidly growing field, but there is currently a lack of evidence regarding proper procedures and risk management strategies that should follow testing, especially when mutations are found for moderate-penetrance genes and when a VUS is found. For this reason, the NCCN Panel recommends that multi-gene testing be offered in the context of professional genetic expertise, with

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

pre- and post-test counseling being offered. Panel recommendations are in agreement with recommendations by ASCO, which issued an updated statement regarding genetic testing in 2015.[248] Carriers of a genetic mutation should be encouraged to participate in clinical trials or genetic registries.

Multi-gene testing is not recommended when: 1) there is an individual from a family with a known mutation and there is no other reason for multi-gene testing; 2) the patient's family history is strongly suggestive of a known hereditary syndrome; and 3) the patient is diagnosed with CRC with MSI or loss of one or more DNA MMR proteins. In these three scenarios, syndrome-specific panels may be considered.

Multi-gene testing may be considered (but may not be limited to based on clinical judgment) the following scenarios:

- A patient has a personal or family history that meets criteria for more than one hereditary cancer syndrome (eg, Lynch syndrome and *BRCA*-related breast and/or ovarian cancer)
- Colonic polyposis with uncertain histology
- Adenomatous polyposis (specific to *APC, MUTYH, GREM1, NTHL1, POLE,* and *POLD1*)
- Family history does not meet criteria for established testing guidelines, but there is suspicion of hereditary cancer, and an appropriate panel is available
- Family history is limited or unknown, but patient has concerns about hereditary cancer
- As second-line testing when first-line testing is inconclusive

Emerging evidence has identified additional genes that may be associated with increased risk for CRC, and the panel has evaluated the strength of the evidence based on published reports. Although research has demonstrated a potential risk for CRC associated with these mutations or pathogenic variants, the value of including these genes for clinical testing (eg, as part of a multi-gene panel) remains uncertain. Nonetheless, the panel recognizes that many testing companies offer panels that include these genes, and that patients are being tested and may need guidance regarding subsequent screening and surveillance. Accordingly, while the panel recommends caution in recommending multi-gene testing, guidance on management of results is discussed below.

Evidence to support screening and surveillance is limited, but the panel has conditionally developed a framework of recommendations for genes commonly included in multi-gene panels, which are outlined after a brief discussion of relevant data.

APC I1307K Pathogenic Variant

The adenomatous polyposis coli (*APC*) gene is a tumor-suppressor gene associated with CRC.[249] There is well-established evidence that the I1307K polymorphism in the *APC* gene, which occurs in approximately 6% to 8% of individuals of Ashkenazi Jewish decent, predisposes carriers to CRC.[250-254] In an analysis of 3305 individuals from Israel who underwent colonoscopic examinations, 8% were identified as carriers of the I1307K polymorphism, and the overall adjusted OR for colorectal neoplasia among carriers was 1.51 (95% CI, 1.16–1.98).[250] A subgroup analysis found that the prevalence of the I1307K polymorphism in individuals of Ashkenazi Jewish descent was 10.1% and the adjusted OR was 1.75 (95% CI, 1.26–2.45).[250] A meta-analysis including 40 studies showed that compared to carriers of wild-type I1307K, individuals of Ashkenazi Jewish descent who carried the I1307K polymorphism had a significantly increased risk of colorectal neoplasia, with a pooled OR of 2.17 (95% CI, 1.64–2.86).[253] Some studies have identified the I1307K polymorphism in the *APC* gene in individuals of non-Ashkenazi Jewish and Arabic descent, though the prevalence is higher in individuals of

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Ashkenazi Jewish descent.[255-257] An analysis of 900 cases from a population-based case-controlled study in northern Israel found the I1307K polymorphism in the *APC* gene in 78 colorectal cancer cases, with a prevalence of 11.2%, 2.7%, or 3.1% among individuals of Ashkenazi Jewish, non-Ashkenazi Jewish, or Arabic descent, respectively.[256] Overall, however, there is insufficient evidence to determine whether risk for CRC associated with the *APC* I1307K polymorphism differs among individuals with versus without Ashkenazi Jewish descent, and the panel recognizes that some individuals may not be aware of Ashkenazi Jewish heritage.

For carriers of the *APC* I1307K pathogenic variant with CRC, the panel recommends colonoscopy surveillance based on the NCCN Guidelines for Colon Cancer and the NCCN Guidelines for Rectal Cancer. For carriers of the *APC* I1307K pathogenic variant unaffected by CRC with a first-degree relative with CRC, the panel recommends colonoscopy surveillance every 5 years beginning at age 40 or 10 years prior to the first-degree relative's age at CRC diagnosis. For carriers unaffected by CRC without a first-degree relative with CRC, the panel recommends colonoscopy screening every 5 years beginning at age 40 years.

AXIN2 Mutations

Mutations in the Axin-related protein (*AXIN2*) gene are associated with polyposis and oligodontia.[258-262] In a study of a four-generation family from Finland, 11 family members had oligodontia and eight of them had either CRC or precancerous lesions, attributed to a nonsense mutation in the *AXIN2* gene.[258] Other studies support the association of *AXIN2* mutations and oligodontia.[260,262] A report described a family with an inherited *AXIN2* mutation (c.1989G>A) segregating in an autosomal dominant pattern with oligodontia and other findings including colonic polyposis, gastric polyps, a mild ectodermal dysplasia phenotype, and early-onset colorectal and breast cancers.[260] A study of 23 families with FAP resulted in the identification of a novel *AXIN2* variant (c.1387C>T) in one family with attenuated polyposis.[261] Carriers of the variant had a variable number of polyps, but no oligodontia or ectodermal dysplasia.[261] For carriers of *AXIN2* mutations, the panel recommends initiation of colonoscopic surveillance at ages 25 to 30 years and if no polyps are detected, to repeat colonoscopy every 2 to 3 years. If polyps are found, colonoscopic surveillance every 1 to 2 years is recommended, with consideration of surgical interventions if the polyp burden becomes unmanageable by colonoscopy.

CHEK2 Mutations

Germline mutations in the cell cycle checkpoint kinase 2 (*CHEK2*) gene are associated with increased risk of breast cancer and CRC, though heterogeneity may exist based on type of *CHEK2* pathogenic variant.[263-266] In a population-based study of 5953 patients with breast, prostate, and colon cancer (1934 patients had colon cancer), 533 were *CHEK2*-positive and 431 were affected relatives.[263] After adjusting for mutation type, the risk of colon cancer was higher among relatives of probands with colon cancer than among relatives of patients with prostate or breast cancer (HR, 4.2; 95% CI, 2.4–7.8; P = .0001).[263] Significant associations between *CHEK2* mutations and CRC risk have been identified in meta-analyses.[265,266] A meta-analysis of seven studies, including 4029 cases and 13,844 controls based on search criteria, found a significant association between the *CHEK2* I157T variant and CRC risk.[265] For carriers of *CHEK2* mutations, the panel recommends similar management strategies as described for carriers of the *APC* I1307K mutation. Some patients may elect for less aggressive screening based on shared decision-making. One model has suggested that earlier screening than the average-risk initiation may be justified for *CHEK2* 1100delC and I157T carriers based on reaching the same risk for CRC at an earlier age than observed among average-risk persons initiating screening at age 50 years.[191]

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

GREM1 Alterations

Hereditary mixed polyposis syndrome (HMPS) is a rare, autosomal-dominant condition that occurs primarily in individuals of Ashkenazi Jewish descent and is characterized by multiple types of colorectal polyps, extracolonic tumors, onset of polyps in adolescence, and progression of some polyps to advanced adenomas.[267,268] HMPS is due to a 40 kb duplication upstream of the gremlin 1 gene (*GREM1*), which increases ectopic *GREM1* expression in normal epithelium.[267] Exome sequencing combined with linkage analyses and detection of copy-number variations identified a 16 kb duplication upstream of *GREM1* in a family of non-Ashkenazi Jewish descent with AFAP.[269] For carriers of *GREM1* alterations, the panel recommends similar management strategies as described for carriers of *AXIN2* mutations.

MSH3 Biallelic Pathogenic Variants

MutS homolog 3 (*MSH3*) is a DNA MMR gene implicated in tumorigenesis of colon cancer with MSI.[270] Some data have linked biallelic *MSH3* germline mutations as a recessive subtype of colorectal adenomatous polyposis.[271] However, given available data, the panel agreed that the strength of evidence linking *MSH3* to increased CRC risk is not currently well established. For carriers of two *MSH3* pathogenic variants, the panel recommends similar management strategies as described for carriers of *AXIN2* mutations.

NTHL1 Biallelic Pathogenic Variants

The endonuclease III-like 1 (*NTHL1*) gene is involved in base excision repair and acts on oxidized pyrimidine residues.[272] A study suggests a role for *NTHL1* mutations in colorectal polyposis.[273] Whole-exome sequencing on 51 individuals from 48 families diagnosed with polyposis identified a homozygous germline nonsense mutation in *NTHL1* in seven affected individuals from three unrelated families.[273] For carriers of two *NTHL1* mutations, the panel recommends similar management strategies as described for carriers of *AXIN2* mutations.

POLD1 and POLE Mutations

DNA polymerases delta [δ]1 (*POLD1*) and epsilon [ε] (*POLE*) are involved in DNA proofreading and replication.[274] Mutations in the *POLD1* and *POLE* genes may be associated with polyposis and increased risk for CRC.[275-279] Using whole-genome sequencing in combination with linkage and association analysis, heterozygous *POLD1* and *POLE* germline variants were identified in multiple adenoma and/or CRC cases.[277] In an analysis of 858 Spanish patients with early-onset and/or familial CRC and/or colonic polyposis, one patient was found to have a *POLE* mutation.[278] In an analysis of 266 unrelated probands with polyposis or who met the Amsterdam criteria, a *POLE* mutation was found in 1.5% of patients.[280] In one study, *POLD1* mutation carriers were also found to have breast and endometrial tumors.[275] Presently, for carriers of *POLD1* and *POLE* mutations, the panel recommends similar management strategies as described for carriers of *AXIN2* mutations.

Emerging Data on Other Mutations

Mutations in the protein-coding gene *GALNT12* are also believed to be associated with increased risk for CRC.[281-283] Heterozygous mutations in the *ATM* gene,[284] and heterozygous mutations in the DNA *RECQL*-helicase gene *BLM*[285-287] may also increase risk for CRC. There are emerging data that *RPS20* mutations may be associated with increased risk for CRC, but more data are required to strengthen this association.[288] Overall, as data regarding the clinical significance of genes associated with CRC risk emerge, the panel expects that these surveillance recommendations will evolve.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019;69:7-34. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30620402.

2. Cheng L, Eng C, Nieman LZ, et al. Trends in colorectal cancer incidence by anatomic site and disease stage in the United States from 1976 to 2005. Am J Clin Oncol 2011;34:573-580. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21217399.

3. Henley SJ, Singh SD, King J, et al. Invasive cancer incidence and survival--United States, 2011. MMWR Morb Mortal Wkly Rep 2015;64:237-242. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25763875.

4. Siegel R, Ward E, Brawley O, Jemal A. Cancer statistics, 2011: the impact of eliminating socioeconomic and racial disparities on premature cancer deaths. CA Cancer J Clin 2011;61:212-236. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21685461.

5. Bailey CE, Hu CY, You YN, et al. Increasing disparities in the age-related incidences of colon and rectal cancers in the United States, 1975-2010. JAMA Surg 2014:1-6. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25372703.

6. Burt R, Neklason DW. Genetic testing for inherited colon cancer. Gastroenterology 2005;128:1696-1716. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15887160.

7. Giardiello FM, Offerhaus JG. Phenotype and cancer risk of various polyposis syndromes. Eur J Cancer 1995;31A:1085-1087. Available at: http://www.ncbi.nlm.nih.gov/pubmed/7576997.

8. Hamilton SR, Liu B, Parsons RE, et al. The molecular basis of Turcot's syndrome. N Engl J Med 1995;332:839-847. Available at: http://www.ncbi.nlm.nih.gov/pubmed/7661930.

9. U.S. National Library of Medicine-Key MEDLINE® Indicators. Available at: http://www.nlm.nih.gov/bsd/bsd_key.html. Accessed July 24, 2014.

10. Slavin TP, Van Tongeren LR, Behrendt CE, et al. Prospective Study of Cancer Genetic Variants: Variation in Rate of Reclassification by Ancestry. J Natl Cancer Inst 2018;110:1059-1066. Available at: https://www.ncbi.nlm.nih.gov/pubmed/29618041.

11. Kastrinos F, Uno H, Ukaegbu C, et al. Development and Validation of the PREMM5 Model for Comprehensive Risk Assessment of Lynch Syndrome. J Clin Oncol 2017;35:2165-2172. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28489507.

12. Aaltonen LA, Salovaara R, Kristo P, et al. Incidence of hereditary nonpolyposis colorectal cancer and the feasibility of molecular screening for the disease. N Engl J Med 1998;338:1481-1487. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9593786.

13. Hampel H, Frankel WL, Martin E, et al. Screening for the Lynch syndrome (hereditary nonpolyposis colorectal cancer). N Engl J Med 2005;352:1851-1860. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15872200.

14. Hampel H, Frankel WL, Martin E, et al. Feasibility of screening for Lynch syndrome among patients with colorectal cancer. J Clin Oncol 2008;26:5783-5788. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18809606.

15. Lynch HT, de la Chapelle A. Hereditary colorectal cancer. N Engl J Med 2003;348:919-932. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12621137.

16. Boland CR, Goel A. Microsatellite instability in colorectal cancer. Gastroenterology 2010;138:2073-2087 e2073. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20420947.

17. Kempers MJ, Kuiper RP, Ockeloen CW, et al. Risk of colorectal and endometrial cancers in EPCAM deletion-positive Lynch syndrome: a cohort study. Lancet Oncol 2011;12:49-55. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21145788.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

18. Rumilla K, Schowalter KV, Lindor NM, et al. Frequency of deletions of EPCAM (TACSTD1) in MSH2-associated Lynch syndrome cases. J Mol Diagn 2011;13:93-99. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21227399.

19. Vasen HF, Watson P, Mecklin JP, Lynch HT. New clinical criteria for hereditary nonpolyposis colorectal cancer (HNPCC, Lynch syndrome) proposed by the International Collaborative group on HNPCC. Gastroenterology 1999;116:1453-1456. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10348829.

20. Vasen HF. Clinical diagnosis and management of hereditary colorectal cancer syndromes. J Clin Oncol 2000;18:81S-92S. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11060333.

21. Barnetson RA, Tenesa A, Farrington SM, et al. Identification and survival of carriers of mutations in DNA mismatch-repair genes in colon cancer. N Engl J Med 2006;354:2751-2763. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16807412.

22. Rodriguez-Bigas MA, Boland CR, Hamilton SR, et al. A National Cancer Institute Workshop on Hereditary Nonpolyposis Colorectal Cancer Syndrome: meeting highlights and Bethesda guidelines. J Natl Cancer Inst 1997;89:1758-1762. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9392616.

23. Umar A, Boland CR, Terdiman JP, et al. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. J Natl Cancer Inst 2004;96:261-268. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14970275.

24. Raedle J, Trojan J, Brieger A, et al. Bethesda guidelines: relation to microsatellite instability and MLH1 promoter methylation in patients with colorectal cancer. Ann Intern Med 2001;135:566-576. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11601928.

25. Pinol V, Castells A, Andreu M, et al. Accuracy of revised Bethesda guidelines, microsatellite instability, and immunohistochemistry for the identification of patients with hereditary nonpolyposis colorectal cancer. JAMA 2005;293:1986-1994. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15855432.

26. Balmana J, Stockwell DH, Steyerberg EW, et al. Prediction of MLH1 and MSH2 mutations in Lynch syndrome. JAMA 2006;296:1469-1478. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17003395.

27. Chen S, Wang W, Lee S, et al. Prediction of germline mutations and cancer risk in the Lynch syndrome. JAMA 2006;296:1479-1487. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17003396.

28. Kastrinos F, Steyerberg EW, Mercado R, et al. The PREMM(1,2,6) model predicts risk of MLH1, MSH2, and MSH6 germline mutations based on cancer history. Gastroenterology 2011;140:73-81. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20727894.

29. Goverde A, Spaander MCW, Nieboer D, et al. Evaluation of current prediction models for Lynch syndrome: updating the PREMM5 model to identify PMS2 mutation carriers. Fam Cancer 2018;17:361-370. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28933000.

30. Moreira L, Balaguer F, Lindor N, et al. Identification of Lynch syndrome among patients with colorectal cancer. JAMA 2012;308:1555-1565. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23073952.

31. Recommendations from the EGAPP Working Group: genetic testing strategies in newly diagnosed individuals with colorectal cancer aimed at reducing morbidity and mortality from Lynch syndrome in relatives. Genet Med 2009;11:35-41. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19125126.

32. Ladabaum U, Wang G, Terdiman J, et al. Strategies to identify the Lynch syndrome among patients with colorectal cancer: a cost-effectiveness analysis. Ann Intern Med 2011;155:69-79. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21768580.

33. Palomaki GE, McClain MR, Melillo S, et al. EGAPP supplementary evidence review: DNA testing strategies aimed at reducing morbidity and

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

mortality from Lynch syndrome. Genet Med 2009;11:42-65. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19125127.

34. Balmana J, Balaguer F, Cervantes A, Arnold D. Familial risk-colorectal cancer: ESMO Clinical Practice Guidelines. Ann Oncol 2013;24 Suppl 6:vi73-80. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23813931.

35. Giardiello FM, Allen JI, Axilbund JE, et al. Guidelines on genetic evaluation and management of Lynch syndrome: a consensus statement by the US Multi-Society Task Force on colorectal cancer. Gastroenterology 2014;147:502-526. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25043945.

36. Mvundura M, Grosse SD, Hampel H, Palomaki GE. The cost-effectiveness of genetic testing strategies for Lynch syndrome among newly diagnosed patients with colorectal cancer. Genet Med 2010;12:93-104. Available at: https://www.ncbi.nlm.nih.gov/pubmed/20084010.

37. Perez-Carbonell L, Ruiz-Ponte C, Guarinos C, et al. Comparison between universal molecular screening for Lynch syndrome and revised Bethesda guidelines in a large population-based cohort of patients with colorectal cancer. Gut 2012;61:865-872. Available at: https://www.ncbi.nlm.nih.gov/pubmed/21868491.

38. Marquez E, Geng Z, Pass S, et al. Implementation of routine screening for Lynch syndrome in university and safety-net health system settings: successes and challenges. Genet Med 2013;15:925-932. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23598716.

39. Hendriks YM, de Jong AE, Morreau H, et al. Diagnostic approach and management of Lynch syndrome (hereditary nonpolyposis colorectal carcinoma): a guide for clinicians. CA Cancer J Clin 2006;56:213-225. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16870997.

40. Boland CR, Thibodeau SN, Hamilton SR, et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. Cancer Res 1998;58:5248-5257. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9823339.

41. Xicola RM, Llor X, Pons E, et al. Performance of different microsatellite marker panels for detection of mismatch repair-deficient colorectal tumors. J Natl Cancer Inst 2007;99:244-252. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17284719.

42. Caldes T, Godino J, Sanchez A, et al. Immunohistochemistry and microsatellite instability testing for selecting MLH1, MSH2 and MSH6 mutation carriers in hereditary non-polyposis colorectal cancer. Oncol Rep 2004;12:621-629. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15289847.

43. Vasen HF, Hendriks Y, de Jong AE, et al. Identification of HNPCC by molecular analysis of colorectal and endometrial tumors. Dis Markers 2004;20:207-213. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15528786.

44. Hampel H, Frankel W, Panescu J, et al. Screening for Lynch syndrome (hereditary nonpolyposis colorectal cancer) among endometrial cancer patients. Cancer Res 2006;66:7810-7817. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16885385.

45. Lindor NM, Burgart LJ, Leontovich O, et al. Immunohistochemistry versus microsatellite instability testing in phenotyping colorectal tumors. J Clin Oncol 2002;20:1043-1048. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11844828.

46. Reyes CM, Allen BA, Terdiman JP, Wilson LS. Comparison of selection strategies for genetic testing of patients with hereditary nonpolyposis colorectal carcinoma: effectiveness and cost-effectiveness. Cancer 2002;95:1848-1856. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12404277.

47. Shia J, Klimstra DS, Nafa K, et al. Value of immunohistochemical detection of DNA mismatch repair proteins in predicting germline mutation in hereditary colorectal neoplasms. Am J Surg Pathol 2005;29:96-104. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15613860.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

48. Pino MS, Chung DC. Application of molecular diagnostics for the detection of Lynch syndrome. Expert Rev Mol Diagn 2010;10:651-665. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20629513.

49. Christakis AG, Papke DJ, Nowak JA, et al. Targeted Cancer Next Generation Sequencing as a Primary Screening Tool for Microsatellite Instability and Lynch Syndrome in Upper Gastrointestinal Tract Cancers. Cancer Epidemiol Biomarkers Prev 2019. Available at: https://www.ncbi.nlm.nih.gov/pubmed/31028081.

50. Latham A, Srinivasan P, Kemel Y, et al. Microsatellite Instability Is Associated With the Presence of Lynch Syndrome Pan-Cancer. J Clin Oncol 2019;37:286-295. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30376427.

51. Soares BL, Brant AC, Gomes R, et al. Screening for germline mutations in mismatch repair genes in patients with Lynch syndrome by next generation sequencing. Fam Cancer 2018;17:387-394. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28932927.

52. Pearlman R, Frankel WL, Swanson B, et al. Prevalence and Spectrum of Germline Cancer Susceptibility Gene Mutations Among Patients With Early-Onset Colorectal Cancer. JAMA Oncol 2017;3:464-471. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27978560.

53. Yurgelun MB, Kulke MH, Fuchs CS, et al. Cancer Susceptibility Gene Mutations in Individuals With Colorectal Cancer. J Clin Oncol 2017;35:1086-1095. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28135145.

54. Stoffel EM, Mangu PB, Gruber SB, et al. Hereditary colorectal cancer syndromes: American Society of Clinical Oncology Clinical Practice Guideline endorsement of the familial risk-colorectal cancer: European Society for Medical Oncology Clinical Practice Guidelines. J Clin Oncol 2015;33:209-217. Available at: https://www.ncbi.nlm.nih.gov/pubmed/25452455.

55. Rubenstein JH, Enns R, Heidelbaugh J, Barkun A. American Gastroenterological Association Institute Guideline on the Diagnosis and Management of Lynch Syndrome. Gastroenterology 2015;149:777-782. Available at: http://www.ncbi.nlm.nih.gov/pubmed/26226577.

56. Haraldsdottir S, Hampel H, Tomsic J, et al. Colon and endometrial cancers with mismatch repair deficiency can arise from somatic, rather than germline, mutations. Gastroenterology 2014;147:1308-1316.e1301. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25194673.

57. Baglietto L, Lindor NM, Dowty JG, et al. Risks of Lynch syndrome cancers for MSH6 mutation carriers. J Natl Cancer Inst 2010;102:193-201. Available at: https://www.ncbi.nlm.nih.gov/pubmed/20028993.

58. Bonadona V, Bonaiti B, Olschwang S, et al. Cancer risks associated with germline mutations in MLH1, MSH2, and MSH6 genes in Lynch syndrome. JAMA 2011;305:2304-2310. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21642682.

59. Moller P, Seppala TT, Bernstein I, et al. Cancer risk and survival in path_MMR carriers by gene and gender up to 75 years of age: a report from the Prospective Lynch Syndrome Database. Gut 2018;67:1306-1316. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28754778.

60. Ryan NAJ, Morris J, Green K, et al. Association of Mismatch Repair Mutation With Age at Cancer Onset in Lynch Syndrome: Implications for Stratified Surveillance Strategies. JAMA Oncol 2017. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28772289.

61. Senter L, Clendenning M, Sotamaa K, et al. The clinical phenotype of Lynch syndrome due to germ-line PMS2 mutations. Gastroenterology 2008;135:419-428. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18602922.

62. Ten Broeke SW, van der Klift HM, Tops CMJ, et al. Cancer Risks for PMS2-Associated Lynch Syndrome. J Clin Oncol 2018;36:2961-2968. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30161022.

63. Win AK, Lindor NM, Young JP, et al. Risks of primary extracolonic cancers following colorectal cancer in lynch syndrome. J Natl Cancer Inst

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

2012;104:1363-1372. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22933731.

64. Lindor NM, Petersen GM, Hadley DW, et al. Recommendations for the care of individuals with an inherited predisposition to Lynch syndrome: a systematic review. JAMA 2006;296:1507-1517. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17003399.

65. Syngal S, Brand RE, Church JM, et al. ACG clinical guideline: Genetic testing and management of hereditary gastrointestinal cancer syndromes. Am J Gastroenterol 2015;110:223-262; quiz 263. Available at: https://www.ncbi.nlm.nih.gov/pubmed/25645574.

66. Vasen HF, Abdirahman M, Brohet R, et al. One to 2-year surveillance intervals reduce risk of colorectal cancer in families with Lynch syndrome. Gastroenterology 2010;138:2300-2306. Available at: https://www.ncbi.nlm.nih.gov/pubmed/20206180.

67. Moller P, Seppala T, Bernstein I, et al. Cancer incidence and survival in Lynch syndrome patients receiving colonoscopic and gynaecological surveillance: first report from the prospective Lynch syndrome database. Gut 2017;66:464-472. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26657901.

68. Hendriks YM, Wagner A, Morreau H, et al. Cancer risk in hereditary nonpolyposis colorectal cancer due to MSH6 mutations: impact on counseling and surveillance. Gastroenterology 2004;127:17-25. Available at: https://www.ncbi.nlm.nih.gov/pubmed/15236168.

69. Jenkins MA, Dowty JG, Ait Ouakrim D, et al. Short-term risk of colorectal cancer in individuals with lynch syndrome: a meta-analysis. J Clin Oncol 2015;33:326-331. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25534380.

70. Haanstra JF, Kleibeuker JH, Koornstra JJ. Role of new endoscopic techniques in Lynch syndrome. Fam Cancer 2013;12:267-272. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23420551.

71. Stoffel EM, Turgeon DK, Stockwell DH, et al. Missed adenomas during colonoscopic surveillance in individuals with Lynch Syndrome (hereditary nonpolyposis colorectal cancer). Cancer Prev Res (Phila) 2008;1:470-475. Available at: https://www.ncbi.nlm.nih.gov/pubmed/19138994.

72. Kohlmann W, Gruber S. Lynch Syndrome. GeneReviews at GeneTests: Medical Genetics Information Resource 2014. Available at: http://www.ncbi.nlm.nih.gov/books/NBK1211/.

73. Watson P, Vasen HF, Mecklin JP, et al. The risk of extra-colonic, extra-endometrial cancer in the Lynch syndrome. Int J Cancer 2008;123:444-449. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18398828.

74. ACOG Practice Bulletin No. 147: Lynch syndrome. Obstet Gynecol 2014;124:1042-1054. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25437740.

75. Auranen A, Joutsiniemi T. A systematic review of gynecological cancer surveillance in women belonging to hereditary nonpolyposis colorectal cancer (Lynch syndrome) families. Acta Obstet Gynecol Scand 2011;90:437-444. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21306348.

76. Chen LM, Yang KY, Little SE, et al. Gynecologic cancer prevention in Lynch syndrome/hereditary nonpolyposis colorectal cancer families. Obstet Gynecol 2007;110:18-25. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17601891.

77. Jarvinen HJ, Renkonen-Sinisalo L, Aktan-Collan K, et al. Ten years after mutation testing for Lynch syndrome: cancer incidence and outcome in mutation-positive and mutation-negative family members. J Clin Oncol 2009;27:4793-4797. Available at: https://www.ncbi.nlm.nih.gov/pubmed/19720893.

78. Renkonen-Sinisalo L, Butzow R, Leminen A, et al. Surveillance for endometrial cancer in hereditary nonpolyposis colorectal cancer syndrome. Int J Cancer 2007;120:821-824. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17096354.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

79. Rijcken FE, Mourits MJ, Kleibeuker JH, et al. Gynecologic screening in hereditary nonpolyposis colorectal cancer. Gynecol Oncol 2003;91:74-80. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14529665.

80. Dove-Edwin I, Boks D, Goff S, et al. The outcome of endometrial carcinoma surveillance by ultrasound scan in women at risk of hereditary nonpolyposis colorectal carcinoma and familial colorectal carcinoma. Cancer 2002;94:1708-1712. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11920532.

81. Schmeler KM, Lynch HT, Chen LM, et al. Prophylactic surgery to reduce the risk of gynecologic cancers in the Lynch syndrome. N Engl J Med 2006;354:261-269. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16421367.

82. Stuckless S, Green J, Dawson L, et al. Impact of gynecological screening in Lynch syndrome carriers with an MSH2 mutation. Clin Genet 2013;83:359-364. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22775459.

83. Dashti SG, Chau R, Ouakrim DA, et al. Female Hormonal Factors and the Risk of Endometrial Cancer in Lynch Syndrome. Jama 2015;314:61-71. Available at: http://www.ncbi.nlm.nih.gov/pubmed/26151267.

84. Capelle LG, Van Grieken NC, Lingsma HF, et al. Risk and epidemiological time trends of gastric cancer in Lynch syndrome carriers in the Netherlands. Gastroenterology 2010;138:487-492. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19900449.

85. Schulmann K, Engel C, Propping P, Schmiegel W. Small bowel cancer risk in Lynch syndrome. Gut 2008;57:1629-1630. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18941010.

86. ten Kate GL, Kleibeuker JH, Nagengast FM, et al. Is surveillance of the small bowel indicated for Lynch syndrome families? Gut 2007;56:1198-1201. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17409122.

87. Koornstra JJ, Kleibeuker JH, Vasen HF. Small-bowel cancer in Lynch syndrome: is it time for surveillance? Lancet Oncol 2008;9:901-905. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18760246.

88. Kastrinos F, Mukherjee B, Tayob N, et al. Risk of pancreatic cancer in families with Lynch syndrome. JAMA 2009;302:1790-1795. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19861671.

89. Renkonen-Sinisalo L, Sipponen P, Aarnio M, et al. No support for endoscopic surveillance for gastric cancer in hereditary non-polyposis colorectal cancer. Scand J Gastroenterol 2002;37:574-577. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12059060.

90. Kim J, Braun D, Ukaegbu C, et al. Clinical Factors Associated With Gastric Cancer in Individuals With Lynch Syndrome. Clin Gastroenterol Hepatol 2019. Available at: https://www.ncbi.nlm.nih.gov/pubmed/31319185.

91. Vasen HF, Blanco I, Aktan-Collan K, et al. Revised guidelines for the clinical management of Lynch syndrome (HNPCC): recommendations by a group of European experts. Gut 2013;62:812-823. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23408351.

92. Correa P, Haenszel W, Cuello C, et al. A model for gastric cancer epidemiology. Lancet 1975;2:58-60. Available at: https://www.ncbi.nlm.nih.gov/pubmed/49653.

93. Blaser MJ. Hypothesis: the changing relationships of Helicobacter pylori and humans: implications for health and disease. J Infect Dis 1999;179:1523-1530. Available at: https://www.ncbi.nlm.nih.gov/pubmed/10228075.

94. Engel C, Loeffler M, Steinke V, et al. Risks of less common cancers in proven mutation carriers with lynch syndrome. J Clin Oncol 2012;30:4409-4415. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23091106.

95. Joost P, Therkildsen C, Dominguez-Valentin M, et al. Urinary Tract Cancer in Lynch Syndrome; Increased Risk in Carriers of MSH2

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Mutations. Urology 2015;86:1212-1217. Available at: http://www.ncbi.nlm.nih.gov/pubmed/26385421.

96. Win AK, Young JP, Lindor NM, et al. Colorectal and other cancer risks for carriers and noncarriers from families with a DNA mismatch repair gene mutation: a prospective cohort study. J Clin Oncol 2012;30:958-964. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22331944.

97. Canto MI, Harinck F, Hruban RH, et al. International Cancer of the Pancreas Screening (CAPS) Consortium summit on the management of patients with increased risk for familial pancreatic cancer. Gut 2013;62:339-347. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23135763.

98. Win AK, Lindor NM, Jenkins MA. Risk of breast cancer in Lynch syndrome: a systematic review. Breast Cancer Res 2013;15:R27. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23510156.

99. Haraldsdottir S, Hampel H, Wei L, et al. Prostate cancer incidence in males with Lynch syndrome. Genet Med 2014;16:553-557. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24434690.

100. Wimmer K, Kratz CP, Vasen HF, et al. Diagnostic criteria for constitutional mismatch repair deficiency syndrome: suggestions of the European consortium 'care for CMMRD' (C4CMMRD). J Med Genet 2014;51:355-365. Available at: http://jmg.bmj.com/content/51/6/355.full.pdf.

101. Burn J, Gerdes AM, Macrae F, et al. Long-term effect of aspirin on cancer risk in carriers of hereditary colorectal cancer: an analysis from the CAPP2 randomised controlled trial. Lancet 2011;378:2081-2087. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22036019.

102. Movahedi M, Bishop DT, Macrae F, et al. Obesity, Aspirin, and Risk of Colorectal Cancer in Carriers of Hereditary Colorectal Cancer: A Prospective Investigation in the CAPP2 Study. J Clin Oncol 2015;33:3591-3597. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26282643.

103. Cleland JG. Does aspirin really reduce the risk of colon cancer? Lancet 2012;379:1586; author reply 1587. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22541575.

104. Jankowski J, Barr H, Moayyedi P. Does aspirin really reduce the risk of colon cancer? Lancet 2012;379:1586-1587; author reply 1587. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22541573.

105. Ait Ouakrim D, Dashti SG, Chau R, et al. Aspirin, Ibuprofen, and the Risk of Colorectal Cancer in Lynch Syndrome. J Natl Cancer Inst 2015;107. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26109217.

106. Mathers JC, Movahedi M, Macrae F, et al. Long-term effect of resistant starch on cancer risk in carriers of hereditary colorectal cancer: an analysis from the CAPP2 randomised controlled trial. Lancet Oncol 2012;13:1242-1249. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23140761.

107. Stanich PP, Pearlman R, Hinton A, et al. Prevalence of Germline Mutations in Polyposis and Colorectal Cancer-Associated Genes in Patients With Multiple Colorectal Polyps. Clin Gastroenterol Hepatol 2018. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30557735.

108. Aretz S, Koch A, Uhlhaas S, et al. Should children at risk for familial adenomatous polyposis be screened for hepatoblastoma and children with apparently sporadic hepatoblastoma be screened for APC germline mutations? Pediatr Blood Cancer 2006;47:811-818. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16317745.

109. Levy RA, Hui VW, Sood R, et al. Cribriform-morular variant of papillary thyroid carcinoma: an indication to screen for occult FAP. Fam Cancer 2014;13:547-551. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24934245.

110. Ito Y, Miyauchi A, Ishikawa H, et al. Our experience of treatment of cribriform morular variant of papillary thyroid carcinoma; difference in clinicopathological features of FAP-associated and sporadic patients. Endocr J 2011;58:685-689. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21670544.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

111. Grover S, Kastrinos F, Steyerberg EW, et al. Prevalence and phenotypes of APC and MUTYH mutations in patients with multiple colorectal adenomas. JAMA 2012;308:485-492. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22851115.

112. Galiatsatos P, Foulkes WD. Familial adenomatous polyposis. Am J Gastroenterol 2006;101:385-398. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16454848.

113. Half E, Bercovich D, Rozen P. Familial adenomatous polyposis. Orphanet J Rare Dis 2009;4:22. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19822006.

114. Ballhausen WG. Genetic testing for familial adenomatous polyposis. Ann N Y Acad Sci 2000;910:36-47; discussion 47-39. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10911904.

115. Mihalatos M, Apessos A, Papadopoulou E, et al. Genetic alterations of the APC gene in familial adenomatous polyposis patients of the hellenic group for the study of colorectal cancer. Anticancer Res 2003;23:2191-2193. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12894596.

116. Mankaney G, Leone P, Cruise M, et al. Gastric cancer in FAP: a concerning rise in incidence. Fam Cancer 2017;16:371-376. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28185118.

117. Yamaguchi T, Ishida H, Ueno H, et al. Upper gastrointestinal tumours in Japanese familial adenomatous polyposis patients. Jpn J Clin Oncol 2016;46:310-315. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26819281.

118. Nieuwenhuis MH, Vasen HF. Correlations between mutation site in APC and phenotype of familial adenomatous polyposis (FAP): a review of the literature. Crit Rev Oncol Hematol 2007;61:153-161. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17064931.

119. Nusliha A, Dalpatadu U, Amarasinghe B, et al. Congenital hypertrophy of retinal pigment epithelium (CHRPE) in patients with familial adenomatous polyposis (FAP); a polyposis registry experience. BMC Res Notes 2014;7:734. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25326340.

120. Sturt NJ, Gallagher MC, Bassett P, et al. Evidence for genetic predisposition to desmoid tumours in familial adenomatous polyposis independent of the germline APC mutation. Gut 2004;53:1832-1836. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15542524.

121. Kennedy RD, Potter DD, Moir CR, El-Youssef M. The natural history of familial adenomatous polyposis syndrome: a 24 year review of a single center experience in screening, diagnosis, and outcomes. J Pediatr Surg 2014;49:82-86. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24439586.

122. Burt RW, Leppert MF, Slattery ML, et al. Genetic testing and phenotype in a large kindred with attenuated familial adenomatous polyposis. Gastroenterology 2004;127:444-451. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15300576.

123. Knudsen AL, Bulow S, Tomlinson I, et al. Attenuated familial adenomatous polyposis: results from an international collaborative study. Colorectal Dis 2010;12:e243-249. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20105204.

124. Aretz S, Uhlhaas S, Caspari R, et al. Frequency and parental origin of de novo APC mutations in familial adenomatous polyposis. Eur J Hum Genet 2004;12:52-58. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14523376.

125. Bisgaard ML, Fenger K, Bulow S, et al. Familial adenomatous polyposis (FAP): frequency, penetrance, and mutation rate. Hum Mutat 1994;3:121-125. Available at: http://www.ncbi.nlm.nih.gov/pubmed/8199592.

126. Hegde MR, Roa BB. Detecting mutations in the APC gene in familial adenomatous polyposis (FAP). Curr Protoc Hum Genet 2006;Chapter 10:Unit 10 18. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18428386.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

127. Giardiello FM, Brensinger JD, Petersen GM, et al. The use and interpretation of commercial APC gene testing for familial adenomatous polyposis. N Engl J Med 1997;336:823-827. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9062090.

128. Guillem JG, Wood WC, Moley JF, et al. ASCO/SSO review of current role of risk-reducing surgery in common hereditary cancer syndromes. J Clin Oncol 2006;24:4642-4660. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17008706.

129. De Cosse JJ, Bulow S, Neale K, et al. Rectal cancer risk in patients treated for familial adenomatous polyposis. The Leeds Castle Polyposis Group. Br J Surg 1992;79:1372-1375. Available at: http://www.ncbi.nlm.nih.gov/pubmed/1336702.

130. Heiskanen I, Jarvinen HJ. Fate of the rectal stump after colectomy and ileorectal anastomosis for familial adenomatous polyposis. Int J Colorectal Dis 1997;12:9-13. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9112143.

131. Vasen HF, van Duijvendijk P, Buskens E, et al. Decision analysis in the surgical treatment of patients with familial adenomatous polyposis: a Dutch-Scandinavian collaborative study including 659 patients. Gut 2001;49:231-235. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11454800.

132. Bulow S, Bulow C, Vasen H, et al. Colectomy and ileorectal anastomosis is still an option for selected patients with familial adenomatous polyposis. Dis Colon Rectum 2008;51:1318-1323. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18523824.

133. Church J, Burke C, McGannon E, et al. Risk of rectal cancer in patients after colectomy and ileorectal anastomosis for familial adenomatous polyposis: a function of available surgical options. Dis Colon Rectum 2003;46:1175-1181. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12972960.

134. Ambroze WL, Jr., Dozois RR, Pemberton JH, et al. Familial adenomatous polyposis: results following ileal pouch-anal anastomosis and ileorectostomy. Dis Colon Rectum 1992;35:12-15. Available at: http://www.ncbi.nlm.nih.gov/pubmed/1310269.

135. Madden MV, Neale KF, Nicholls RJ, et al. Comparison of morbidity and function after colectomy with ileorectal anastomosis or restorative proctocolectomy for familial adenomatous polyposis. Br J Surg 1991;78:789-792. Available at: http://www.ncbi.nlm.nih.gov/pubmed/1651799.

136. Soravia C, Klein L, Berk T, et al. Comparison of ileal pouch-anal anastomosis and ileorectal anastomosis in patients with familial adenomatous polyposis. Dis Colon Rectum 1999;42:1028-1033; discussion 1033-1024. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10458126.

137. Van Duijvendijk P, Slors JF, Taat CW, et al. Quality of life after total colectomy with ileorectal anastomosis or proctocolectomy and ileal pouch-anal anastomosis for familial adenomatous polyposis. Br J Surg 2000;87:590-596. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10792315.

138. van Duijvendijk P, Slors JF, Taat CW, et al. Functional outcome after colectomy and ileorectal anastomosis compared with proctocolectomy and ileal pouch-anal anastomosis in familial adenomatous polyposis. Ann Surg 1999;230:648-654. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10561088.

139. Ziv Y, Church JM, Oakley JR, et al. Surgery for the teenager with familial adenomatous polyposis: ileo-rectal anastomosis or restorative proctocolectomy? Int J Colorectal Dis 1995;10:6-9. Available at: http://www.ncbi.nlm.nih.gov/pubmed/7745328.

140. Bjork JA, Akerbrant HI, Iselius LE, Hultcrantz RW. Risk factors for rectal cancer morbidity and mortality in patients with familial adenomatous polyposis after colectomy and ileorectal anastomosis. Dis Colon Rectum 2000;43:1719-1725. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11156457.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

141. Nugent KP, Phillips RK. Rectal cancer risk in older patients with familial adenomatous polyposis and an ileorectal anastomosis: a cause for concern. Br J Surg 1992;79:1204-1206. Available at: http://www.ncbi.nlm.nih.gov/pubmed/1334761.

142. Spigelman AD, Williams CB, Talbot IC, et al. Upper gastrointestinal cancer in patients with familial adenomatous polyposis. Lancet 1989;2:783-785. Available at: http://www.ncbi.nlm.nih.gov/pubmed/2571019.

143. Bulow S, Christensen IJ, Hojen H, et al. Duodenal surveillance improves the prognosis after duodenal cancer in familial adenomatous polyposis. Colorectal Dis 2012;14:947-952. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21973191.

144. Kallenberg FGJ, Bastiaansen BAJ, Dekker E. Cap-assisted forward-viewing endoscopy to visualize the ampulla of Vater and the duodenum in patients with familial adenomatous polyposis. Endoscopy 2017;49:181-185. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27760435.

145. Abraham SC, Nobukawa B, Giardiello FM, et al. Fundic gland polyps in familial adenomatous polyposis: neoplasms with frequent somatic adenomatous polyposis coli gene alterations. Am J Pathol 2000;157:747-754. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10980114.

146. Leone PJ, Mankaney G, Sarvapelli S, et al. Endoscopic and histologic features associated with gastric cancer in familial adenomatous polyposis. Gastrointest Endosc 2019;89:961-968. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30597145.

147. Walton SJ, Frayling IM, Clark SK, Latchford A. Gastric tumours in FAP. Fam Cancer 2017;16:363-369. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28271232.

148. Groen EJ, Roos A, Muntinghe FL, et al. Extra-intestinal manifestations of familial adenomatous polyposis. Ann Surg Oncol 2008;15:2439-2450. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18612695.

149. Bulow C, Bulow S. Is screening for thyroid carcinoma indicated in familial adenomatous polyposis? The Leeds Castle Polyposis Group. Int J Colorectal Dis 1997;12:240-242. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9272455.

150. Giardiello FM, Offerhaus GJ, Lee DH, et al. Increased risk of thyroid and pancreatic carcinoma in familial adenomatous polyposis. Gut 1993;34:1394-1396. Available at: http://www.ncbi.nlm.nih.gov/pubmed/8244108.

151. Herraiz M, Barbesino G, Faquin W, et al. Prevalence of thyroid cancer in familial adenomatous polyposis syndrome and the role of screening ultrasound examinations. Clin Gastroenterol Hepatol 2007;5:367-373. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17258512.

152. Steinhagen E, Hui VW, Levy RA, et al. Results of a prospective thyroid ultrasound screening program in adenomatous polyposis patients. Am J Surg 2014;208:764-769. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25073656.

153. Truta B, Allen BA, Conrad PG, et al. Genotype and phenotype of patients with both familial adenomatous polyposis and thyroid carcinoma. Fam Cancer 2003;2:95-99. Available at: http://www.ncbi.nlm.nih.gov/pubmed/14574158.

154. Feng X, Milas M, O'Malley M, et al. Characteristics of benign and malignant thyroid disease in familial adenomatous polyposis patients and recommendations for disease surveillance. Thyroid 2015;25:325-332. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25585202.

155. Church J, Lynch C, Neary P, et al. A desmoid tumor-staging system separates patients with intra-abdominal, familial adenomatous polyposis-associated desmoid disease by behavior and prognosis. Dis Colon Rectum 2008;51:897-901. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18322756.

156. Church J, Berk T, Boman BM, et al. Staging intra-abdominal desmoid tumors in familial adenomatous polyposis: a search for a uniform approach

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

to a troubling disease. Dis Colon Rectum 2005;48:1528-1534. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15906134.

157. Quintini C, Ward G, Shatnawei A, et al. Mortality of intra-abdominal desmoid tumors in patients with familial adenomatous polyposis: a single center review of 154 patients. Ann Surg 2012;255:511-516. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22323009.

158. Attard TM, Giglio P, Koppula S, et al. Brain tumors in individuals with familial adenomatous polyposis: a cancer registry experience and pooled case report analysis. Cancer 2007;109:761-766. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17238184.

159. Algra AM, Rothwell PM. Effects of regular aspirin on long-term cancer incidence and metastasis: a systematic comparison of evidence from observational studies versus randomised trials. Lancet Oncol 2012;13:518-527. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22440112.

160. Baron JA, Cole BF, Sandler RS, et al. A randomized trial of aspirin to prevent colorectal adenomas. N Engl J Med 2003;348:891-899. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12621133.

161. Brasky TM, Potter JD, Kristal AR, et al. Non-steroidal anti-inflammatory drugs and cancer incidence by sex in the VITamins And Lifestyle (VITAL) cohort. Cancer Causes Control 2012;23:431-444. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22212612.

162. Chan AT, Giovannucci EL, Meyerhardt JA, et al. Long-term use of aspirin and nonsteroidal anti-inflammatory drugs and risk of colorectal cancer. JAMA 2005;294:914-923. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16118381.

163. Ruder EH, Laiyemo AO, Graubard BI, et al. Non-steroidal anti-inflammatory drugs and colorectal cancer risk in a large, prospective cohort. Am J Gastroenterol 2011;106:1340-1350. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21407185.

164. Sandler RS, Halabi S, Baron JA, et al. A randomized trial of aspirin to prevent colorectal adenomas in patients with previous colorectal cancer. N Engl J Med 2003;348:883-890. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12621132.

165. Arber N, Eagle CJ, Spicak J, et al. Celecoxib for the prevention of colorectal adenomatous polyps. N Engl J Med 2006;355:885-895. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16943401.

166. Arber N, Spicak J, Racz I, et al. Five-year analysis of the prevention of colorectal sporadic adenomatous polyps trial. Am J Gastroenterol 2011;106:1135-1146. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21503000.

167. Baron JA, Sandler RS, Bresalier RS, et al. A randomized trial of rofecoxib for the chemoprevention of colorectal adenomas. Gastroenterology 2006;131:1674-1682. Available at: http://www.ncbi.nlm.nih.gov/pubmed/17087947.

168. Bertagnolli MM, Eagle CJ, Zauber AG, et al. Celecoxib for the prevention of sporadic colorectal adenomas. N Engl J Med 2006;355:873-884. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16943400.

169. Bertagnolli MM, Eagle CJ, Zauber AG, et al. Five-year efficacy and safety analysis of the Adenoma Prevention with Celecoxib Trial. Cancer Prev Res (Phila Pa) 2009;2:310-321. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19336730.

170. Bresalier RS, Sandler RS, Quan H, et al. Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. N Engl J Med 2005;352:1092-1102. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15713943.

171. Solomon SD, McMurray JJ, Pfeffer MA, et al. Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. N Engl J Med 2005;352:1071-1080. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15713944.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

172. Giardiello FM, Yang VW, Hylind LM, et al. Primary chemoprevention of familial adenomatous polyposis with sulindac. N Engl J Med 2002;346:1054-1059. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11932472.

173. Burn J, Bishop DT, Chapman PD, et al. A randomized placebo-controlled prevention trial of aspirin and/or resistant starch in young people with familial adenomatous polyposis. Cancer Prev Res (Phila) 2011;4:655-665. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21543343.

174. Coffey RJ, Hawkey CJ, Damstrup L, et al. Epidermal growth factor receptor activation induces nuclear targeting of cyclooxygenase-2, basolateral release of prostaglandins, and mitogenesis in polarizing colon cancer cells. Proc Natl Acad Sci U S A 1997;94:657-662. Available at: https://www.ncbi.nlm.nih.gov/pubmed/9012840.

175. Eisinger AL, Nadauld LD, Shelton DN, et al. The adenomatous polyposis coli tumor suppressor gene regulates expression of cyclooxygenase-2 by a mechanism that involves retinoic acid. J Biol Chem 2006;281:20474-20482. Available at: https://www.ncbi.nlm.nih.gov/pubmed/16699180.

176. Roberts RB, Min L, Washington MK, et al. Importance of epidermal growth factor receptor signaling in establishment of adenomas and maintenance of carcinomas during intestinal tumorigenesis. Proc Natl Acad Sci U S A 2002;99:1521-1526. Available at: https://www.ncbi.nlm.nih.gov/pubmed/11818567.

177. Samadder NJ, Neklason DW, Boucher KM, et al. Effect of Sulindac and Erlotinib vs Placebo on Duodenal Neoplasia in Familial Adenomatous Polyposis: A Randomized Clinical Trial. JAMA 2016;315:1266-1275. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27002448.

178. Steinbach G, Lynch PM, Phillips RK, et al. The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. N Engl J Med 2000;342:1946-1952. Available at: http://www.ncbi.nlm.nih.gov/pubmed/10874062.

179. Cruz-Correa M, Hylind LM, Romans KE, et al. Long-term treatment with sulindac in familial adenomatous polyposis: a prospective cohort study. Gastroenterology 2002;122:641-645. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11874996.

180. West NJ, Clark SK, Phillips RK, et al. Eicosapentaenoic acid reduces rectal polyp number and size in familial adenomatous polyposis. Gut 2010;59:918-925. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20348368.

181. Al-Tassan N, Chmiel NH, Maynard J, et al. Inherited variants of MYH associated with somatic G:C-->T:A mutations in colorectal tumors. Nat Genet 2002;30:227-232. Available at: http://www.ncbi.nlm.nih.gov/pubmed/11818965.

182. Jones S, Emmerson P, Maynard J, et al. Biallelic germline mutations in MYH predispose to multiple colorectal adenoma and somatic G:C-->T:A mutations. Hum Mol Genet 2002;11:2961-2967. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12393807.

183. Theodoratou E, Campbell H, Tenesa A, et al. A large-scale meta-analysis to refine colorectal cancer risk estimates associated with MUTYH variants. Br J Cancer 2010;103:1875-1884. Available at: https://www.ncbi.nlm.nih.gov/pubmed/21063410.

184. Vogt S, Jones N, Christian D, et al. Expanded extracolonic tumor spectrum in MUTYH-associated polyposis. Gastroenterology 2009;137:1976-1985 e1971-1910. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19732775.

185. Win AK, Cleary SP, Dowty JG, et al. Cancer risks for monoallelic MUTYH mutation carriers with a family history of colorectal cancer. Int J Cancer 2011;129:2256-2262. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21171015.

186. Win AK, Dowty JG, Cleary SP, et al. Risk of colorectal cancer for carriers of mutations in MUTYH, with and without a family history of cancer. Gastroenterology 2014;146:1208-1211 e1201-1205. Available at: https://www.ncbi.nlm.nih.gov/pubmed/24444654.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

187. Win AK, Hopper JL, Jenkins MA. Association between monoallelic MUTYH mutation and colorectal cancer risk: a meta-regression analysis. Fam Cancer 2011;10:1-9. Available at: https://www.ncbi.nlm.nih.gov/pubmed/21061173.

188. Rosner G, Bercovich D, Daniel YE, et al. Increased risk for colorectal adenomas and cancer in mono-allelic MUTYH mutation carriers: results from a cohort of North-African Jews. Fam Cancer 2015;14:427-436. Available at: https://www.ncbi.nlm.nih.gov/pubmed/25822476.

189. Lubbe SJ, Di Bernardo MC, Chandler IP, Houlston RS. Clinical implications of the colorectal cancer risk associated with MUTYH mutation. J Clin Oncol 2009;27:3975-3980. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19620482.

190. Ma X, Zhang B, Zheng W. Genetic variants associated with colorectal cancer risk: comprehensive research synopsis, meta-analysis, and epidemiological evidence. Gut 2014;63:326-336. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23946381.

191. Katona BW, Yurgelun MB, Garber JE, et al. A counseling framework for moderate-penetrance colorectal cancer susceptibility genes. Genet Med 2018;20:1324-1327. Available at: https://www.ncbi.nlm.nih.gov/pubmed/29493579.

192. Nieuwenhuis MH, Vogt S, Jones N, et al. Evidence for accelerated colorectal adenoma--carcinoma progression in MUTYH-associated polyposis? Gut 2012;61:734-738. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21846783.

193. Tomlinson IP, Houlston RS. Peutz-Jeghers syndrome. J Med Genet 1997;34:1007-1011. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9429144.

194. Shaco-Levy R, Jasperson KW, Martin K, et al. Morphologic characterization of hamartomatous gastrointestinal polyps in Cowden syndrome, Peutz-Jeghers syndrome, and juvenile polyposis syndrome. Hum Pathol 2016;49:39-48. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26826408.

195. Burdick D, Prior JT. Peutz-Jeghers syndrome. A clinicopathologic study of a large family with a 27-year follow-up. Cancer 1982;50:2139-2146. Available at: http://www.ncbi.nlm.nih.gov/pubmed/7127254.

196. Giardiello FM, Welsh SB, Hamilton SR, et al. Increased risk of cancer in the Peutz-Jeghers syndrome. N Engl J Med 1987;316:1511-1514. Available at: http://www.ncbi.nlm.nih.gov/pubmed/3587280.

197. Linos DA, Dozois RR, Dahlin DC, Bartholomew LG. Does Peutz-Jeghers syndrome predispose to gastrointestinal malignancy? A later look. Arch Surg 1981;116:1182-1184. Available at: http://www.ncbi.nlm.nih.gov/pubmed/7283716.

198. Spigelman AD, Murday V, Phillips RK. Cancer and the Peutz-Jeghers syndrome. Gut 1989;30:1588-1590. Available at: http://www.ncbi.nlm.nih.gov/pubmed/2599445.

199. Lim W, Hearle N, Shah B, et al. Further observations on LKB1/STK11 status and cancer risk in Peutz-Jeghers syndrome. Br J Cancer 2003;89:308-313. Available at: https://www.ncbi.nlm.nih.gov/pubmed/12865922.

200. Jenne DE, Reimann H, Nezu J, et al. Peutz-Jeghers syndrome is caused by mutations in a novel serine threonine kinase. Nat Genet 1998;18:38-43. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9425897.

201. Hemminki A, Markie D, Tomlinson I, et al. A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. Nature 1998;391:184-187. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9428765.

202. Dunlop MG. Guidance on gastrointestinal surveillance for hereditary non-polyposis colorectal cancer, familial adenomatous polypolis, juvenile polyposis, and Peutz-Jeghers syndrome. Gut 2002;51 Suppl 5:V21-27. Available at: http://www.ncbi.nlm.nih.gov/pubmed/12221036.

203. Latchford AR, Neale K, Phillips RK, Clark SK. Juvenile polyposis syndrome: a study of genotype, phenotype, and long-term outcome. Dis

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Colon Rectum 2012;55:1038-1043. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22965402.

204. Coburn MC, Pricolo VE, DeLuca FG, Bland KI. Malignant potential in intestinal juvenile polyposis syndromes. Ann Surg Oncol 1995;2:386-391. Available at: http://www.ncbi.nlm.nih.gov/pubmed/7496832.

205. Cichy W, Klincewicz B, Plawski A. Juvenile polyposis syndrome. Arch Med Sci 2014;10:570-577. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25097590.

206. Iyer NK, Burke CA, Leach BH, Parambil JG. SMAD4 mutation and the combined syndrome of juvenile polyposis syndrome and hereditary haemorrhagic telangiectasia. Thorax 2010;65:745-746. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20685751.

207. Howe JR, Mitros FA, Summers RW. The risk of gastrointestinal carcinoma in familial juvenile polyposis. Ann Surg Oncol 1998;5:751-756. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9869523.

208. Brosens LA, Langeveld D, van Hattem WA, et al. Juvenile polyposis syndrome. World J Gastroenterol 2011;17:4839-4844. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22171123.

209. Jass JR, Williams CB, Bussey HJ, Morson BC. Juvenile polyposis--a precancerous condition. Histopathology 1988;13:619-630. Available at: http://www.ncbi.nlm.nih.gov/pubmed/2853131.

210. Snover DC, Ahnen DJ, Burt RW, Odze RD. Serrated Polyps of the Colon and Rectum and Serrated Polyposis. In: Bosman FT, Carneiro, F., Hruban, R. H., Theise, N.D., ed. WHO Classification of Tumours of the Digestive System. Lyon: IARC; 2010:160-165.

211. Noffsinger AE, Hart J. Serrated adenoma: a distinct form of non-polypoid colorectal neoplasia? Gastrointest Endosc Clin N Am 2010;20:543-563. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20656251.

212. Rex DK, Ahnen DJ, Baron JA, et al. Serrated lesions of the colorectum: review and recommendations from an expert panel. Am J Gastroenterol 2012;107:1315-1329; quiz 1314, 1330. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22710576.

213. De Sousa EMF, Wang X, Jansen M, et al. Poor-prognosis colon cancer is defined by a molecularly distinct subtype and develops from serrated precursor lesions. Nat Med 2013;19:614-618. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23584090.

214. Guarinos C, Sanchez-Fortun C, Rodriguez-Soler M, et al. Serrated polyposis syndrome: molecular, pathological and clinical aspects. World J Gastroenterol 2012;18:2452-2461. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22654442.

215. Rosty C, Walsh MD, Walters RJ, et al. Multiplicity and molecular heterogeneity of colorectal carcinomas in individuals with serrated polyposis. Am J Surg Pathol 2013;37:434-442. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23211288.

216. Rosty C, Brosens LA, Dekker E, Nagtegaal ID. Serrated Polyposis. In: Lokuhetty D, White VA, Watanabe R, Cree IA, eds. WHO Classification of Tumours: Digestive System Tumours. Lyon, France: International Agency for Research on Cancer; 2019:532-534.

217. Egoavil C, Juarez M, Guarinos C, et al. Increased Risk of Colorectal Cancer in Patients With Multiple Serrated Polyps and Their First-Degree Relatives. Gastroenterology 2017;153:106-112 e102. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28400194.

218. Boparai KS, Mathus-Vliegen EM, Koornstra JJ, et al. Increased colorectal cancer risk during follow-up in patients with hyperplastic polyposis syndrome: a multicentre cohort study. Gut 2010;59:1094-1100. Available at: http://www.ncbi.nlm.nih.gov/pubmed/19710031.

219. Yeoman A, Young J, Arnold J, et al. Hyperplastic polyposis in the New Zealand population: a condition associated with increased colorectal cancer risk and European ancestry. N Z Med J 2007;120:U2827. Available at: http://www.ncbi.nlm.nih.gov/pubmed/18264196.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN®), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

220. Jasperson KW, Kanth P, Kirchhoff AC, et al. Serrated polyposis: colonic phenotype, extracolonic features, and familial risk in a large cohort. Dis Colon Rectum 2013;56:1211-1216. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24104994.

221. Edelstein DL, Cruz-Correa M, Soto-Salgado M, et al. Risk of Colorectal and Other Cancers in Patients With Serrated Polyposis. Clin Gastroenterol Hepatol 2015;13:1697-1699. Available at: https://www.ncbi.nlm.nih.gov/pubmed/25681317.

222. Gala MK, Mizukami Y, Le LP, et al. Germline mutations in oncogene-induced senescence pathways are associated with multiple sessile serrated adenomas. Gastroenterology 2014;146:520-529. Available at: https://www.ncbi.nlm.nih.gov/pubmed/24512911.

223. Taupin D, Lam W, Rangiah D, et al. A deleterious RNF43 germline mutation in a severely affected serrated polyposis kindred. Hum Genome Var 2015;2:15013. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27081527.

224. Valle L. Recent Discoveries in the Genetics of Familial Colorectal Cancer and Polyposis. Clin Gastroenterol Hepatol 2017;15:809-819. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27712984.

225. Yan HHN, Lai JCW, Ho SL, et al. RNF43 germline and somatic mutation in serrated neoplasia pathway and its association with BRAF mutation. Gut 2017;66:1645-1656. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27329244.

226. Boparai KS, Reitsma JB, Lemmens V, et al. Increased colorectal cancer risk in first-degree relatives of patients with hyperplastic polyposis syndrome. Gut 2010;59:1222-1225. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20584785.

227. Lage P, Cravo M, Sousa R, et al. Management of Portuguese patients with hyperplastic polyposis and screening of at-risk first-degree relatives: a contribution for future guidelines based on a clinical study. Am J Gastroenterol 2004;99:1779-1784. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15330918.

228. Win AK, Walters RJ, Buchanan DD, et al. Cancer risks for relatives of patients with serrated polyposis. Am J Gastroenterol 2012;107:770-778. Available at: http://www.ncbi.nlm.nih.gov/pubmed/22525305.

229. Oquinena S, Guerra A, Pueyo A, et al. Serrated polyposis: prospective study of first-degree relatives. Eur J Gastroenterol Hepatol 2013;25:28-32. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23011040.

230. Kallenberg FGJ, Latchford A, Lips NC, et al. Duodenal Adenomas in Patients With Multiple Colorectal Adenomas Without Germline APC or MUTYH Mutations. Dis Colon Rectum 2018;61:58-66. Available at: https://www.ncbi.nlm.nih.gov/pubmed/29215473.

231. Tieu AH, Edelstein D, Axilbund J, et al. Clinical Characteristics of Multiple Colorectal Adenoma Patients Without Germline APC or MYH Mutations. J Clin Gastroenterol 2016;50:584-588. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26485104.

232. Hall MJ, Forman AD, Pilarski R, et al. Gene panel testing for inherited cancer risk. J Natl Compr Canc Netw 2014;12:1339-1346. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25190699.

233. Gallego CJ, Shirts BH, Bennette CS, et al. Next-Generation Sequencing Panels for the Diagnosis of Colorectal Cancer and Polyposis Syndromes: A Cost-Effectiveness Analysis. J Clin Oncol 2015;33:2084-2091. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25940718.

234. Walsh T, Casadei S, Coats KH, et al. Spectrum of mutations in BRCA1, BRCA2, CHEK2, and TP53 in families at high risk of breast cancer. JAMA 2006;295:1379-1388. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16551709.

235. Walsh T, Lee MK, Casadei S, et al. Detection of inherited mutations for breast and ovarian cancer using genomic capture and massively parallel sequencing. Proc Natl Acad Sci U S A 2010;107:12629-12633. Available at: http://www.ncbi.nlm.nih.gov/pubmed/20616022.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

236. Bombard Y, Bach PB, Offit K. Translating genomics in cancer care. J Natl Compr Canc Netw 2013;11:1343-1353. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24225968.

237. Rainville IR, Rana HQ. Next-generation sequencing for inherited breast cancer risk: counseling through the complexity. Curr Oncol Rep 2014;16:371. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24488544.

238. Cragun D, Radford C, Dolinsky JS, et al. Panel-based testing for inherited colorectal cancer: a descriptive study of clinical testing performed by a US laboratory. Clin Genet 2014;86:510-520. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24506336.

239. LaDuca H, Stuenkel AJ, Dolinsky JS, et al. Utilization of multigene panels in hereditary cancer predisposition testing: analysis of more than 2,000 patients. Genet Med 2014;16:830-837. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24763289.

240. Mauer CB, Pirzadeh-Miller SM, Robinson LD, Euhus DM. The integration of next-generation sequencing panels in the clinical cancer genetics practice: an institutional experience. Genet Med 2014;16:407-412. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24113346.

241. Kapoor NS, Curcio LD, Blakemore CA, et al. Multigene panel testing detects equal rates of pathogenic BRCA1/2 mutations and has a higher diagnostic yield compared to limited BRCA1/2 analysis alone in patients at risk for hereditary breast cancer. Ann Surg Oncol 2015;22:3282-3288. Available at: http://www.ncbi.nlm.nih.gov/pubmed/26219241.

242. Kurian AW, Hare EE, Mills MA, et al. Clinical evaluation of a multiple-gene sequencing panel for hereditary cancer risk assessment. J Clin Oncol 2014;32:2001-2009. Available at: https://www.ncbi.nlm.nih.gov/pubmed/24733792.

243. Tung N, Battelli C, Allen B, et al. Frequency of mutations in individuals with breast cancer referred for BRCA1 and BRCA2 testing using next-generation sequencing with a 25-gene panel. Cancer 2015;121:25-33. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25186627.

244. Yurgelun MB, Allen B, Kaldate RR, et al. Identification of a Variety of Mutations in Cancer Predisposition Genes in Patients With Suspected Lynch Syndrome. Gastroenterology 2015;149:604-613.e620. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25980754.

245. Mersch J, Brown N, Pirzadeh-Miller S, et al. Prevalence of Variant Reclassification Following Hereditary Cancer Genetic Testing. JAMA 2018;320:1266-1274. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30264118.

246. David KL, Best RG, Brenman LM, et al. Patient re-contact after revision of genomic test results: points to consider-a statement of the American College of Medical Genetics and Genomics (ACMG). Genet Med 2019;21:769-771. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30578420.

247. Slavin TP, Manjarrez S, Pritchard CC, et al. The effects of genomic germline variant reclassification on clinical cancer care. Oncotarget 2019;10:417-423. Available at: https://www.ncbi.nlm.nih.gov/pubmed/30728895.

248. Robson ME, Bradbury AR, Arun B, et al. American Society of Clinical Oncology Policy statement update: genetic and genomic testing for cancer susceptibility. J Clin Oncol 2015;33:3660-3667. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26324357.

249. Markowitz SD, Bertagnolli MM. Molecular origins of cancer: Molecular basis of colorectal cancer. N Engl J Med 2009;361:2449-2460. Available at: https://www.ncbi.nlm.nih.gov/pubmed/20018966.

250. Boursi B, Sella T, Liberman E, et al. The APC p.I1307K polymorphism is a significant risk factor for CRC in average risk Ashkenazi Jews. Eur J Cancer 2013;49:3680-3685. Available at: http://www.ncbi.nlm.nih.gov/pubmed/23896379.

251. Gryfe R, Di Nicola N, Lal G, et al. Inherited colorectal polyposis and cancer risk of the APC I1307K polymorphism. Am J Hum Genet 1999;64:378-384. Available at: http://www.ncbi.nlm.nih.gov/pubmed/9973276.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

252. Laken SJ, Petersen GM, Gruber SB, et al. Familial colorectal cancer in Ashkenazim due to a hypermutable tract in APC. Nat Genet 1997;17:79-83. Available at: https://www.ncbi.nlm.nih.gov/pubmed/9288102.

253. Liang J, Lin C, Hu F, et al. APC polymorphisms and the risk of colorectal neoplasia: a HuGE review and meta-analysis. Am J Epidemiol 2013;177:1169-1179. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23576677.

254. Locker GY, Kaul K, Weinberg DS, et al. The I1307K APC polymorphism in Ashkenazi Jews with colorectal cancer: clinical and pathologic features. Cancer Genet Cytogenet 2006;169:33-38. Available at: http://www.ncbi.nlm.nih.gov/pubmed/16875934.

255. Patael Y, Figer A, Gershoni-Baruch R, et al. Common origin of the I1307K APC polymorphism in Ashkenazi and non-Ashkenazi Jews. Eur J Hum Genet 1999;7:555-559. Available at: https://www.ncbi.nlm.nih.gov/pubmed/10439961.

256. Rennert G, Almog R, Tomsho LP, et al. Colorectal polyps in carriers of the APC I1307K polymorphism. Dis Colon Rectum 2005;48:2317-2321. Available at: https://www.ncbi.nlm.nih.gov/pubmed/16228836.

257. Shtoyerman-Chen R, Friedman E, Figer A, et al. The I1307K APC polymorphism: prevalence in non-Ashkenazi Jews and evidence for a founder effect. Genet Test 2001;5:141-146. Available at: https://www.ncbi.nlm.nih.gov/pubmed/11551102.

258. Lammi L, Arte S, Somer M, et al. Mutations in AXIN2 cause familial tooth agenesis and predispose to colorectal cancer. Am J Hum Genet 2004;74:1043-1050. Available at: http://www.ncbi.nlm.nih.gov/pubmed/15042511.

259. Lejeune S, Guillemot F, Triboulet JP, et al. Low frequency of AXIN2 mutations and high frequency of MUTYH mutations in patients with multiple polyposis. Hum Mutat 2006;27:1064. Available at: https://www.ncbi.nlm.nih.gov/pubmed/16941501.

260. Marvin ML, Mazzoni SM, Herron CM, et al. AXIN2-associated autosomal dominant ectodermal dysplasia and neoplastic syndrome. Am J Med Genet A 2011;155a:898-902. Available at: http://www.ncbi.nlm.nih.gov/pubmed/21416598.

261. Rivera B, Perea J, Sanchez E, et al. A novel AXIN2 germline variant associated with attenuated FAP without signs of oligondontia or ectodermal dysplasia. Eur J Hum Genet 2014;22:423-426. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23838596.

262. Wong S, Liu H, Bai B, et al. Novel missense mutations in the AXIN2 gene associated with non-syndromic oligodontia. Arch Oral Biol 2014;59:349-353. Available at: https://www.ncbi.nlm.nih.gov/pubmed/24581859.

263. Gronwald J, Cybulski C, Piesiak W, et al. Cancer risks in first-degree relatives of CHEK2 mutation carriers: effects of mutation type and cancer site in proband. Br J Cancer 2009;100:1508-1512. Available at: https://www.ncbi.nlm.nih.gov/pubmed/19401704.

264. Han FF, Guo CL, Liu LH. The effect of CHEK2 variant I157T on cancer susceptibility: evidence from a meta-analysis. DNA Cell Biol 2013;32:329-335. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23713947.

265. Liu C, Wang QS, Wang YJ. The CHEK2 I157T variant and colorectal cancer susceptibility: a systematic review and meta-analysis. Asian Pac J Cancer Prev 2012;13:2051-2055. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22901170.

266. Xiang HP, Geng XP, Ge WW, Li H. Meta-analysis of CHEK2 1100delC variant and colorectal cancer susceptibility. Eur J Cancer 2011;47:2546-2551. Available at: https://www.ncbi.nlm.nih.gov/pubmed/21807500.

267. Jaeger E, Leedham S, Lewis A, et al. Hereditary mixed polyposis syndrome is caused by a 40-kb upstream duplication that leads to increased and ectopic expression of the BMP antagonist GREM1. Nat

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

Genet 2012;44:699-703. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22561515.

268. Lieberman S, Walsh T, Schechter M, et al. Features of Patients With Hereditary Mixed Polyposis Syndrome Caused by Duplication of GREM1 and Implications for Screening and Surveillance. Gastroenterology 2017;152:1876-1880 e1871. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28242209.

269. Rohlin A, Eiengard F, Lundstam U, et al. GREM1 and POLE variants in hereditary colorectal cancer syndromes. Genes Chromosomes Cancer 2016;55:95-106. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26493165.

270. Ikeda M, Orimo H, Moriyama H, et al. Close correlation between mutations of E2F4 and hMSH3 genes in colorectal cancers with microsatellite instability. Cancer Res 1998;58:594-598. Available at: https://www.ncbi.nlm.nih.gov/pubmed/9485005.

271. Adam R, Spier I, Zhao B, et al. Exome Sequencing Identifies Biallelic MSH3 Germline Mutations as a Recessive Subtype of Colorectal Adenomatous Polyposis. Am J Hum Genet 2016;99:337-351. Available at: https://www.ncbi.nlm.nih.gov/pubmed/27476653.

272. Krokan HE, Bjoras M. Base excision repair. Cold Spring Harb Perspect Biol 2013;5:a012583. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23545420.

273. Weren RD, Ligtenberg MJ, Kets CM, et al. A germline homozygous mutation in the base-excision repair gene NTHL1 causes adenomatous polyposis and colorectal cancer. Nat Genet 2015;47:668-671. Available at: http://www.ncbi.nlm.nih.gov/pubmed/25938944.

274. Garg P, Burgers PM. DNA polymerases that propagate the eukaryotic DNA replication fork. Crit Rev Biochem Mol Biol 2005;40:115-128. Available at: https://www.ncbi.nlm.nih.gov/pubmed/15814431.

275. Bellido F, Pineda M, Aiza G, et al. POLE and POLD1 mutations in 529 kindred with familial colorectal cancer and/or polyposis: review of reported cases and recommendations for genetic testing and surveillance. Genet Med 2016;18:325-332. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26133394.

276. Esteban-Jurado C, Gimenez-Zaragoza D, Munoz J, et al. POLE and POLD1 screening in 155 patients with multiple polyps and early-onset colorectal cancer. Oncotarget 2017;8:26732-26743. Available at: https://www.ncbi.nlm.nih.gov/pubmed/28423643.

277. Palles C, Cazier JB, Howarth KM, et al. Germline mutations affecting the proofreading domains of POLE and POLD1 predispose to colorectal adenomas and carcinomas. Nat Genet 2013;45:136-144. Available at: https://www.ncbi.nlm.nih.gov/pubmed/23263490.

278. Valle L, Hernandez-Illan E, Bellido F, et al. New insights into POLE and POLD1 germline mutations in familial colorectal cancer and polyposis. Hum Mol Genet 2014;23:3506-3512. Available at: https://www.ncbi.nlm.nih.gov/pubmed/24501277.

279. Elsayed FA, Kets CM, Ruano D, et al. Germline variants in POLE are associated with early onset mismatch repair deficient colorectal cancer. Eur J Hum Genet 2015;23:1080-1084. Available at: https://www.ncbi.nlm.nih.gov/pubmed/25370038.

280. Spier I, Holzapfel S, Altmuller J, et al. Frequency and phenotypic spectrum of germline mutations in POLE and seven other polymerase genes in 266 patients with colorectal adenomas and carcinomas. Int J Cancer 2015;137:320-331. Available at: https://www.ncbi.nlm.nih.gov/pubmed/25529843.

281. Guda K, Moinova H, He J, et al. Inactivating germ-line and somatic mutations in polypeptide N-acetylgalactosaminyltransferase 12 in human colon cancers. Proc Natl Acad Sci U S A 2009;106:12921-12925. Available at: https://www.ncbi.nlm.nih.gov/pubmed/19617566.

282. Clarke E, Green RC, Green JS, et al. Inherited deleterious variants in GALNT12 are associated with CRC susceptibility. Hum Mutat 2012;33:1056-1058. Available at: https://www.ncbi.nlm.nih.gov/pubmed/22461326.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.

Printed by Arena OncoPath on 12/13/2019 1:33:20 PM. For personal use only. Not approved for distribution. Copyright © 2019 National Comprehensive Cancer Network, Inc., All Rights Reserved.

283. Segui N, Pineda M, Navarro M, et al. GALNT12 is not a major contributor of familial colorectal cancer type X. Hum Mutat 2014;35:50-52. Available at: https://www.ncbi.nlm.nih.gov/pubmed/24115450.

284. Thompson D, Duedal S, Kirner J, et al. Cancer risks and mortality in heterozygous ATM mutation carriers. J Natl Cancer Inst 2005;97:813-822. Available at: https://www.ncbi.nlm.nih.gov/pubmed/15928302.

285. Baris HN, Kedar I, Halpern GJ, et al. Prevalence of breast and colorectal cancer in Ashkenazi Jewish carriers of Fanconi anemia and Bloom syndrome. Isr Med Assoc J 2007;9:847-850. Available at: https://www.ncbi.nlm.nih.gov/pubmed/18210922.

286. Cleary SP, Zhang W, Di Nicola N, et al. Heterozygosity for the BLM(Ash) mutation and cancer risk. Cancer Res 2003;63:1769-1771. Available at: https://www.ncbi.nlm.nih.gov/pubmed/12702560.

287. Laitman Y, Boker-Keinan L, Berkenstadt M, et al. The risk for developing cancer in Israeli ATM, BLM, and FANCC heterozygous mutation carriers. Cancer Genet 2016;209:70-74. Available at: https://www.ncbi.nlm.nih.gov/pubmed/26778106.

288. Nieminen TT, O'Donohue MF, Wu Y, et al. Germline mutation of RPS20, encoding a ribosomal protein, causes predisposition to hereditary nonpolyposis colorectal carcinoma without DNA mismatch repair deficiency. Gastroenterology 2014;147:595-598.e595. Available at: http://www.ncbi.nlm.nih.gov/pubmed/24941021.

Version 3.2019, 12/13/19 © 2019 National Comprehensive Cancer Network® (NCCN©), All rights reserved. NCCN Guidelines® and this illustration may not be reproduced in any form without the express written permission of NCCN.